**Quantifying leaf trait covariation and its controls** **across climates and biomes**

Yanzheng Yang1,2,3,\*, Han Wang1,3, Sandy P. Harrison3,4, I. Colin Prentice1,3,5,6 , Ian J. Wright6, Changhui Peng3,7,\* and Guanghui Lin1,8,\*

1Ministry of Education Key Laboratory for Earth System Modeling, Department of Earth System Science, Tsinghua University, Beijing 100084, China.

2Joint Center for Global Change Studies (JCGCS), Beijing 100875, China

3State Key Laboratory of Soil Erosion and Dryland Farming on the Loess Plateau, College of Forestry, Northwest A&F University, Yangling, Shaanxi 712100, China

4School of Archaeology, Geography and Environmental Sciences (SAGES), University of Reading, Reading, RG6 6AH, UK

5AXA Chair of Biosphere and Climate Impacts, Imperial College London, Department of Life Sciences, Silwood Park Campus, Buckhurst Road, Ascot SL5 7PY, UK

6Department of Biological Sciences, Macquarie University, North Ryde, NSW 2109, Australia

7Department of Biological Sciences, Institute of Environmental Sciences, University of Quebec at Montreal, C.P. 8888, Succ. Centre-Ville, Montréal H3C 3P8, QC, Canada

8Key Laboratory of Stable Isotope and Gulf Ecology, Graduate School at Shenzhen, Tsinghua University, Shenzhen, Guangdong 518055, China

(\*Authors for correspondence: tel +86(10)62797230; email [yanzheng148@163.com](mailto:peng.changhui@uqam.ca) (Y.Y.); tel +86(10)62797230; email [lingh@tsinghua.edu.cn](mailto:lingh@tsinghua.edu.cn) (G.L.); tel +86(29)87080608; email cpeng86@yahoo.com (C.P.))

**Summary**

* Plant functional ecology requires the quantification of trait variation and its controls. Field measurements on 483 species at 48 sites across China were used to analyse variation in leaf traits, and assess their predictability.
* Principal components analysis (PCA) was used to characterize trait variation, redundancy analysis (RDA) to reveal climate effects, and RDA with variance partitioning to estimate separate and overlapping effects of site, climate, life-form and family membership.
* Four orthogonal dimensions of total trait variation were identified: leaf area (LA), internal-to-ambient CO2 ratio (*χ*), leaf economics spectrum traits (specific leaf area (SLA) *versus* leaf dry matter content (LDMC) and nitrogen per area (*N*area)), and photosynthetic capacities (*V*cmax, *J*max at 25˚C). LA and *χ* covaried with moisture index. Site, climate, life form and family together explained 70% of trait variance. Families accounted for 17%, and climate and families together 29%. LDMC and SLA showed the largest family effects. Independent life-form effects were small.
* Climate influences trait variation in part by selection for different life forms and families. Trait values derived from climate data via RDA showed substantial predictive power for trait values in the available global data sets. Systematic trait data collection across all climates and biomes is still necessary.

**Key words:** climate, leaf economics spectrum, multivariate analysis, photosynthetic capacity, phylogeny, plant functional traits, vegetation modelling.

**Introduction**

Functional traits generally do not vary independently, but show broadly predictable patterns of covariation (Armbruster *et al*., 1996; Watson *et al*., 2016). The covariation of traits may mean that traits share genetic controls, or that they have related roles in community assembly and function (Wright *et al*., 2007; Fajardo *et al*., 2011). Quantifying the covariation of vegetative traits and their controls is important for an understanding of how plants drive ecosystem processes and determine the responses of ecosystems to environmental change (Wright et al., 2007; Shipley *et al.*, 2011; Swenson 2013; van Bodegom *et al.*, 2014; Kong et al., 2014; Kraft *et al.*, 2015). Although a number of large-scale studies have quantified both trait covariation (e.g. Wright *et al.*, 2004; Armbruster *et al*., 2014; Peiman & Robinson, 2017) and trait-environment relationships,(e.g. Wright *et al.*, 2005; Harrison et al., 2010; Liu et al., 2012; Maire et al., 2015; Meng et al., 2015), a number of general issues await resolution. These include:

(1) The dimensionality of trait space – that is, the extent to which combinations of different traits are independent, *versus* belonging to a set of covarying traits as exemplified by the leaf economics spectrum (LES) (Wright *et al*., 2004, 2005). The intrinsic dimensionality of traits is the minimum number of independent axes that adequately describe the functional variation among species, and is therefore an important quantity in comparative ecology (Laughlin, 2014).

(2) The extent to which trait variation is determined by climate, versus the co-existence of multiple trait values in the same climate (Adler *et al*., 2013; Valladares *et al*., 2015).

(3) The extent to which trait variation and trait-environment correlations are linked to ‘hard-wired’ physiognomic (life-form) and/or phylogenetic differences among species, and the role of environment in selecting among life forms and clades (Díaz *et al*., 2013; Ackerly, 2009; Donovan *et al*., 2014).

The dimensionality question has received attention in plant functional ecology partly because of the universal nature of the LES, which is considered as the outcome of a tradeoff between resource acquisition and conservation – representing different general strategies for existence, rather than adaptations to environment (Wright *et al.*, 2007; Kong *et al.*, 2014; Reich, 2014). An early synthesis led to a proposal for four trait dimensions indexed by leaf mass per area and lifespan (i.e. the LES), seed mass and seed output, leaf and twig size, and plant height (Westoby *et al.*, 2002). Wright *et al*. (2007) found three independent trait dimensions represented by specific leaf area (SLA), seed/fruit size and leaf size in seven neotropical forests. The most extensive study (in terms of the number of species considered) to date was by Díaz *et al.* (2016), who showed that variation among species in height, stem specific density, leaf mass per area, seed mass, and nitrogen per unit mass (*N*mass) could be reduced to two dimensions, the first indexing plant size, the second the LES. However, these various studies have considered only a limited set of traits or combined information from disparate sources, and did not attempt to quantify the climatic or phylogenetic controls on traits.

In this paper, we examine a suite of leaf traits, using co-located measurements to quantify the contributions of climate, site, life form and phylogeny to trait variation at a large geographic scale. Our analysis is based on an extensive data set (Wang *et al*., 2018), containing information on multiple leaf traits from different regions of China. We focused on seven leaf traits that together capture many functions of plants (Table S1). The traits considered include four commonly measured traits: leaf area (LA), specific leaf area (SLA), leaf dry matter content (LDMC) and leaf nitrogen per unit area (*N*area), and also three traits that determine photosynthetic rates: maximum carboxylation rate (*V*cmax) and maximum electron transport rate (*J*max), derived from gas exchange measurements in the field, and the ratio of intercellular to ambient carbon dioxide (CO2) concentration (often denoted as *c*i:*c*a but called *χ* here following Prentice *et al.*, 2014) derived from leaf stable carbon isotope (δ13C) measurements. We used multivariate analysis to quantify the dimensionality of variation in this set of traits, and the nature and dimensionality of trait-climate relationships. We used variance partitioning to attribute trait variations (for all traits, and each trait separately) to differences among sites, climate variations across sites, and distinctions among life forms and plant families. We finally applied the trait-climate relationships derived from the data set to various global datasets for specific traits, in order to assess their generality and potential wider application.

**Materials and methods**

**Dataset description**

The data are derived from the China Plant Trait Database (Wang *et al.*, 2018), which contains information on morphological, physical, chemical and photosynthetic traits from 122 sites and provides information on more than 1215 species. The database was designed to provide comprehensive sampling of different vegetation types and climates. It employs a standardized taxonomy and includes information on life form, plant family, site location, elevation, and climate. LA, SLA, *N*area, LDMC and leaf δ13C data from multiple species were available at 48 sites, including 483 species altogether, distributed through the eastern half of China (Fig. 1a, Table S2). The sites from northeastern China are distributed along an aridity gradient (Prentice et al., 2011), including steppes, grasslands and temperate deciduous broadleaf forests. The sites from southwestern China represent tropical and subtropical evergreen broadleaf forests, and tropical dry woodlands. Temperate deciduous forests in central China and boreal forests in the far north of China were also included. Collectively these data cover the principal climatic and vegetation zones of the region (Fig. 1b). At each site, a stratified sampling strategy ensured that measurements were available for the main species in each canopy stratum, including up to 25 species of trees. Species were classified by life form as trees, small trees, lianas, shrubs, forbs and graminoids. Bamboos, herbaceous climbers, geophytes and pteridophytes were present only in small numbers in the dataset and were not included in our analysis. Fig. S1 shows frequency distributions of each trait within each life form for forest and non-forest sites. Table S3 lists the total number of samples in each class.

Details of trait measurement methods can be found in Wang *et al.* (2018). LA, SLA, *N*area and LDMC were measured on samples collected in the field following standard protocols (Cornelissen *et al.*, 2003). LA was taken as the projected area of a leaf, or leaflet in the case of compound leaves.*V*cmax was calculated from the light-saturated rate of net CO2 fixation at ambient CO2 (*A*sat) using the so-called one-point method, which provides a rapid and effective alternative to the measurement of a full *A*-*c*i curve (De Kauwe *et al.*, 2016). *J*max was calculated from the light-saturated rate of net CO2 fixation at high CO2 (*A*max). Both *V*cmax and *J*max were adjusted to a standard temperature of 25oC using the methods proposed by Niinemets *et al.* (2014). The adjusted values are called *V*cmax25 and *J*max25. Leaf 13C measurements were converted to 13C discrimination and thence to *χ,* eliminating the effects of latitude and sampling year as described in Cornwell *et al.* (2017):

(1)

where *φ* is latitude and *a*, *b* and *c* are parameters estimated by regression with values *a* = 0.0819, *b* = 0.0983 and *c* = 7.7521 (Cornwell *et al.*, 2017), and

(2)

where is the sampling year and *g* = –0.0467, and

(3)

where *a'* is the discrimination against 13CO2 during diffusion through stomata (4.4‰) and *b'* is the discrimination against 13CO2 duringcarboxylation (27‰) (Farquhar *et al.*, 1982). Cernusak *et al.* (2013) showed that about 80% of the variation in instantaneous gas exchange measurements of χ could be accounted for by a linear relationship to δ13C, supporting the use of equation (3). Estimates of *χ* based on δ13C measurements are used here, however, because they reflect longer-term growth conditions better.

Three bioclimate variables adequately represent the controls on vegetation structure and composition across China (Wang *et al.*, 2013). These are the accumulated photosynthetically active radiation during the thermal growing season (PAR0), defined as the period when daily temperature is above 0oC; the daily mean temperature during the thermal growing season (mGDD0); and the ratio of mean annual precipitation to annual equilibrium evapotranspiration (moisture index, MI), calculated using SPLASH (Davis *et al.*, 2017). The primary data for the calculation of these bioclimatic variables were derived from 1814 meteorological stations (740 stations with data from 1971 to 2000, the rest from 1981 to 1990), interpolated to 1 km resolution with elevation as a covariate using ANUSPLIN V4.37 (Hutchinson 2007).

**Gap filling**

Photosynthetic measurements were only available for 14 sites in the China Plant Trait Database; however, these sites comprise 53% of the species represented in the data set. Photosynthetic measurements were not available for the temperate forests of Changbai Mountain, and the Inner Mongolia grasslands. In order to allow multivariate analysis of a larger data set, *V*cmax values for species at these sites were gap-filled using a back-propagation neural network using LMA, *N*area, LA, *χ* and moisture index (MI) as predictors (newff function in Matlab 2010a). The neural network is a machine learning technique that often provides better performance than conventional statistical methods for this type of application (Paruelo *et al*., 1997; Papale *et al*., 2003; Moffat *et al*., 2010). The data were divided into two parts: a calibration data set used to determine the weights in the neural network (75% of data points), and a validation data set used to assess the network performance (25% of data points). The method achieved an acceptable accuracy with *R*2 = 0.49 between observed and predicted values for the calibration data set and 0.50 for the validation data set. *J*max values were then estimated from *V*cmax values using a linear regression fitted to data from all sites where both *A*sat and *A*max were measured. The regression equation used for gap-filling is loge *Jmax,25* = –0.0221 mGDD0 + 0.7329 loge *Vcmax,25* + 2.0362 (*R*2 = 0.75, *P* < 0.01).

**Multivariate analysis and variance partitioning**

*Principal components analysis* (PCA) and *redundancy analysis* (RDA) are powerful multivariate analysis techniques with many ecological applications (White *et al*., 2005; Maire *et al*., 2015; Scheibe *et al*., 2015). As a dimensionality reduction technique, PCA projects a set of data on correlated variables on to a series of composite, uncorrelated variables called principal components (James *et al*., 1990). In RDA, these variables are chosen to maximize the extent of their correlation with a set of predictor variables (Borcard *et al*., 1992) and are therefore described as “constrained” axes of variation. RDA also extracts further “unconstrained” axes, which are the principal components of the variation that remains after the fitted effects of the predictor variables have been removed. Here, PCA is used to analyse trait covariation; RDA is used to analyse the relationships of trait variation to climate variables; and the unconstrained axes of RDA are used to characterize the residual (within-site) variation in traits. These analyses were performed using the vegan package in R (Oksanen *et al.*, 2017). LA was square-root transformed before analysis to yield a linear measure of leaf size. *χ* was logit-transformed (logit *χ* = loge [*χ*/(1 – *χ*)]). All other traits (including √LA) were natural log-transformed. All traits were thus converted to dimensionless quantities in the range (, ), allowing PCA and RDA to be carried out using the covariance matrix among traits with no need for further standardization. Each trait thereby has its ‘natural’ weight in the analysis. For log-transformed variables, this treatment implies that a trait with, say, 10-fold variation has twice the weight of a trait with 5-fold variation. The weight can be quantified by the standard deviation of the transformed variables (loge √LA: 1.17, loge SLA: 0.50, loge LDMC: 0.38, loge *N*area: 0.59, loge *V*cmax25: 0.58, loge *J*max25: 0.48, logit *χ*: 1.37; see also Table 3). PCA and RDA were repeated using only the species-site combinations for which actual (as opposed to gap-filled) photosynthetic trait data were available (Figs S2-S4, Tables S4-S5).

*Variation partitioning* quantifies the amount of variation in a predicted quantity (in multiple regression) or set of quantities (in RDA) that can be explained by different groups of predictors (Legendre & Legendre, 2012). We used the Legendre method (Legendre & Anderson, 1999; Peres-Neto *et al*., 2006; Meng *et al*., 2015), which explicitly accounts for correlations between groups by distinguishing unique and overlapping contributions from each group. The results are most conveniently displayed as Venn diagrams. The method was used here with RDA to assign trait variation to components linked to climate, sites, life forms, families, and the intersections of these controls.

**Trait prediction**

We evaluated the predictive power of the fitted trait-climate relationships in the RDA analysis, first on the data set as a whole and then using a cross-validation approach (Picard & Cook, 1984; Kohavi 1995). We performed five iterations, in which 80% of the data was used for training and 20% retained for validation. The average root-mean-squared error (RMSE) across all five trials provides the final measure of goodness-of-fit.

The general predictive power of the trait-climate relationships was then tested using four independent global trait data sets: leaf economics traits (SLA, LDMC, *N*area) from Wright *et al.* (2004); √LA from Wright *et al.* (2017); photosynthetic traits (*V*cmax25 , *J*max25) from De Kauwe *et al.* (2016), including data from Bahar et al. (2017); and *χ* from Cornwell *et al.* (2017) (Table S6). Each of these data sets provides geolocated site-based measurements across continents, vegetation types and climates (Figure S5). We derived climate variables for each site from the nearest 10-minute grid cell in the CRU 2.0 dataset (New *et al*. 2002), which provides long-term monthly means of temperature, precipitation, and sunshine duration for the standard period 1961-1990. PAR0, mGDD0, and MI were calculated in the same way as for the sites in China, using SPLASH to calculate MI (Davis *et al.*, 2017).

We screened out measurements from sites in the global data sets where MI > 1.4 or mGDD0 < 10 because these are beyond the limits of the climates sampled in China. Some of the δ13C measurements in Cornwell *et al.* (2017) are < –30‰. We assume that these reflect incomplete mixing of CO2 between the free atmosphere and the forest understorey. We excluded these measurements. The number of sites and individual measurements from each global data set used to test the climate-trait predictions is shown in Table S6. Trait values at each global site were directly predicted from climate inputs, using the RDA model previously derived from the data in China. Ordinary least-squares regression was used to compare observed (*y*) with predicted (*x*) trait values.

**Results**

**Four dimensions of trait variation**

PCA of traits from all species and sampling sites revealed four independent axes of trait variation (Fig. 2, Table 1). The first four principal components together account for 95% of total trait variation. The first two axes are dominated by LA and *χ*, orthogonal to one another. These two axes together account for 79% of total trait variation: this large fraction draws attention to the large span of variability in these traits, especially leaf area. The third axis, accounting for 11% of total trait variation, primarily represents the LES, with SLA opposed to *N*area and LDMC. The plot of axis 3 against axis 4, which accounts for 6% of total trait variation, shows that *V*cmax and *J*max vary closely together, but orthogonally to the LES.

Analysis based on sites with complete data only (Fig. S2, Table S4) shows that the first four principal components have similar explanatory power to the main analysis (93%) and, although the axes are rotated with respect to the axes derived from the larger data set, they show the same four dimensions of variation with LA, LES, photosynthetic capacity and χ varying independently of one another. The patterns of trait covariation can also be seen by examining the matrix of pairwise correlations between traits (Fig. S6). The differences between Fig. S6 (a) based on the gap-filled data set, and Fig. S6(b) based on sites with complete data, show the (slight) effect of gap-filling. *V*cmax and *J*max are highly correlated (0.84) before gap filling. The largest difference is that the negative correlations of both *V*cmax and *J*max with leaf area *increase* due to the gap filling. This evidently does not contradict our inference from PCA on the gap-filled data set, i.e. that photosynthetic capacities are largely uncorrelated with the other traits.

**Trait variation related to climate**

The three bioclimatic variables together account for 37% of trait variation (Table 2). Three successive RDA axes (Fig. 3, Table 2) describe the patterns of trait variation with climate, and show that the between-site patterns of trait covariation imposed by climatic gradients differ from those found in the data set as a whole. The first RDA axis is overwhelmingly dominant, and is related to the gradient of MI from desert-steppe to moist forests. LA and χ vary together along this gradient, with both large leaves and large *χ* characteristic of wetter environments. The second RDA axis accounts for 2% of trait variation, and is related to the covariation of mean growing-season temperature and total growing-season light availability along the latitudinal gradient from the boreal zone to the tropics. Trait variation on this axis resembles the LES: warmer, higher irradiance climates are characterized by plants with lower SLA, higher LDMC and higher *N*area. The third RDA axis accounts for only 0.4% of trait variation. Analysis based on sites with complete data only (Fig. S3, Table S5) shows the same patterns.

**Residual trait variation, unrelated to climate**

The unconstrained axes (or residual principal components) calculated by RDA after climatic differences among sites have been accounted for (Fig. 4, Table 2) provide insight into trait variation that is expressed within sites and across all climates. The patterns of this residual variation, as shown by the first four unconstrained axes, are similar to the patterns shown by the principal components of the whole data set (Fig. 2, Table 1), with evidence for four independent dimensions of variation associated with successive components dominated by *χ*, LA, LES traits and photosynthetic capacities, respectively. Analysis based on sites with complete data only (Fig. S4, Table S5) shows the same four dimensions.

The same general patterns of non-climate-related trait covariation are also clear on inspection of the partial correlations among transformed trait values, after the effects of climatic predictors have been removed (Fig. 5). Deeper colours in Fig. 5 indicate larger absolute magnitudes of correlation. The traits can be seen to fall into four blocks: one comprising *V*cmax and *J*max (positively correlated), one comprising the traits that contribute to the LES (SLA negatively correlated with LDMC and *N*area), *χ*, and LA. While *χ* shows almost no correlation with any of the other traits, LA is weakly negatively correlated with *V*cmax and *J*max (Fig. 5), as is SLA.

**Multiple controls of trait variation**

Venn diagrams (Fig. 6) summarize the percentage contributions of climate, site, life form and family (including intersecting contributions) to total trait variation, and to variation in each separate trait. The intersection regions represent trait variation that cannot be unambiguously attributed to one control or another, because of correlations among the controls. For example, substantial intersections between climate and family occur because these controls are not independent: different families are selected for in different climates. Anomalously large values (relative to the all-traits analysis) are highlighted in bold in Fig. 6. Climate is shown nested within site, because differences in climate are determined by site location. Table 3 also shows the total percentage of variance associated with each control (including intersections with other controls).

Considering the variation among all traits together (Fig. 6), climate, site, family and life form jointly account for 70% of total trait variance. The most important features of the partitioning are (1) the joint effect of climate with family independent of life form (17%); (2) the substantial fraction of variance due to family alone (17%), independent of site or life form; and (3) the fact that most of the total variance associated with life form (16%) is also linked to climate (13%). There is some additional effect of climate independent of family (8%); and some effect of site independent of climate (12%), which is presumably related to edaphic or microclimatic factors.

The partitioning of trait variance for individual traits (Fig. 6) generally resembles that for all traits. However, 48% of total trait variation in LDMC is linked to family, and 41% linked to family independent of other controls. Only 4% of the variation in LDMC is linked to climate, and none to climate and family together. For SLA, 40% of total trait variation is linked to family (with 14% linked to family and life form together independent of other controls); 9% is linked to family and site together independent of other controls, but only 4% to climate and family together. These anomalies indicate a particularly strong phylogenetic component to variation in LDMC and, to a lesser extent, SLA. The unexplained variation is greater for *V*cmax25 (46%) and *J*max25 (41%) than for the other traits.

After climate, site and family effects have been accounted for, the remaining (independent) contribution of life form to trait variation is small. The total life-form contribution is < 10% for all traits except LA and *χ*, and the unique contribution of life form independent of all other controls is very slight, < 2.5% for all traits. Forbs and graminoids show different ranges of trait values in forest and non-forest vegetation (Fig. S1). Specifically, SLA and LDMC of forbs and graminoids decrease between forests and non-forests while *N*area, *V*cmax and *J*max increase. That is, for all these traits, life forms occupying the understorey in forest vegetation become more ‘tree-like’ in non-forest vegetation, suggesting that these traits are more determined by the light environment than by any intrinsic difference among life forms.

**Worldwide prediction of traits based on the observed climate-trait relationships**

The RDA analyses show that climate (including indirect effects mediated by selection for life forms and families) is the major determinant of trait variation for most of the traits examined, except for LDMC and SLA, which show a substantial independent phylogenetic component. This generalization is supported by predictions of the mean site values for each trait (Fig S7). At species level, the adjusted *R*2 between observed and predicted values for LDMC is only 0.08, and for SLA 0.16 (Table S7), while the relationship is better for other traits – from 0.24 for *V*cmax25 to 0.52 for √LA. The average adjusted *R*2 across traits is 0.28. Partitioning the data into woody and non-woody components has little impact on the quality of the prediction for most traits, but prediction of LDMC and SLA is better for non-woody than woody species (Table S7). Although predictability is imperfect, because of the (demonstrated) influence of non-climatic factors on all of the traits, these analyses nonetheless show that it is possible to predict all four dimensions of trait variation, to first order, from climate.

The prediction of trait values in global data sets provides a more stringent test of the universality of the derived climate-trait relationships (Fig. 7, Table 4). At site level, the lowest adjusted *R*2 value between observed and predicted trait values is again for LDMC (0.01), but for SLA it is 0.31. For other traits, adjusted *R*2 ranged from 0.25 (*J*max) to 0.34 (√LA). The average across traits is 0.31, excluding LDMC. The observed values for loge *V*cmax25 tend to be higher than the predicted values, whereas the observed values of loge SLA tend to be lower than the predicted values (Fig. 7). However the regression slopes for these traits are not significantly different from unity (Table 4). The OLS regression slopes for loge √LA, *J*max25 andloge χ are in the range from 0.48 to 1. RMSE values (Table 4) are larger in the global comparison than in the calibration set for loge √LA and SLA; but closely similar for *N*area, *V*cmax25 and *J*max25, and *χ*. The average RMSE across traits excluding LDMC is slightly less in the global comparison (0.42) than in the calibration set (0.61).

**Discussion**

**The ecological significance of leaf-trait dimensions**

The four dimensions of total leaf-trait variation reported here indicate the existence of independent variation among species in LA, *χ*, photosynthetic capacity, and the LES. The RDA based on climate shows a smaller dimensionality, with most of the variation concentrated on a single axis from wet to dry environments. LA is both expected and observed to increase with plant-available moisture, due to energy-balance constraints (Wright *et al.*, 2017). *χ* is both expected and observed to increase with atmospheric moisture according to the least-cost hypothesis (Prentice *et al.*, 2014). These hydroclimatic controls on both LA and *χ* are presumed to be the cause of (a) the dominance of a single dimension of trait-environment relationships across the region, related to moisture/aridity, and (b) the observed close covariation of LA and *χ* between sites along the aridity gradient– contrasting with their independence in the data as a whole. Analysis of the residual (non-climatic) component of trait variation however shows, once again, four independent dimensions, with a pattern closely similar to that shown in total leaf-trait variation, and orthogonal variation of LA and *χ*.

Multivariate analysis confirms the universal nature of the LES, as indexed here by SLA, LDMC (which tends to be high when SLA is low), and *N*area. Unlike *N*mass (N concentration per unit mass), *N*area increases with *decreasing* SLA because the structural component of leaf N increases in proportion to LMA (see e.g. Onoda *et al.*, 2004, 2017; Wright *et al.*, 2005; Osnas *et al.*, 2013; Dong *et al.*, 2017a). The LES is identified in the PCA, and in the residual trait variation after consideration of climate effects in RDA. However, it also appears in the climatically constrained RDA as a second-order pattern correlated with the latitudinal gradient. In other words, there is a shift in the average position of species along the LES (towards lower SLA) with increasing growing-season length and warmth, although this shift accounts only for a small proportion (2%) of total trait variance. The LES reflects the inescapable linkage between high construction costs and long payback times of leaves with low SLA (Kikuzawa, 1991; Reich *et al*., 1997; McMurtrie & Dewar, 2011; Funk & Cornwell, 2013). The shift towards lower-SLA leaves in warmer climates is primarily due to the shift of dominance from deciduous to evergreen woody plants. The increase in growing-season length (towards a year-round growing season in the tropics) favours longer-lived evergreen leaves with lower SLA in warmer climates, as shown here and in other studies.

Both the gap-filled data set and the non-gap-filled subset show that the two photosynthetic capacities (*V*cmax and *J*max) covary closely (Fig. S6), as is expected from the co-ordination hypothesis – which predicts that leaves should not possess excess capacity in either carboxylation or electron transport, as photosynthesis depends on both (Chen *et al.*, 1993; Maire *et al.*, 2012). However both traits show substantial variation within sites. When *V*cmax and *J*max were entered into the analysis after adjustment to local growth temperature, as opposed to 25˚C, the results were very similar (not shown). Opposite trends of variation in *V*cmax and *J*max are shown only in the (minor) third axis of the RDA, accounting for 0.4% of total trait variance and driven by differences among sites in summer temperature that are independent of the latitudinal gradient. This pattern is consistent with expectations, as a decline in the *J*max:*V*cmax ratio with increasing temperature has been shown experimentally (Kattge & Knorr, 2007) and predicted theoretically (Wang *et al*., 2017a). The decline is larger when the two photosynthetic capacities are estimated at prevailing growth temperature, but persists when they are adjusted to 25˚C.

**Contributions to leaf trait variation**

The variance partitioning results presented here demonstrate that family and climate effects (except for LDMC and SLA) overlap considerably. In other words, a substantial part of trait variation with climate is due to families replacing one another along environmental gradients. After family, climate and site effects have been taken into account, independent life-form effects become unimportant. Thus, to first order, the principal controls on trait variation in this data set are family identity, climate, and climatic selection among families. Additional effects of site (independent of climate) could in principle be due to microclimatic and/or edaphic differences among sites, which have not been investigated. LDMC and to a lesser extent SLA show stronger family effects than other traits, while the effects of climate on these traits appear to be largely independent of family identity.

**Implications for vegetation modelling**

Vegetation models based on continuous variation in trait space sample ‘plants’ from a continuum of trait values (e.g. Scheiter *et al*., 2013; Fyllas *et al.*, 2014). This approach requires specifying which traits can vary; by how much; and the extent to which different traits covary, in other words, the effective dimensionality of trait space. Our analyses of leaf traits, including traits derived from stable isotope and gas exchange measurements, indicate that at least four independent dimensions of trait variation need to be considered; that realistic modelling of functional diversity must allow for within-site variation in each of these dimensions; and that environmental differences force patterns of trait covariation across sites that can be different from patterns observed within sites.

With the exception of LDMC, which shows a particularly strong phylogenetic component, the trait-environment relationships found here should be amenable to process-based modelling. The energy balance implications of leaf size (Michaletz *et al.*, 2016; Dong *et al*., 2017b; Wright *et al.*, 2017) mean that this trait is crucial for survival, particularly in cold climates or in hot, dry climates. As the biophysical controls of leaf size are relatively well understood, it should be straightforward to build energy-balance constraints on leaf size into trait-based models. Shifts in the LES along environmental gradients could also be modelled, given the well-established relationship of leaf longevity and SLA (Wright *et al.*, 2004) and the experimentally determined variations of SLA with environmental factors (Poorter *et al.*, 2009). The distribution of SLA within communities could be represented by a pattern of covariation in leaf longevity, SLA, LDMC and the structural component of *N*area, as shown here and in other studies.

The co-ordination hypothesis predicts both *V*cmax and the ratio of *J*max to *V*cmax, including the observed dependence of both quantities on growth temperature (Wang *et al.*, 2017b). Large-scale patterns in *V*cmax and the metabolic component of *N*area can be predicted theoretically (Dong *et al.*, 2017a). The co-ordination hypothesis also predicts the observed seasonal acclimation of *V*cmax and *J*max (Togashi *et al.*, 2018). Thus, at the level of community mean values, it seems likely that *V*cmax can be successfully modelled as a function of environment (Ali *et al.*, 2016). A temperature-dependent ratio of *J*max to *V*cmax would then allow prediction of *J*max.

The CO2 drawdown from air to leaf, indexed by *χ*, is predicted by most vegetation models by simultaneous solution of the FvCB equations to predict assimilation rate as a function of leaf-internal CO2 (*c*i) and the diffusion equation to predict *c*i as a function of ambient CO2 (*c*a), stomatal conductance and assimilation rate (Farquhar *et al*., 1980). Theoretically and empirically well-founded relationships between *χ* and environmental variables (Wang *et al.*, 2017b) provide an alternative way to model *χ* directly as a function of environment, and thus to predict assimilation rates more straightforwardly than in many current models.

**Challenges and future directions**

This analysis illustrates the power of large trait data sets spanning a large range of climates, and including measurements from multiple co-existing species at each field site, to reveal general patterns. It also shows the utility of multivariate analysis to summarize patterns, and variance partitioning to attribute trait variability to different (and sometimes intersecting) causes. But despite the availability of large plant-trait data compilations (e.g. Kattge *et al.*, 2011), the number of sites that include all of any specified set of plant traits is often disappointingly small – because different research groups typically collect data on different sets of traits. There remains a need for more extensive trait data collection including photosynthetic traits and isotopic measurements in addition to conventional leaf traits, and for such data collection to extend to the full range of the world’s climates. There has been a limited amount of comparative work, for example, on photosynthetic traits, which are essential for all process-based vegetation modelling. Moreover, compared to leaf traits, there is a paucity of data on other field-measurable traits (notably stem hydraulic properties) that may be equally important for plant functional ecology. As is well illustrated by the global data sets that we used to test the predictive capacity of trait-climate relationships, the site- and/or species-metadata available are often limited. There remains a need for extensive, targeted collection and analysis of plant trait data, including co-located morphological, gas-exchange and isotopic measurements, and spanning the world’s major environmental and floristic gradients.

**Acknowledgments**

This research has been by supported by High-end Foreign Expert Programmes of China (GDW20156100290, GDW20166100147, GDW20181100161) (ICP and SPH), the National Natural Science Foundation of China (41701051, 31600388) (YY and HW), the National Basic Research Program of China (2013CB956600) (GL and CP), the Fundamental Research Funds for the Central Universities (YY), the QianRen Program, and the Natural Sciences and Engineering Research Council of Canada (NSERC) Discover Grant (CP). SPH acknowledges support from the ERC-funded project GC2.0 (Global Change 2.0: Unlocking the past for a clearer future, grant number 694481). This research contributes to the AXA Chair Programme in Biosphere and Climate Impacts and the Imperial College initiative on Grand Challenges in Ecosystems and the Environment (ICP). We thank O. Atkin, K. Crous, T. Domingues, D. Ellsworth, H. Togashi, Ü. Niinemets and L. Weerasinghe for providing the photosynthesis data (*V*cmax25, *J*max25) used in the validation.

**Author contributions**

YY, HW, SPH and ICP collectively devised the analysis strategy and interpreted the results. YY carried out all of the statistical analyses and wrote the first draft of the manuscript. IJW provided additional advice on the analysis and interpretation of trait variation patterns. CP and GL contributed to the revision of the text. All authors provided input to the final draft.

**References**

**Ackerly D. 2009.** Conservatism and diversification of plant functional traits: evolutionary rates versus phylogenetic signal. *Proceedings of the National Academy of Sciences* **106 (Suppl. 2)**: 19699-19706.

**Ackerly DD, Cornwell WK. 2007.** A trait-based approach to community assembly: partitioning of species trait values into within- and among-community components. *Ecology Letters* **10**: 135-145.

**Adler PB, Fajardo A, Kleinhesselink AR, Kraft NJ. 2013.** Trait‐based tests of coexistence mechanisms. *Ecology Letters* **16**: 1294-1306.

**Ali A, Xu C, Rogers A, Fisher R, Wullschleger S, McDowell N, Massoud E, Vrugt J, Muss J, Fisher J et al. 2016.** A global scale mechanistic model of the photosynthetic capacity (LUNA V1.0). *Geoscientific Model Development* **9**: 587-606.

**Armbruster WS, Pélabon C, Bolstad GH, Hansen TF. 2014.** Integrated phenotypes: understanding trait covariation in plants and animals. *Philosophical Transactions of the Royal Society B: Biological Sciences* **369**: 20130245.

**Armbruster WS, Schwaegerle KE.** **1996.** Causes of covariation of phenotypic traits among populations. *Journal of Evolutionary Biology* **9**:261-276.

**Bahar NHA, Ishida FY, Weerasinghe LK, Guerrieri R, O'Sullivan OS, Bloomfield KJ, Asner GP, Martin RE, Lloyd J, Malhi Y et al. 2017.** Leaf-level photosynthetic capacity in lowland Amazonian and high-elevation Andean tropical moist forests of Peru. *New Phytologist* **214**: 1002-1018.

**Borcard D, Legendre P, Drapeau P**.1992. [Partialling out the spatial component of ecological variation](http://dx.doi.org/10.2307/1940179). *Ecology* **84**: 511-525.

**Cernusak LA, Ubierna N, Winter K, Holtum JAM, Marshall JD, Farquhar GD. 2013.** Environmental and physiological determinants of carbon isotope discrimination in terrestrial plants. *New Phytologist* **200**: 950-965.

**Chen JL, Reynolds JF, Harley PC, Tenhunen JD. 1993.** Coordination theory of leaf nitrogen distribution in a canopy. *Oecologia* **93**: 63-69.

**Cornwell WK, Wright I, Turner J, Maire V, Barbour M, Cernusak L, Dawson T, Ellsworth D, Farquhar G, Griffiths H et al. 2017.** A global dataset of leaf ∆ 13 C values. 10.5281/zenodo.569501

**Cornelissen J, Lavorel S, Garnier E, Díaz S, Buchmann N, Gurvich D, Reich P, Ter Steege H, Morgan H, van der Heijden M. 2003.** A handbook of protocols for standardised and easy measurement of plant functional traits worldwide. *Australian Journal of Botany* **51**: 335-380.

**De Kauwe MG, Lin YS, Wright IJ, Medlyn BE, Crous KY, Ellsworth DS, Maire V, Prentice IC, Atkin OK, Rogers A. 2016.** A test of the 'one-point method' for estimating maximum carboxylation capacity from field-measured, light-saturated photosynthesis. *New Phytologist* **210**: 1130-1144.

**Davis TW, Prentice IC, Stocker BD, Thomas RT, Whitley RJ, Wang H, Evans BJ, Gallego-Sala AV, Sykes MT, Cramer W. 2017.** Simple process-led algorithms for simulating habitats (SPLASH v. 1.0): robust indices of radiation, evapotranspiration and plant-available moisture. *Geoscientific Model Development* **10**: 689-708.

**Díaz S, Purvis A, Cornelissen JH, Mace G M, Donoghue MJ, Ewers RM, Jordano P, Pearse WD. 2013.** Functional traits, the phylogeny of function, and ecosystem service vulnerability. *Ecology and Evolution*, **3**: 2958-2975.

**Díaz S, Kattge J, Cornelissen JHC, Wright IJ, Lavorel S, Dray S, Reu B, Kleyer M, Wirth C, Prentice IC et al. 2016.** The global spectrum of plant form and function. *Nature* **529**: 167-173.

**Dong N, Prentice IC, Evans BJ, Caddy-Retalic S, Lowe AJ, Wright IJ. 2017a.** Leaf nitrogen from first principles: field evidence for adaptive variation with climate. *Biogeosciences* **14**: 481-495.

**Dong N, Prentice IC**, **Harrison SP, Song Q, Zhang YP. 2017b.** Biophysical homoeostasis of leaf temperature: a neglected process for vegetation and land-surface modelling. *Global Ecology and Biogeography* **9**: 998-1007.

**Donovan LA, Mason CM, Bowsher AW, Goolsby EW, Ishibashi CD. 2014.** Ecological and evolutionary lability of plant traits affecting carbon and nutrient cycling. *Journal of Ecology* **102**:302-314

**Fajardo A, Piper FI. 2011.** Intraspecific trait variation and covariation in a widespread tree species (*Nothofagus pumilio*) in southern Chile. *New Phytologist***189:** 259-271.

**Farquhar G, von Caemmerer S, Berry J. 1980.** A biochemical model of photosynthetic CO2 assimilation in leaves of C3 species. *Planta* **149**: 78-90.

**Farquhar G, O'Leary M, Berry J. 1982.** On the relationship between carbon isotope discrimination and the intercellular carbon dioxide concentration in leaves. *Functional Plant Biology* **9**: 121-137.

**Funk JL, Cornwell WK. 2013.** Leaf traits within communities: context may affect the mapping of traits to function. *Ecology* **94**: 1893-1897.

**Fyllas NM, Gloor E, Mercado LM, Sitch S, Quesada CA, Domingues TF, Galbraith DR, Torre-Lezama A, Vilanova E, Ramírez-Angulo H et al. 2014.** Analysing Amazonian forest productivity using a new individual and trait-based model (TFS v.1). *Geoscientific Model Development* **7**: 1251-1269.

**Harrison SP, Morfopoulos C, Dani KGS, Prentice IC, Arneth A, Atwell BJ, Barkley MP, Leishman MR, Loreto F, Medlyn BE et al. 2013.** Volatile isoprenoid emissions from plastid to planet. *New Phytologist* **197**: 49-57.

**Hutchinson, M.2007.** ANUSPLIN Version 4.37 User Guide. Canberra: *The Australian National University.*

**James FC, McCulloch CE. 1990.** Multivariate Analysis in Ecology and Systematics:Panacea or Pandora's Box? *Annual Review of Ecology and Systematics* **21**:129−166.

**Kattge J, Díaz S, Lavorel S, Prentice IC, Leadley P, Bönisch G, Garnier E, Westoby M, Reich PB, Wright IJ et al. 2011.** TRY – a global database of plant traits. *Global Change Biology* **17**: 2905-2935.

**Kattge J, Knorr W. 2007.** Temperature acclimation in a biochemical model of photosynthesis: a reanalysis of data from 36 species. *Plant, Cell & Environment*, **30**: 1176-1190.

**Kikuzawa K. 1991.** A cost-benefit analysis of leaf habit and leaf longevity of trees and their geographical pattern. *American Naturalist* **138**: 1250-1263.

**Kohavi R. 1995.** A study of cross-validation and bootstrap for accuracy estimation and model selection. *International Joint Conference on Artificial Intelligence* **14**: 1137-1145.

**Kong D, Ma C, Zhang Q, Li L, Chen X, Zeng H, Guo D. 2014.** Leading dimensions in absorptive root trait variation across 96 subtropical forest species. *New Phytologist* **203**: 863-872.

**Kraft NJ, Godoy O, Levine JM. 2015.** Plant functional traits and the multidimensional nature of species coexistence. *Proceedings of the National Academy of Sciences* **112**: 797-802.

**Laughlin DC. 2014.** Applying trait‐based models to achieve functional targets for theory‐driven ecological restoration. *Ecology letters* **17:**771-784.

**Legendre P, Anderson MJ. 1999.** Distance‐based redundancy analysis: testing multispecies responses in multifactorial ecological experiments. *Ecological monographs.* **69**:1-24.

**Legendre P, Legendre L. 2012.** *Numerical ecology, 3rd Edition.* Amsterdam, Netherland: Elsevier Science.

**Liu X, Swenson NG, Wright SJ, Zhang L, Song K, Du Y J, Zhang JL, Mi XC, Ren HB, Ma KP. 2012**. Covariation in Plant Functional Traits and Soil Fertility within Two Species-Rich Forests. *PLoS One* **7**: e34767.

**Maire V, Martre P, Kattge J, Gastal F, Esser G, Fontaine S, Soussana JF. 2012.** The coordination of leaf photosynthesis links C and N fluxes in C3 plant species. *PLoS One* **7**: e38345.

**Maire V, Wright IJ, Prentice IC, Batjes NH, Bhaskar R, Bodegom PM van, Cornwell WK, Ellsworth D, Niinemets Ü, Ordonez A et al. 2015.** Global effects of soil and climate on leaf photosynthetic traits and rates. *Global Ecology and Biogeography* **24**:706-717.

**McMurtrie RE, Dewar RC. 2011.** Leaf-trait variation explained by the hypothesis that plants maximize their canopy carbon export over the lifespan of leaves. *Tree Physiology* **31**: 1007-1023.

**Meng TT, Wang H, Harrison SP, Prentice IC, Ni J, Wang G. 2015.** Responses of leaf traits to climatic gradients: adaptive variation versus compositional shifts. *Biogeosciences* **12**: 5339-5352.

**Michaletz ST, Weiser MD, McDowell NG, Zhou J, Kaspari M, Helliker BR, Enquist BJ. 2016.** The energetic and carbon economic origins of leaf thermoregulation. *Nature Plants* **2**: 16129.

**Moffat AM, Beckstein C, Churkina G, Mund M, Heimann M. 2010.** Characterization of ecosystem responses to climatic controls using artificial neural networks. *Global Change Biology* **16**:2737-2749.

**New M, Lister D, Hulme M, Makin I. 2002.** A high-resolution data set of surface climate over global land areas**.***Climate Research***, 21**: 1-25.

**Niinemets Ü, Keenan TF, Hallik L. 2014.** A worldwide analysis of within-canopy variations in leaf structural, chemical and physiological traits across plant functional types. *New Phytologist* **205**: 973-993.

**Oksanen J, Blanchet FG, Friendly M, Kindt R, Legendre P, McGlinn D, Minchin PR, O’Hara RB, Simpson GL, Solymos P, Stevens MH, Szoecs E. 2017.** *vegan: Community Ecology Package. R package Version 2.4-4.* [WWW document] URL <https://github.com/vegandevs/vegan>. [Accessed 1 May 2016].

**Onoda Y, Hikosaka K, Hirose T. 2004.** Allocation of nitrogen to cell walls decreases photosynthetic nitrogen-use efficiency. *Functional Ecology* **18**: 419-425.

**Onoda Y, Wright IJ, Evans JR, Hikosaka K, Kitajima K, Niinemets Ü, Poorter H, Tosens T, Westoby M. 2017.** Physiological and structural tradeoffs underlying the leaf economics spectrum. *New Phytologist* **214**: 1447-1463

**Osnas JL, Lichstein JW, Reich PB, Pacala SW. 2013.** Global leaf trait relationships: mass, area, and the leaf economics spectrum. *Science* **340**: 741-744.

**Papale D, Valentini R. 2003**. A new assessment of European forests carbon exchanges by eddy fluxes and artificial neural network spatialization. *Global change biology* **9**:525-535.

**Paruelo JM, Tomasel F**. **1997**. Prediction of functional characteristics of ecosystems: a comparison of Article neutral networks and regression models. *Ecological modelling* **98**:173-186.

**Peiman KS, Robinson BW. 2017.** Comparative analyses of phenotypic trait covariation within and among populations. *The American Naturalist* **190**: 451-468.

**Peres-Neto PR, Legendre P, Dray S, Borcard D. 2006.** Variation partitioning of species data matrices: estimation and comparison of fractions. *Ecology* **87**: 2614-2625.

**Picard RR, Cook RD. 1984.** Cross-validation of regression models. *Journal of the American Statistical Association* **79:** 575-583.

**Pierce S, Brusa G, Vagge I, Cerabolini, BEL. 2013**. Allocating CSR plant functional types: the use of leaf economics and size traits to classify woody and herbaceous vascular plants. *Functional Ecology* **27**: 1002-1010.

**Poorter H, Niinemets Ü, Poorter L, Wright IJ, Villar R. 2009.** Causes and consequences of variation in leaf mass per area (LMA): a meta-analysis. *New Phytologist* **182**: 565-588.

**Prentice IC, Dong N, Gleason SM, Maire V, Wright IJ. 2014.** Balancing the costs of carbon gain and water transport: testing a new theoretical framework for plant functional ecology. *Ecology Letters* **17**: 82-91.

**Prentice IC, Meng T, Wang H, Harrison SP, Ni J, Wang G. 2011.** Evidence of a universal scaling relationship for leaf CO2 drawdown along an aridity gradient. *New Phytologist* **190**: 169-180.

**Reich PB. 2014.** The world-wide ‘fast-slow’ plant economics spectrum: a traits manifesto. *Journal of Ecology* **102**: 275-301.

**Reich PB, Walters MB, Ellsworth DS. 1997.** From tropics to tundra: global convergence in plant functioning. Proceedings of the National Academy of *Sciences* **94**: 13730-13734.

**Scheiter S, Langan L, Higgins SI. 2013.** Next-generation dynamic global vegetation models: learning from community ecology. *New Phytologist* **198**: 957-969.

**Scheibe A, Steffens C, Seven J, Jacob A, Hertel D, Leuschner C, Gleixner G.** 2015. Effects of tree identity dominate over tree diversity on the soil microbial community structure. *Soil Biology and Biochemistry* **81**: 219-227.

**Shipley B, Laughlin DC, Sonnier G, Otfinowski R. 2011.** A strong test of a maximum entropy model of trait-based community assembly. *Ecology* **92**: 507-517.

**Swenson NG. 2013.** The assembly of tropical tree communities–the advances and shortcomings of phylogenetic and functional trait analyses. *Ecograph* **36:**264-76.

**Togashi HF, Prentice IC, Atkin OK, Macfarlane C, Prober SM, Bloomfield KJ, Evans BJ 2018.** Thermal acclimation of leaf photosynthetic traits in an evergreen woodland, consistent with the coordination hypothesis. *Biogeosciences* **15**: 3461–3474.

**Valladares F, Bastias CC, Godoy O, Granda E, Escudero A. 2015**. Species coexistence in a changing world. *Frontiers in Plant Science* **6**: 866.

**van Bodegom PM, Douma JC, Verheijen LM. 2014.** A fully traits-based approach to modeling global vegetation distribution. *Proceedings of the National Academy of Sciences* **111**: 13733-13738.

**Wang H, Prentice IC, Ni J. 2013.** Data-based modelling and environmental sensitivity of vegetation in China. *Biogeosciences* **10**: 5817-5830.

**Wang H, Prentice IC, Cornwell WK, Keenan TF, Davis TW, Wright IJ, Evans BJ, Peng C. 2017a.** Towards a universal model for carbon dioxide uptake by plants. *Nature Plants* **3**: 734-741.

**Wang H, Prentice IC, Davis TW, Keenan TF, Wright IJ, Peng C. 2017b.** Photosynthetic responses to altitude: an explanation based on optimality principles. *New Phytologist* **213**: 976-982.

**Wang H, Harrison SP, Prentice IC, Yang Y, Togashi HF, Wang M, Zhou S, Bai F, Ni J 2018**. The China Plant Trait Database: towards a comprehensive regional compilation of functional traits for land plants. *Ecology* **99**: 500.

**Watson RA, Szathmáry E. 2016.** How can evolution learn? *Trends in Ecology & Evolution* **31**:147-157.

**Westoby M, Falster DS, Moles AT, Vesk PA, Wright IJ. 2002.** Plant ecological strategies: Some leading dimensions of variation between species. *Annual Review of Ecology and Systematics* **33**: 125-159.

**White C, Tardif JC, Adkins A, Staniforth R. 2005.** Functional diversity of microbial communities in the mixed boreal plain forest of central Canada. *Soil Biology & Biochemistry* 37: 1359-1372.

**Wright IJ, Ackerly DD, Bongers F, Harms KE, Ibarra-Manriquez G, Martinez-Ramos M, Mazer SJ, Muller-Landau HC, Paz H, Pitman NCA et al. 2007.** Relationships among ecologically important dimensions of plant trait variation in seven neotropical forests. *Annals of Botany* **99**: 1003-1015.

**Wright IJ, Reich PB, Cornelissen JHC, Falster DS, Groom PK, Hikosaka K, Lee W, Lusk CH, Niinemets Ü, Oleksyn J et al. 2005.** Modulation of leaf economic traits and trait relationships by climate. *Global Ecology and Biogeography* **14**: 411-421.

**Wright IJ, Reich PB, Westoby M, Ackerly DD, Baruch Z, Bongers F, Cavender-Bares J, Chapin T, Cornelissen JH, Diemer M et al. 2004.** The worldwide leaf economics spectrum. *Nature* **428**: 821-827.

**Wright IJ, Dong N, Maire V, Prentice IC, Westoby M, Díaz S, Gallagher RV, Jacobs BF, Kooyman R, Law EA et al.** **2017**. Global climatic drivers of leaf size. *Science* **357**: 917-921.

**Supporting InformationTable S1** Leaf traits and their ecological significance

**Table S2** Characteristics of the sites

**Table S3** Total number of samples for each life form.

**Table S4** Trait loadings, eigenvalues, and the percentage of trait variation explained by successive principal components in the trait PCA (only sites with complete data).

**Table S5** Trait loadings, eigenvalues, and the percentage of trait variation explained by successive RDA axes (constrained by climate) and residual principal components.

**Table S6** Number of sites from the data sets used for the global assessment of trait-climate relationships (before and after screening).

**Table S7** Calibration accuracy of the predictive model of trait-climate RDA relationships for sites in China.

**Figure S1** Frequency distributions of traits in forest and non-forest sites.

**Figure S2** Trait dimensions from principal components analysis (sites with complete data).

**Figure S3** Climate-related trait dimensions from redundancy analysis (only sites with complete data).

**Figure S4** Residual (climate-independent) dimensions of trait variation (only sites with complete data).

**Figure S5** Distribution of sites used to test the trait predictions globally.

**Figure S6** Pairwise correlations between traits.

**Figure S7** Calibration of the predictive model of trait~climate RDA relationships in China.

**Figures Legends**

**Figure 1** Geographical and climatic coverage of the trait dataset. The individual sites are shown as red dots superimposed on a simplified vegetation map of China in (a); these sites have been grouped into eight named regions. The distribution of sites in climate space is shown in (b), where the moisture index (MI) is defined as the ratio of mean annual precipitation to annual equilibrium evapotranspiration, PAR0 is the accumulated photosynthetically active radiation during the thermal growing season, and the daily mean temperature during the thermal growing season (mGDD0) is shown by the colour of the dots. The grey shading indicates the frequency of different climates, as defined by MI and PAR0, in eastern China as a whole.

**Figure 2** Trait dimensions from principal component analysis: grey circles are species-site combinations. (a) PC1 *versus* PC2, (b) PC2 *versus* PC3, (c) PC3 *versus* PC4.The traits are LA: leaf area, SLA: specific leaf area, LDMC: leaf dry matter content, *N*area: leaf nitrogen per unit area, *V*cmax25: maximum carboxylation rate standardized to 25˚C, *J*max25: maximum electron transport rate standardized to 25˚C, and *χ* :the ratio of intercellular to ambient CO2 concentration. The four axes of variability related to LA, *χ*, the leaf economic spectrum and the photosynthetic traits are shown by coloured ellipses on each plot.

**Figure 3** Climate-related trait dimensions from redundancy analysis: grey circles are species-site combinations and coloured dots signify named regions as defined in Fig. 1. (a) RDA1 *versus* RDA2, (b) RDA2 *versus* RDA3. The traits are: LA: leaf area, SLA: specific leaf area, LDMC: leaf dry matter content, *N*area: leaf nitrogen per unit area, *V*cmax25: maximum carboxylation rate standardized to 25˚C, *J*max25: maximum electron transport rate standardized to 25˚C, and *χ*: the ratio of intercellular to ambient CO2 concentration. The climate variables are the ratio of mean annual precipitation to annual equilibrium evapotranspiration (MI), the accumulated photosynthetically active radiation during the thermal growing season (PAR0) and the daily mean temperature during the thermal growing season (mGDD0).

**Figure 4** Residual (climate-independent) dimensions of trait variation: grey circles are species-site combinations. (a) PC1 *versus* PC2, (b) PC2 *versus* PC3, (c) PC3 *versus* PC4.The traits are: LA: leaf area, SLA: specific leaf area, LDMC: leaf dry matter content, *N*area: leaf nitrogen per unit area, *V*cmax25: maximum carboxylation rate standardized to 25˚C, *J*max25: maximum electron transport rate standardized to 25˚C, and *χ*: the ratio of intercellular to ambient CO2 concentration.

**Figure 5** Partial correlations between traits after removal of climate effects. The traits are: LA: leaf area, SLA: specific leaf area, LDMC: leaf dry matter content, *N*area: leaf nitrogen per unit area, *V*cmax25: maximum carboxylation rate standardized to 25˚C, *J*max25: maximum electron transport rate standardized to 25˚C, and *χ*: the ratio of intercellular to ambient CO2 concentration. Colours indicate the strength of the correlation, where dark blue indicates perfect correlation. n.s. stands for not significant.

**Figure 6** Variance partitioning (%) for all traits considered together, and each trait separately. The traits are: LA: leaf area, SLA: specific leaf area, LDMC: leaf dry matter content, *N*area: leaf nitrogen per unit area, *V*cmax25: maximum carboxylation rate standardized at 25˚C, *J*max25: maximum electron transport rate standardized at 25˚C, and *χ*: the ratio of intercellular to ambient CO2 concentration.

**Figure 7** Predicting traits globally at site level, from the trait-climate relationships derived from data in China. The traits are: LA: leaf area, SLA: specific leaf area, LDMC: leaf dry matter content, *N*area: leaf nitrogen per unit area, *V*cmax25: maximum carboxylation rate standardized to 25˚C, *J*max25: maximum electron transport rate standardized to 25˚C, and *χ*: the ratio of intercellular to ambient CO2 concentration. (a) Predicted loge √LA versus observed loge √LA (Wright et al., 2017). (b) Predicted loge SLA versus observed loge SLA (Wright et al., 2004). (c) Predicted loge LDMC versus observed loge LDMC (Wright et al., 2004). (d) Predicted loge *N*area versus observed loge *N*area (Wright et al., 2004). (e) Predicted loge *V*cmax25 versus observed loge *V*cmax25 (De Kauwe et al., 2016). (f) Predicted loge *J*max25 versus observed loge *J*max25 (De Kauwe et al., 2016). (g) Predicted logit χ versus observed logit χ (Cornwell et al., 2017). Gray points are species trait values and red squares are site means. Blue lines are the regresion lines and their shades are 95% confidence intervals.

Table 1 Trait loadings, eigenvalues, and the percentage of trait variation explained by successive principal components in the trait PCA. Loadings > 0.3 in magnitude are shown in **bold**.

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
|  | PC1 | PC2 | PC3 | PC4 |
| loge √LA | **0.57** | **0.69** | 0.29 | **0.31** |
| loge SLA | 0.07 | 0.04 | **0.61** | 0.28 |
| loge LDMC | 0.04 | 0.03 | **0.31** | 0.09 |
| loge *N*area | 0.12 | 0.11 | **0.60** | 0.24 |
| loge *V*cmax,25 | 0.19 | 0.24 | 0.23 | **0.70** |
| loge *J*max,25 | 0.16 | 0.19 | 0.17 | **0.52** |
| logit *χ* | **0.76** | **0.64** | 0.05 | 0.02 |
| **Eigenvalue** | 2.57 | 0.90 | 0.50 | 0.25 |
| **Explained (%)** | 58.0 | 20.4 | 11.3 | 5.6 |
| **Cumulative (%)** | 58.0 | 78.5 | 89.8 | 95.4 |

Table 2 Trait loadings, eigenvalues, and the percentage of trait variation explained by successive RDA axes (constrained by climate) and residual principal components, with axes 1 and 2 mirrored to facilitate comparison with the PCA. Loadings > 0.3 in magnitude are shown in **bold**.

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
|  | RDA1 | RDA2 | RDA3 | PC1 | PC2 | PC3 | PC4 |
| loge √LA | **0.66** | 0.24 | **0.51** | 0.12 | **0.85** | **0.44** | 0.25 |
| loge SLA | 0.01 | **0.67** | 0.11 | 0.11 | 0.20 | **0.53** | **0.33** |
| loge LDMC | 0.02 | 0.14 | **0.43** | 0.08 | 0.05 | **0.32** | 0.17 |
| loge *N*area | 0.15 | **0.67** | **0.30** | 0.04 | 0.18 | **0.55** | **0.30** |
| loge *V*cmax,25 | 0.22 | 0.07 | 0.19 | 0.04 | **0.33** | 0.26 | **0.68** |
| loge *J*max,25 | 0.18 | 0.11 | 0.29 | 0.05 | 0.26 | 0.22 | **0.49** |
| logit *χ* | **0.67** | 0.08 | **0.58** | **0.98** | 0.17 | 0.07 | 0.04 |
| **Eigenvalue** | 1.55 | 0.08 | 0.02 | 1.19 | 0.75 | 0.42 | 0.24 |
| **Explained (%)** | 34.9 | 1.8 | 0.4 | 26.8 | 17.0 | 9.6 | 5.3 |
| **Cumulative (%)** | 34.9 | 36.7 | 37.1 | 63.9 | 80.9 | 90.5 | 95.9 |

Table 3 Total contributions (%) of climate, family, site and life form to trait variation. Standard deviations (weights) of the transformed variables are also given. For each trait, values less than 0.5 or more than 1.5 times the all-traits value are shown in *italics* and **bold** respectively.

|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  | All traits | loge √LA | loge SLA | loge LDMC | loge *N*area | loge *V*cmax25 | loge *J*max25 | logit χ |
| Weights |  | 1.17 | 0.50 | 0.38 | 0.59 | 0.58 | 0.48 | 1.37 |
| Climate | 37.2 | 51.3 | *14.6* | *3.8* | 24.6 | 23.5 | 28.0 | 38.0 |
| Family | 54.6 | 60.6 | 40.4 | 47.9 | 36.6 | 38.8 | 46.2 | 59.1 |
| Site | 49.4 | 59.4 | 35.8 | *17.9* | 39.5 | 33.7 | 37.8 | 51.9 |
| Life form | 16.3 | **25.7** | *7.4* | *9.5* | *1.3* | *3.4* | *5.1* | 16.8 |

Table 4 Prediction accuracy of the trait-climate RDA model for independent global data sets at site level. \* indicates that the slope is significantly different from 1 (*P* < 0.01), # indicatesthat the intercept is significantly different from 0 (*P* < 0.01). \*\* indicates that the regression is significant (*P* < 0.01). n.s. stands for not significant.

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| Traits | Slope | Intercept |  | *n* | RMSE | Source of data |
| loge √LA | 0.60\*  (0.52, 0.70) | –1.45#  (–1.72, –1.10) | 0.34\*\* | 388 | 0.70 | Wright et al. (2017) |
| loge SLA | 0.99  (0.68, 1.31) | –0.61  (–1.41, 0.19) | 0.31\*\* | 87 | 0.53 | Wright et al. (2004) |
| loge LDMC | n.s. | n.s. | 0.01 | 9 | 0.20 | Wright et al. (2004) |
| loge *N*area | 0.38\*  (0.24, 0.52) | 0.45#  (0.34, 0.56) | 0.28\*\* | 77 | 0.26 | Wright et al. (2004) |
| loge *V*cmax25 | 1.16  (0.62, 1.69) | –0.11  (–1.97, 1.76) | 0.33\*\* | 38 | 0.40 | De Kauwe et al. (2016) |
| loge *J*max25 | 0.59\*  (0.27, 0.92) | 1.99#  (0.62, 3.36) | 0.25\*\* | 38 | 0.33 | De Kauwe et al. (2016) |
| logit χ | 0.48\*  (0.40, 0.57) | 0.35#  (0.30, 0.40) | 0.33\*\* | 281 | 0.29 | Cornwell et al. (2017) |