

# **An early and late peak in microglial activation in Alzheimer's disease trajectory**

Zhen Fan<sup>1</sup>, David J. Brooks<sup>1,2</sup>, Aren Okello<sup>1</sup>, Paul Edison<sup>1</sup>

<sup>1</sup>Neurology Imaging Unit, Imperial College London, Hammersmith Hospital, Du

Cane Road, London, W12 0NN, UK.

<sup>2</sup>Department of Nuclear Medicine, Institute of Medicine, Aarhus University, Denmark

## **Corresponding author's full address:**

Dr Paul Edison MBBS, MRCP, MPhil, PhD, FRCPI

Neurology Imaging Unit, Imperial College London, B Block, Hammersmith Hospital Campus,

Du Cane Road, London, W12 0NN, UK.

Tel: +44(0)20 3313 3725      Fax: +44(0)20 8383 1783

Email: paul.edison@imperial.ac.uk

**Running head:** Neuroinflammation in Alzheimer's trajectory

## **Abstract**

Beta-amyloid deposition, neuroinflammation and tau tangle formation all play a significant role in Alzheimer's disease. We hypothesised that there is microglial activation early on in Alzheimer's disease trajectory, where in the initial phase, microglial may be trying to repair the damage, while later on in the disease these microglia could be ineffective and produce proinflammatory cytokines leading to progressive neuronal damage. In this longitudinal study, we have evaluated the temporal profile of microglial activation and its relationship between fibrillar amyloid load at baseline and follow-up in subjects with mild cognitive impairment, and this was compared with subjects with Alzheimer's disease. Thirty subjects (8 mild cognitive impairment, 8 Alzheimer's disease and 14 controls) aged between 54-77 years underwent [11C](R)PK11195, [11C]PIB PET and MRI scans. Patients were followed up after  $14 \pm 4$  months. Region of interest and Statistical Parametric Mapping analysis were used to determine longitudinal alterations. Single subject analysis was performed to evaluate the individualized pathological changes over time. Correlations between levels of microglial activation and amyloid deposition at a voxel level were assessed using Biological Parametric Mapping. We demonstrated that both baseline and follow-up microglial activation in the mild cognitive impairment cohort compared to controls were increased by 41% and 21%, respectively. There was a longitudinal reduction of 18% in microglial activation in mild cognitive impairment cohort over 14 months, which was associated with a mild elevation in fibrillar amyloid load. Cortical clusters of microglial activation and amyloid deposition spatially overlapped in the subjects with mild cognitive impairment. Baseline microglial activation was increased by 36% in Alzheimer's disease subjects compared with controls. Longitudinally, Alzheimer's disease subjects showed an increase in microglial activation. In conclusion, this is one of the first longitudinal PET studies evaluating longitudinal changes in microglial activation in mild cognitive impairment and Alzheimer's disease subjects. We found

there is an initial longitudinal reduction in microglial activation in subjects with mild cognitive impairment, while Alzheimer's disease subjects showed an increase in microglial activation. This could reflect that activated microglia in mild cognitive impairment initially may adopt a protective activation phenotype, which later change to a pro-inflammatory phenotype as disease progresses and amyloid clearance fails. Thus, we speculate that there might be two peaks of microglial activation in the Alzheimer's disease trajectory; an early protective peak and a later pro-inflammatory peak. If so, anti-microglial agents targeting the pro-inflammatory phenotype would be most beneficial in the later stages of the disease.

**Keywords:** Alzheimer's disease; mild cognitive impairment; microglial activation; amyloid imaging; neuropathology.

**Abbreviations:** *TSPO* = 18-kDa mitochondrial translocator protein; ROI = region of interest; SPM = Statistical Parametric Mapping; BPM = Biological Parametric Mapping; MNI = Montreal Neurologic Institute; CNS = central nervous system; TREM2 = myeloid cells 2; IL = interleukin;

## **Introduction**

Amnesic mild cognitive impairment is a transitional stage between cognitively normal individuals and Alzheimer's disease. The amyloid cascade-neuroinflammation hypothesis suggests that an increase in levels of beta-amyloid, due to its excess synthesis or a failure of clearance, is the primary event and that this leads to neuroinflammation and subsequent neuronal damage. However, other studies have emphasised an early role of neuroinflammation in the disease process (Heneka et al., 2015; Okello et al., 2009).

It is now accepted that neuroinflammation plays a significant role in Alzheimer's disease process (Amor et al., 2010; Heneka et al., 2015). Activated microglia can adopt two main functional states: pro-inflammatory and anti-inflammatory states of microglial activation. It has been suggested that in the early stages of Alzheimer's disease, the initial microglial activation may serve a protective function (anti-inflammatory) trying to clear the amyloid and release nerve growth factors (Hamelin et al., 2016). When this process fails due to accumulation of beta-amyloid or other toxic products, there is activation of pro-inflammatory phenotypes, which release pro-inflammatory cytokines, leading to self-perpetuating damage to the neurons. However, genetic data from GWAS suggest that microglial activation may play a significant role early on in the disease and independent of amyloid pathology (R. Guerreiro and Hardy, 2013; R. J. Guerreiro and Hardy, 2011; Hollingworth et al., 2011a; Hollingworth et al., 2011b; Varnum and Ikezu, 2012). This is further supported by evidence from ageing brain, where microglial activation and neurodegeneration can be present in the absence of amyloid plaques (von Bernhardi et al., 2015). Epidemiological studies have shown a lower prevalence of Alzheimer's disease in people taking anti-inflammatory NSAIDs, suggesting a potential protective role of lowering neuroinflammation, although randomised trials have not shown efficacy of these agents in subjects with established Alzheimer's disease (Jaturapatporn et al., 2012). However, no studies to date have evaluated levels of microglial activation in subjects

with mild cognitive impairment longitudinally to assess the influence of microglial activation in the Alzheimer's disease trajectory. We hypothesised that there is microglial activation in subjects with mild cognitive impairment early on in the disease process, which may decrease over time as the protective phenotype becomes ineffective, while during later stages of the Alzheimer's disease trajectory, there is persistent increase in microglial activation.

TSPO expression in the healthy brain is very low, but it elevates after brain injury and neuroinflammation, therefore TSPO has been widely used as a PET imaging biomarker to visualise microglial activation in CNS (Fan et al., 2015a; Varley et al., 2015). The aim of the present study was to evaluate the influence of microglial activation in Alzheimer's disease trajectory using [11C](R)PK11195 PET in subjects with mild cognitive impairment and Alzheimer's disease and how these changes are influenced by the levels of amyloid deposition measured with [11C]PIB PET longitudinally.

## **Materials and Methods**

Eight participants who had a diagnosis of mild cognitive impairment based on the Petersen criteria and 14 healthy controls were recruited into this study. These subjects were compared with our previous data on eight subjects with Alzheimer's disease (Fan et al., 2015b). All subjects underwent detailed neurological examinations. All subjects underwent [11C](R)PK11195, [11C]PIB PET scans, and T1/T2 volumetric MRI to evaluate any structural lesions at baseline. Subjects with mild cognitive impairment and Alzheimer's disease were followed up with repeat [11C](R)PK11195 PET, [11C]PIB PET and MRI after 14 $\pm$ 4 months.

### *Image acquisition*

MRI scans were acquired with a 1.5 Tesla GE scanner (acquisition times repetition time 30 ms, echo time 3 ms, flip angle 30 degrees, field of view 25 cm, matrix 156  $\times$  256, voxel dimensions 0.98  $\times$  0.98  $\times$  1.6 mm). ECAT EXACT HR ++ (CTI/Siemens, Knoxville, TN) PET scanner

was used to acquire [11C](R)PK11195 PET, while [11C]PIB scans were acquired using ECAT EXACT HR+ scanner. T1-weighted MRI was used to generate an individualised brain atlas as detailed below, whereas T2-weighted MRI was acquired to exclude any structural abnormalities. A mean activity of  $290 \pm 16$  MBq [11C](R)PK11195 was injected intravenously and scanned for 60mins to generate an dynamic [11C](R)PK11195 PET (Edison et al., 2008). For [11C]PIB PET scans,  $370 \pm 18$  MBq [11C]PIB was injected and scanned for 90mins.

### *Image processing*

Owing to widespread distribution of microglial activation, it is difficult to identify a single region as reference. Therefore, supervised cluster analysis was used to generate parametric images of [11C](R)PK11195 BP (binding potentials) for each individual by extracting distributed clusters of voxels that kinetically behaved either like a normal population cortical reference input function or showed evidence of tracer retention (Anderson et al., 2007; Turkheimer et al., 2007). With the supervised clustering algorithm, the [11C](R)PK11195 PET shows a better test-retest variability (10.6%, ICC=0.878) than unsupervised approaches (Turkheimer et al., 2007). A summed 60-90 minute add image was used to create [11C]PIB SUVR (RATIO) maps with cerebellar cortex as the reference tissue.

### *Region of Interest analysis*

We used statistical parametric mapping software, SMP8 (<http://www.fil.ion.ucl.ac.uk/spm>) and Analyze 11 (AnalyzeDirect) for image analysis. Initially, [11C](R)PK11195 BP and [11C]PIB PET (60-90 min) images were co-registered to the individual MRI space. Then the subjects' MRIs were segmented to grey matter, white matter, and cerebrospinal fluid. The grey matter binary image was created at 50% threshold. Probabilistic brain atlas in MNI space was transformed to individual MRI space. The grey matter binary image was then multiplied with the probabilistic brain atlas in Analyze11 to create an individualised object map. [11C]PIB

RATIO images were created by dividing the individual PET with cerebellar grey matter [11C]PIB uptake. Finally, the [11C](R)PK11195 BP and [11C]PIB RATIO images were sampled in the following regions: frontal, parietal, temporal, and occipital cortices, anterior, posterior cingulate, thalamus striatum and hippocampus. In addition, partial volume correction (PVC) was applied to the [11C](R)PK11195 BP PET data at baseline and during follow up to minimize the impact of brain atrophy using PVC\_SFSRR software (Shidahara et al., 2009).

### *SPM analysis*

All PET and MRI scans were then normalised into MNI space and smoothed using a 6\*6\*6 mm filter in SPM. Initially a cluster threshold of  $p < 0.05$  with extent threshold of 50 voxels were applied, if the significance was reached (Supplementary Figure 1), higher threshold was applied to delineate minimal changes. Between-group clusters of significant mean differences at a voxel level between subjects with mild cognitive impairment and controls were localised with SPM8 by applying the Student's t-test with a threshold of  $p < 0.01$  and cluster extent threshold 50 voxels for [11C](R)PK11195 PET; and a threshold of  $p < 0.005$  and cluster extent threshold 50 voxels for [11C]PIB. Similar thresholds were used for the follow-up PET scans. The overall change from baseline to follow up in microglial activation and amyloid deposition were assessed using paired t-test in SPM with a threshold of  $p < 0.01$  with an extent threshold of 50 voxels. Similar SPM analysis were performed for subjects with Alzheimer's disease (Fan et al., 2015b).

### *Single subject analyses*

As a classical ROI approach for interrogating inhomogeneous microglial activation tends to underestimate the localised voxel-wise small changes within any predefined ROI region, we developed individualised SPM maps by comparing each patient against a group of healthy controls to localise significant clusters of binding change (Edison et al., 2013). VOI maps for

increased [11C](R)PK11195 BP and [11C]PIB uptake for each subject were generated by extracting the significant clusters from the single subject result in SPM8. The volume of each pathological process was calculated using Analyze11 (Fan et al., 2015a).

### *3D intensity T-map*

We developed 3D intensity T-maps in MATLAB to demonstrate a clear visualisation of the significant changes in microglial activation and amyloid between baseline and follow-up. Each 3D intensity T-map utilises the SPM T-map to generate surface plots where the colour intensity and height reflects the T-score for the corresponding voxel with significant increase or decrease in the microglial activation and amyloid deposition.

### *BPM correlations*

Voxel-wise statistical correlations between [11C](R)PK11195 BP and [11C]PIB RATIO were performed using the Biological Parametric Mapping (Casanova et al., 2007), which provides sophisticated voxel-wise comparison across imaging modalities with  $p < 0.05$ . In order to directly compare different pathological processes using PET scans, Z-score maps were created to represent the brain profiles for levels of microglial activation and amyloid deposition using the following formulae for BPM analysis:

$$Z[11C](R)PK11195 = \frac{[11C](R)PK11195BP - \bar{X} Control[11C](R)PK11195BP}{SD of Control[11C](R)PK11195BP}$$

$$Z [11C]PIB = \frac{[11C]PIB - \bar{X} Control[11C]PIB}{SD of Control[11C]PIB},$$

where Z denotes Z-score map,  $\bar{X}$  denotes mean, and SD denotes standard deviation.

As the predefined ROI regional mean may underestimate the true changes at voxel level, hence a pixel-wise Pearson correlation graph (scatter plot) between the group mean image of [11C]PIB RATIO PET and [11C](R)PK11195 BP PET was applied in the whole cortex and in frontal, temporal, parietal and occipital cortices. The scatter plots were generated for all subjects with mild cognitive impairment, subgroup of amyloid positive subjects and subgroup



of amyloid negative subjects.

### *Statistical analysis*

All ROI data statistical analysis was performed using SPSS for Windows version 22 (SPSS, Chicago, Illinois, USA). The p values for voxel-wise SPM comparison was corrected for family-wise error rate (FEW) at cluster level. Mean differences between patients and healthy controls, as well as the regional differences between baseline and follow-up were calculated using Student's t-test. The Pplot multiple comparison (Turkheimer et al., 2001) was applied to the regional paired t-test between baseline and follow-up [11C](R)PK11195 BP and [11C]PIB RATIO using Hochberg multiple comparison correction.

## **Results**

Demographics are shown in Table 1. There were no significant group differences in age, age range or gender. The majority of subjects with mild cognitive impairment demonstrated a deterioration in MMSE score at follow-up, while one subject the MMSE score changed from 25/30 to 28/30.

### *[11C](R)PK11195 PET*

The mild cognitive impairment subjects as group showed longitudinal regional reductions in microglial activation. ROI analysis detected significant reductions in [11C](R)PK11195 BP in frontal, temporal, parietal, and occipital cortices, anterior cingulate, posterior cingulate and hippocampus when baseline data was compared against 14 months' follow-up (Table 2A). This longitudinal reduction in microglial activation in mild cognitive impairment was still persistent even after the data was corrected for partial volume effect (Table 3). Apart from frontal cortex, all the above regions remained significant after correcting for multiple comparison (Turkheimer et al., 2001). Individually, ROI analysis found that 6/8 subjects with mild cognitive impairment showed  $20 \pm 6\%$  mean reductions of [11C](R)PK11195 BP at follow-up.

This longitudinal reduction in microglial activation from baseline to follow up was further supported by the reduction demonstrated using paired t-test in SPM (Fig. 1A). Based on the amyloid load at baseline, there were four amyloid positive subjects who demonstrated global increase in amyloid deposition compared with controls, while the other four showed no elevation in amyloid load in any ROIs. When the subjects were separated as amyloid positive and amyloid negative subjects, amyloid positive group demonstrated significant reduction in microglial activation using ROI and SPM analysis in temporal, parietal and occipital cortices, anterior cingulate, posterior cingulate and hippocampus over time. However, the amyloid negative subject group demonstrated only small clusters of reduction in microglial activation using SPM analysis. In the amyloid negative subjects, there was a trend of reduction in microglial activation using ROI analysis, however, it didn't reach statistical significance (Fig. 1 and Table 2B). [11C](R)PK11195 PET shows a test-retest variability of 10.6% (ICC=0.878) (Turkheimer et al., 2007). The reductions seen in microglial activation in this study are up to 33% to 42%; and are unlikely to be due to repeat measurements.

Single subject volumetric analysis (VOI) found that the total number of voxels with significantly raised microglial activation decreased by  $50 \pm 27\%$  during follow up (Fig. 2E and 2F, and Table 4). Compared to the healthy controls, SPM localised significant increases in microglial activation in subjects with mild cognitive impairment at both baseline and at follow-up in medial temporal lobe, hippocampus, anterior temporal lobe, posterior temporal lobe, parietal and frontal gyri at a cluster threshold of  $p < 0.01$  and extent threshold of 50 voxels (Fig. 2A and 2B).

Subjects with Alzheimer's disease showed 30% ( $p < 0.03$ ) higher microglial activation compared to healthy controls in four cortices, hippocampus and striatum at baseline, while this microglial activation increased to 46% ( $p < 0.01$ ) compared to healthy controls after 14 months' follow-up as previously reported (Supplementary Table 1). Six out of eight subjects with

Alzheimer's disease individually showed a longitudinal increase in the total volume of microglial activation measured by single subject VOI analysis (Table 4). (Fan et al., 2015b)

### *[11C]PIB PET*

Longitudinally, subjects with mild cognitive impairment showed around 6% increases in amyloid deposition in frontal lobe, temporal lobe, occipital lobe and posterior cingulate at ROI level based on the paired t-test (Table 2A). Individually, six subjects with mild cognitive impairment who experienced longitudinal decreases in microglial activation demonstrated small increases (6%) in mean [11C]PIB uptake across ROIs, including posterior cingulate, striatum, hippocampus and frontal, temporal, parietal and occipital cortices. VOI analysis demonstrated a 39% increase in the total number of voxels with significantly increased amyloid deposition. Interestingly, one patient (MCI8) who revealed a longitudinal increase in microglial activation (both regional BP and larger volume of microglial activation using single subject SPM analysis) during follow-up, showed a 2% reduction in mean amyloid uptake RATIO and a 26% reduction in the total number of voxels (using single subject SPM) over time (Table 4). Similar to Alzheimer's disease subjects (Fan et al., 2015b), subjects with mild cognitive impairment also demonstrated whole brain elevation of amyloid deposition compared with healthy controls at baseline and follow-up (Fig. 2C, 2D and Table 2A).

### *Correlation*

In subjects with mild cognitive impairment, BPM revealed a positive correlation between microglial activation and amyloid deposition at baseline in frontal, temporal and occipital lobe at threshold of  $p < 0.05$  (Fig. 3A). At follow-up, the clusters with a positive correlation at baseline had expanded in frontal, temporal, and parietal cortical regions using the same threshold (Fig. 3B). In subjects with Alzheimer's disease, clusters of microglial activation were positively correlated with amyloid deposition mainly in temporal lobe, parietal lobe and

occipital lobe at baseline, and this positive correlation has persisted at follow-up (Fig. 3C and 3D).

Using pixel-wise correlation, we found significant correlation (Pearson) ( $r=0.37$ ,  $p<0.0001$ ) between [11C](R)PK11195 BP and [11C]PIB RATIO in the composite cortical region (combining frontal, temporal, parietal and occipital cortical region), and individually in all the four cortical regions in MCI subjects (Supplementary Figure 2A). When we separated the MCI subjects based on amyloid status, amyloid positive MCI subjects demonstrated even more significant correlation ( $r=0.53$ ,  $p<0.0001$ ) in the composite cortex and individually in different cortices, while amyloid negative MCI subjects failed to show correlation between amyloid deposition and microglial activation (Supplementary Figure 2B).

## **Discussion**

In this study, we have detected a longitudinal reduction in microglial activation in a majority of our subjects with mild cognitive impairment, associated with a marginal increase in amyloid load. This contrasted with subjects with Alzheimer's disease who showed an increase in their microglial activation over 14 months (Fan et al., 2015b). These findings could be explained by the notion that there is initially protective microglial activation in the prodromal stage of Alzheimer's disease trying to clear the amyloid which then fails, while during the later stages of disease there is progressive increase in microglial pro-inflammatory activation. This study also demonstrates that microglial activation is positive correlated with amyloid load. Such an association is compatible with microglial cells being activated by amyloid, but some of the microglial activation could also occur independently of amyloid deposition and may be triggered by other factors. This suggests a bimodal distribution of microglial activation in Alzheimer's trajectory as depicted in Fig. 4.

This is the first study demonstrates that there are two peaks of microglial activation in the Alzheimer's disease trajectory in vivo. For the first peak, significant microglial activation in

the subjects with mild cognitive impairment at baseline is consistent with pathological studies showing that there is significant inflammation during the initial stages of the disease. For instance, the increased YKL-40, a putative indicator of inflammation, was found in preclinical stages of AD, indicating neuroinflammation could occur in the early stage (Craig-Schapiro et al., 2010). It has also been suggested that, in the initial stages of the disease, microglia are trying to clear amyloid and other toxic products as they accumulate (Pan et al., 2011; Rogers et al., 2002). This notion is supported by the presence of microglial cells surrounding amyloid plaques in Alzheimer's disease cerebral cortex, the presence of A $\beta$  deposits in T-cells, activated microglia and reactive astrocytes in the brains of subjects with mild cognitive impairment (Sastre et al., 2006). This could explain our finding of increased microglial activation in the early stages of the disease, as in subjects with mild cognitive impairment. It is suggested that failure of the microglia to clear amyloid leads to accumulation of amyloid. One could argue that this may be due to switching from protective to pro-inflammatory state at this stage, causing a decrease in anti-inflammatory microglial activation and, as disease advances as in Alzheimer's disease, there is an increase in pro-inflammatory microglial activation, leading to two peaks of microglial activation. Recent studies on TREM2 further substantiate an early and late peak of microglial activation during the Alzheimer's disease trajectory (Heslegrave et al., 2016; Piccio et al., 2016). TREM2, expressed on microglia, are believed to function in microglial activation. While previous studies have reported discrepant results for TREM2 levels in AD, recently a large number of subjects have demonstrated that the level of soluble TREM2 might be dependent on the different stages of AD, where TREM2 has a protective role and peaks during the early stage of the disease and later the levels drop as disease further progresses (Jiang et al., 2013; Suárez - Calvet et al., 2016).

A recent PET study has demonstrated a protective role for microglial activation using a TSPO marker, [18F]DPA-714, which has showed higher TSPO binding in slow decliners than fast

decliners, with no difference in amyloid load (Hamelin et al., 2016). Hamelin et al have also demonstrated a positive correlation between TSPO binding and MMSE score and grey matter volume, which further indicates microglial activation may be beneficial in the early phase of the disease. Studies in later stages of Alzheimer's disease have suggested progressive increase in microglial activation. In this study, we have demonstrated that there is an early peak of microglial activation in MCI stage, and one could speculate that in the early stages microglia are trying to prevent the damage by the protective mechanism.

While multiple studies have suggested that microglial activation is present in the early stage of AD, it is still argued whether microglial activation is beneficial or harmful to the neurons and, as recent studies suggest, it is possible that different microglial functional phenotypes could have different effects (Cunningham, 2013; Lyman et al., 2014). Currently, there is compelling evidence to indicate that microglia may change from one phenotype to another during different phases of the disease (Cai et al., 2013; Lynch, 2013). In this study, a longitudinal decline in TSPO binding was observed in subjects with mild cognitive impairment. This could be explained as disease advances, it is possible that protective anti-inflammatory microglial activation could become ineffective and exhausted over a period of time, resulting in predominant activation of pro-inflammatory microglia releasing multiple pro-inflammatory cytokines causing neuronal damage and neurodegeneration (Varnum and Ikezu, 2012), which leads to a second peak as we have reported in subjects with Alzheimer's disease. On an individual basis, a majority of subjects with mild cognitive impairment had early increased microglial activation at baseline, and this reduced longitudinally. In one subject with mild cognitive impairment, there was a longitudinal increase in microglial activation (12%) and an associated reduction in amyloid deposition (5%) with an improvement in cognitive function, as the MMSE score increased from 25 to 28. Even though it is a single subject, one could speculate that this could be due to further activation of the protective anti-inflammatory

phenotype which helped to clear A $\beta$  in the early stage of the disease (Cai et al., 2013; Prokop et al., 2013; Zotova et al., 2013).

While [11C](R)PK11195 PET biomarker cannot differentiate between different microglial activation (functional) states, recent studies have demonstrated the shift from the anti-inflammatory to pro-inflammatory state plays a central role in Alzheimer's disease pathogenesis, which may be triggered by changes in the micro-environment in the brain (Sanchez-Guajardo et al., 2013; Varnum and Ikezu, 2012). In the current study, we found that amyloid positive subjects with mild cognitive impairment have revealed a longitudinal reduction in microglial activation during follow-up, while relatively less reduction was found in amyloid negative subjects. This may indicate that amyloid pathology causes significant microglial activation early on in the disease process, perhaps in an attempt to clear the amyloid load. One could argue that the presence of the amyloid causes excessive glial activation in the beginning, and as disease advances, these microglia become ineffective (less activated) at an accelerated rate than those subjects who are amyloid negative (Lim et al., 2014; Villemagne et al., 2013). Several PET studies evaluating microglia in vivo have demonstrated conflicting results, some suggesting increased microglial activation in early stages of the disease (Hamelin et al., 2016; Yasuno et al., 2012) while others found no microglial activation (Schuitemaker et al., 2013). However, none of these studies has evaluated microglial activation longitudinally using PET and selected subjects at different stages of the disease. This dual peak in a heterogeneous population could account for some of the differences in results of microglial activation in different studies. Interestingly, studies evaluating anti-inflammatory agents in Alzheimer's disease have also produced conflicting results. One could speculate that if any anti-microglial agents are to be effective, they have to specifically target the pro-inflammatory phenotype, and hence the inconsistencies in the results.

BPM correlation analysis revealed a positive correlation between microglial activation and amyloid deposition. At follow-up, more clusters of positive correlation between microglial activation and amyloid were seen in temporoparietal lobe which could imply that, as amyloid increases, local microglial activation is also increased and one could speculate that, as amyloid increases, microglia are trying to clear the amyloid. Microglia are key innate immune cells and it is possible that, in the process of removing the cell debris, the toxic products accumulate and damage the neurons in the vicinity (Doens and Fernandez, 2014; Mandrekar-Colucci and Landreth, 2010). In Alzheimer's disease brain, it has been shown that plaques cause direct activation of the complement system by binding to the C1q protein in pyramidal neurons, which is significantly increased. Thus, in agreement with our previous findings, it is possible that, due to continuous toxic insult, microglia lose their beneficial property and become toxically over-activated perhaps by the unexpected complement system induced by C1q protein, which may lead to further damage to the neurons, cognitive impairment and worsening of Alzheimer's disease (Fraser et al., 2010).

While we noticed a voxel-wise positive correlation between microglial activation and amyloid deposition, we found the level of microglial activation in subjects with mild cognitive impairment decreased over time. In spite of the regional reduction in microglial activation, more clusters of positive association between amyloid and microglial activation were observed at the voxel level during mild cognitive impairment follow-up, suggesting more microglia are specifically targeting amyloid. However, there was a group-wise deterioration of cognitive function suggesting that, as disease advances, amyloid deposition and microglial activation could play a synergistic role in accelerated neurodegeneration. This suggests that, rather than a simple direct relationship, it is more likely to imply a complex relationship between microglial activation and amyloid, potentially due to different functional microglial activation in different stages of disease.



This is a study with small numbers and cohorts followed longitudinally at different time points. A larger sample size with longer longitudinal follow-up is necessary to confirm or refute the bimodal relationship between microglial activation and amyloid that we are postulating. [11C](R)PK11195 PET provides a relatively low signal-to-noise ratio across varying neurodegeneration. Currently, our laboratory and others have been evaluating second-generation TSPO ligands ([11C]PBR28, [11C]DPA713, and [18F]GE180) with the aim of achieving better signal-to-noise ratio and discrimination of active disease states from health. However, these newer ligands are influenced by TSPO genotype and, to date, they have not been proven to be superior to [11C](R)PK11195 PET for discriminating Alzheimer's disease from healthy controls. The future goal is to confirm the pathological relationship between amyloid aggregation and microglial activation and to determine whether anti-microglial agents show efficacy in suppressing cell activity.

In conclusion, this is one of the first longitudinal PET studies evaluating microglial activation changes associated with mild cognitive impairment and Alzheimer's disease progression, and correlating these with levels of amyloid deposition. It has revealed that, while inflammation is initially present in mild cognitive impairment, it diminishes over 14 months but then subsequently rises as progressing to Alzheimer's disease. This supports the view that microglial activation is a dynamic process, and are probably undergoing constant transformation between phenotypes. We speculate that activated microglia in subjects with mild cognitive impairment initially adopt a protective phenotype but change to a pro-inflammatory phenotype as disease progresses and amyloid clearance fails. This results in two peaks of microglial activation and suggests that anti-microglial agents may have their most beneficial effect in later stages of the disease when they target the pro-inflammatory phenotypes.

## **Acknowledgement**

The authors thank Hammersmith Imanet, GE Healthcare, for the provision of radiotracers, scanning, and blood analysis equipment. Dr Edison was funded by the Medical Research Council and now by Higher Education Funding Council for England (HEFCE). The PET scans and MRI scans were funded by the Medical Research Council and Alzheimer's Research UK. This article presents independent research funded by Medical Research Council and Alzheimer's Research, UK and supported by the NIHR CRF and BRC at Imperial College Healthcare NHS Trust.

## **Funding**

The PET scans and MRI scans were funded by the Medical Research Council and part of the study was funded by Alzheimer's Research UK.

Dr. Edison was funded by the Medical Research Council and now by Higher Education Funding Council for England (HEFCE). He has also received grants from Alzheimer's Research, UK, Alzheimer's Drug Discovery Foundation, Alzheimer's Society, UK, Novo Nordisk and GE Healthcare. Prof. Brooks has received research grants and non-financial support from the Medical Research Council, grants from Alzheimer's Research Trust, during the conduct of the study; other from GE Healthcare, personal fees from AstraZeneca, personal fees from Cytos, personal fees from Shire, personal fees from Novartis, personal fees from GSK, Holland, personal fees from Navidea, personal fees from UCB, personal fees from Acadia, grants from Michael J Fox Foundation, grants from European Commission, outside the submitted work. Dr Okello and Miss Fan have nothing to disclose.

## **Supplementary material**

Supplementary material is available at Brain online.

## References

- Amor, S., Puentes, F., Baker, D., van der Valk, P., Inflammation in neurodegenerative diseases. *Immunology*. 2010; 129:2, 154-69.
- Anderson, A. N., Pavese, N., Edison, P., Tai, Y. F., Hammers, A., Gerhard, A., et al., A systematic comparison of kinetic modelling methods generating parametric maps for [(11)C]-(R)-PK11195. *Neuroimage*. 2007; 36:1, 28-37.
- Cai, Z., Hussain, M. D., Yan, L. J., Microglia, neuroinflammation, and beta-amyloid protein in Alzheimer's disease. *Int J Neurosci*. 2013.
- Casanova, R., Srikanth, R., Baer, A., Laurienti, P. J., Burdette, J. H., Hayasaka, S., et al., Biological parametric mapping: A statistical toolbox for multimodality brain image analysis. *Neuroimage*. 2007; 34:1, 137-43.
- Craig-Schapiro, R., Perrin, R. J., Roe, C. M., Xiong, C., Carter, D., Cairns, N. J., et al., YKL-40: a novel prognostic fluid biomarker for preclinical Alzheimer's disease. *Biol Psychiatry*. 2010; 68:10, 903-12.
- Cunningham, C., Microglia and neurodegeneration: the role of systemic inflammation. *Glia*. 2013; 61:1, 71-90.
- Doens, D., Fernandez, P. L., Microglia receptors and their implications in the response to amyloid beta for Alzheimer's disease pathogenesis. *Journal of Neuroinflammation*. 2014; 11.
- Edison, P., Ahmed, I., Fan, Z., Hinz, R., Gelosa, G., Ray Chaudhuri, K., et al., Microglia, amyloid, and glucose metabolism in Parkinson's disease with and without dementia. *Neuropsychopharmacology*. 2013; 38:6, 938-49.
- Edison, P., Archer, H. A., Gerhard, A., Hinz, R., Pavese, N., Turkheimer, F. E., et al., Microglia, amyloid, and cognition in Alzheimer's disease: An [11C](R)PK11195-PET and [11C]PIB-PET study. *Neurobiol Dis*. 2008; 32:3, 412-9.
- Fan, Z., Aman, Y., Ahmed, I., Chetelat, G., Landeau, B., Ray Chaudhuri, K., et al., Influence of microglial activation on neuronal function in Alzheimer's and Parkinson's disease dementia. *Alzheimers Dement*. 2015a; 11:6, 608-21 e7.
- Fan, Z., Okello, A. A., Brooks, D. J., Edison, P., Longitudinal influence of microglial activation and amyloid on neuronal function in Alzheimer's disease. *Brain*. 2015b; 138:Pt 12, 3685-98.
- Fraser, D. A., Pisalyaput, K., Tenner, A. J., C1q enhances microglial clearance of apoptotic neurons and neuronal blebs, and modulates subsequent inflammatory cytokine production. *J Neurochem*. 2010; 112:3, 733-43.
- Guerreiro, R., Hardy, J., TREM2 and neurodegenerative disease. *N Engl J Med*. 2013; 369:16, 1569-70.
- Guerreiro, R. J., Hardy, J., Alzheimer's disease genetics: lessons to improve disease modelling. *Biochem Soc Trans*. 2011; 39:4, 910-6.
- Hamelin, L., Lagarde, J., Dorothee, G., Leroy, C., Labit, M., Comley, R. A., et al., Early and protective microglial activation in Alzheimer's disease: a prospective study using 18F-DPA-714 PET imaging. *Brain*. 2016; 139:Pt 4, 1252-64.
- Heneka, M. T., Carson, M. J., El Khoury, J., Landreth, G. E., Brosseron, F., Feinstein, D. L., et al., Neuroinflammation in Alzheimer's disease. *Lancet Neurol*. 2015; 14:4, 388-405.
- Heslegrave, A., Heywood, W., Paterson, R., Magdalinou, N., Svensson, J., Johansson, P., et al., Increased cerebrospinal fluid soluble TREM2 concentration in Alzheimer's disease. *Molecular Neurodegeneration*. 2016; 11.
- Hollingsworth, P., Harold, D., Jones, L., Owen, M. J., Williams, J., Alzheimer's disease genetics: current knowledge and future challenges. *Int J Geriatr Psychiatry*. 2011a; 26:8, 793-802.
- Hollingsworth, P., Harold, D., Sims, R., Gerrish, A., Lambert, J. C., Carrasquillo, M. M., et al., Common variants at ABCA7, MS4A6A/MS4A4E, EPHA1, CD33 and CD2AP are associated with Alzheimer's disease. *Nat Genet*. 2011b; 43:5, 429-35.

- Jack, C. R., Jr., Knopman, D. S., Jagust, W. J., Shaw, L. M., Aisen, P. S., Weiner, M. W., et al., Hypothetical model of dynamic biomarkers of the Alzheimer's pathological cascade. *Lancet Neurol.* 2010; 9:1, 119-28.
- Jaturapatporn, D., Isaac, M. G., McCleery, J., Tabet, N., Aspirin, steroidal and non-steroidal anti-inflammatory drugs for the treatment of Alzheimer's disease. *Cochrane Database Syst Rev.* 2012; 2, CD006378.
- Jiang, T., Yu, J. T., Zhu, X. C., Tan, L., TREM2 in Alzheimer's disease. *Molecular Neurobiology.* 2013; 48:1, 180-185.
- Lim, Y. Y., Maruff, P., Pietrzak, R. H., Ames, D., Ellis, K. A., Harrington, K., et al., Effect of amyloid on memory and non-memory decline from preclinical to clinical Alzheimer's disease. *Brain.* 2014; 137, 221-231.
- Lyman, M., Lloyd, D. G., Ji, X., Vizcaychipi, M. P., Ma, D., Neuroinflammation: the role and consequences. *Neurosci Res.* 2014; 79, 1-12.
- Lynch, M. A., The impact of neuroimmune changes on development of amyloid pathology; relevance to Alzheimer's disease. *Immunology.* 2013.
- Mandrekar-Colucci, S., Landreth, G. E., Microglia and Inflammation in Alzheimer's Disease. *Cns & Neurological Disorders-Drug Targets.* 2010; 9:2, 156-167.
- Okello, A., Koivunen, J., Edison, P., Archer, H. A., Turkheimer, F. E., Nagren, K., et al., Conversion of amyloid positive and negative MCI to AD over 3 years: an 11C-PIB PET study. *Neurology.* 2009; 73:10, 754-60.
- Pan, X. D., Zhu, Y. G., Lin, N., Zhang, J., Ye, Q. Y., Huang, H. P., et al., Microglial phagocytosis induced by fibrillar beta-amyloid is attenuated by oligomeric beta-amyloid: implications for Alzheimer's disease. *Mol Neurodegener.* 2011; 6, 45.
- Piccio, L., Deming, Y., Del-Águila, J. L., Ghezzi, L., Holtzman, D. M., Fagan, A. M., et al., Cerebrospinal fluid soluble TREM2 is higher in Alzheimer disease and associated with mutation status. *Acta neuropathologica.* 2016, 1-9.
- Prokop, S., Miller, K. R., Heppner, F. L., Microglia actions in Alzheimer's disease. *Acta Neuropathol.* 2013.
- Rogers, J., Strohmeier, R., Kovelowski, C. J., Li, R., Microglia and inflammatory mechanisms in the clearance of amyloid beta peptide. *Glia.* 2002; 40:2, 260-269.
- Sanchez-Guajardo, V., Barnum, C. J., Tansey, M. G., Romero-Ramos, M., Neuroimmunological processes in Parkinson's disease and their relation to alpha-synuclein: microglia as the referee between neuronal processes and peripheral immunity. *ASN Neuro.* 2013; 5:2, 113-39.
- Sastre, M., Klockgether, T., Heneka, M. T., Contribution of inflammatory processes to Alzheimer's disease: molecular mechanisms. *Int J Dev Neurosci.* 2006; 24:2-3, 167-76.
- Schuitmaker, A., Kropholler, M. A., Boellaard, R., van der Flier, W. M., Kloet, R. W., van der Doef, T. F., et al., Microglial activation in Alzheimer's disease: an (R)-[(1)(1)C]PK11195 positron emission tomography study. *Neurobiol Aging.* 2013; 34:1, 128-36.
- Shidahara, M., Tsoumpas, C., Hammers, A., Boussion, N., Visvikis, D., Suhara, T., et al., Functional and structural synergy for resolution recovery and partial volume correction in brain PET. *Neuroimage.* 2009; 44:2, 340-8.
- Suárez - Calvet, M., Kleinberger, G., Caballero, M. Á. A., Brendel, M., Rominger, A., Alcolea, D., et al., sTREM2 cerebrospinal fluid levels are a potential biomarker for microglia activity in early - stage Alzheimer's disease and associate with neuronal injury markers. *EMBO molecular medicine.* 2016, e201506123.
- Turkheimer, F. E., Edison, P., Pavese, N., Roncaroli, F., Anderson, A. N., Hammers, A., et al., Reference and target region modeling of [11C]-(R)-PK11195 brain studies. *J Nucl Med.* 2007; 48:1, 158-67.
- Turkheimer, F. E., Smith, C. B., Schmidt, K., Estimation of the number of "true" null hypotheses in multivariate analysis of neuroimaging data. *Neuroimage.* 2001; 13:5, 920-930.

- Varley, J., Brooks, D. J., Edison, P., Imaging neuroinflammation in Alzheimer's disease and other dementias: Recent advances and future directions. *Alzheimers Dement.* 2015; 11:9, 1110-20.
- Varnum, M. M., Ikezu, T., The classification of microglial activation phenotypes on neurodegeneration and regeneration in Alzheimer's disease brain. *Arch Immunol Ther Exp (Warsz).* 2012; 60:4, 251-66.
- Villemagne, V. L., Burnham, S., Bourgeat, P., Brown, B., Ellis, K. A., Salvado, O., et al., Amyloid beta deposition, neurodegeneration, and cognitive decline in sporadic Alzheimer's disease: a prospective cohort study. *Lancet Neurology.* 2013; 12:4, 357-367.
- von Bernhardi, R., Eugenin-von Bernhardi, L., Eugenin, J., Microglial cell dysregulation in brain aging and neurodegeneration. *Front Aging Neurosci.* 2015; 7, 124.
- Yasuno, F., Kosaka, J., Ota, M., Higuchi, M., Ito, H., Fujimura, Y., et al., Increased binding of peripheral benzodiazepine receptor in mild cognitive impairment-dementia converters measured by positron emission tomography with [(1)(1)C]DAA1106. *Psychiatry Res.* 2012; 203:1, 67-74.
- Zotova, E., Bharambe, V., Cheaveau, M., Morgan, W., Holmes, C., Harris, S., et al., Inflammatory components in human Alzheimer's disease and after active amyloid-beta42 immunization. *Brain.* 2013; 136:Pt 9, 2677-96.

## Figure Legends

### **Figure 1. SPM analysis between baseline and follow up microglial activation in mild cognitive impairment subjects.**

Figure 1(A) Paired t-test SPM analysis between baseline and follow up microglial activation in all subjects with mild cognitive impairment. Figure 2(B) Paired t-test SPM analysis between baseline and follow up microglial activation in amyloid positive subjects. Figure 1 (C) Paired t-test SPM analysis between baseline and follow up microglial activation in amyloid negative subjects. Supplementary Table 2 details the coordinates with their statistical results. MCI = subjects with mild cognitive impairment.

### **Figure 2. Longitudinal changes in [11C](R)PK11195 BP and [11C]PIB in subjects with mild cognitive impairment compared to controls at baseline and follow-up.**

Figure 2(A) and (B) demonstrate clusters of significantly increased [11C](R)PK11195 BP in subjects with mild cognitive impairment compared to healthy controls at baseline and follow-up, respectively, with the same voxel threshold of  $P < 0.01$  and extent threshold of 50 voxels. Figure 2(C) and (D) show clusters of significantly increased [11C]PIB in subjects with mild cognitive impairment compared to healthy controls at baseline and follow-up ( $p < 0.0001$  and extent of 200 voxels). Supplementary Table 3 details the coordinates with their statistical results. Figure 2(E) and (F) demonstrates 3D intensity T-map of [11C](R)PK11195 BP and [11C]PIB RATIO for two individual subjects with mild cognitive impairment at baseline and follow-up. The surface plot represents significant increase in the microglial activation (left panel) and amyloid deposition (right panel) against the control group at baseline superimposed on a 3D matrix at baseline and follow-up.

### **Figure 3. Voxel-based correlation between [11C](R)PK11195 BP and [11C]PIB uptake.**

Figure 3 demonstrates BPM positive correlation between microglial activation and amyloid deposition superimposed on a SPM render brain image, along with 3D intensity T-map of BPM

correlation in sagittal view in mild cognitive impairment and Alzheimer's disease cohort. Figure 3(A) and (B) show positive correlations between [11C]PIB RATIO and [11C](R)PK11195 BP in subjects with mild cognitive impairment at baseline and follow-up respectively at a cluster threshold of  $p < 0.05$  with an extent threshold of 50-voxel. Figure 3 (C) and (D) show positive correlations between [11C]PIB RATIO and [11C](R)PK11195 BP in subjects with Alzheimer's disease at baseline and follow-up respectively at a cluster threshold of  $p < 0.05$  with an extent threshold of 50-voxel. Supplementary Table 4 details the significant clusters.

**Figure 4. Hypothetical model of dual peak of microglial activation in the Alzheimer's trajectory.** The upper panel demonstrates the hypothetical model of morphological changes in microglia in Alzheimer's disease trajectory, where ramified microglia transform to anti-inflammatory (protective) microglial phenotype and pro-inflammatory (toxic) microglial phenotypes. The lower panel shows the microglial activation in relation to other biomarkers detectable using positron emission tomography where two peaks of microglial activation are present in Alzheimer's trajectory (modified from Jack et al) (Jack et al., 2010).

**Supplementary Figure 1. SPM analysis between patients and healthy controls using a cluster threshold of  $p < 0.05$  with 50 voxels.**

Supplementary Figure 1(A) and (B) demonstrate clusters of significantly increased [11C](R)PK11195 BP in MCI subjects compared to healthy controls at baseline and follow-up with a voxel threshold of  $P < 0.05$  and extent threshold of 50 voxels. Supplementary Figure 1 (C) and (D) show clusters of significantly increased [11C]PIB uptake in subjects with mild cognitive impairment subjects compared to healthy controls at baseline and follow-up ( $p < 0.05$  and extent of 50 voxels).

**Supplementary Figure 2. Pixel-wise correlation (scatter plot) between group mean [11C]PIB uptake and [11C](R)PK11195 binding potential.**

Supplementary Figure 2 (A) demonstrates the pixel-wise correlation for all subjects with mild cognitive impairment between group mean [11C]PIB and [11C](R)PK11195 binding potential. The top scatter plot represents a significant Pearson correlation ( $r=0.37$ ,  $p<0.0001$ ) in the whole cortex between group mean [11C]PIB uptake and [11C](R)PK11195 binding potential. The lower four scatter plots show the correlation in frontal (blue), temporal (purple), parietal (orange), and occipital (yellow) cortices. Supplementary Figure 2 (B) demonstrates the pixel-wise correlation for amyloid positive (Left) and amyloid negative subjects (Right) with mild cognitive impairment. The scatter plot in the top panel represents Pearson correlation in the whole cortex between group mean [11C]PIB uptake and [11C](R)PK11195 binding potential in amyloid positive subjects ( $r=0.53$ ,  $p<0.0001$ ) and amyloid negative subjects ( $r=0.0065$ ,  $p=0.09$ ). The lower four scatter plots show the correlation in frontal (blue), temporal (purple), parietal (orange), and occipital (yellow) cortices. MCI = mild cognitive impairment.



**Figure 1**

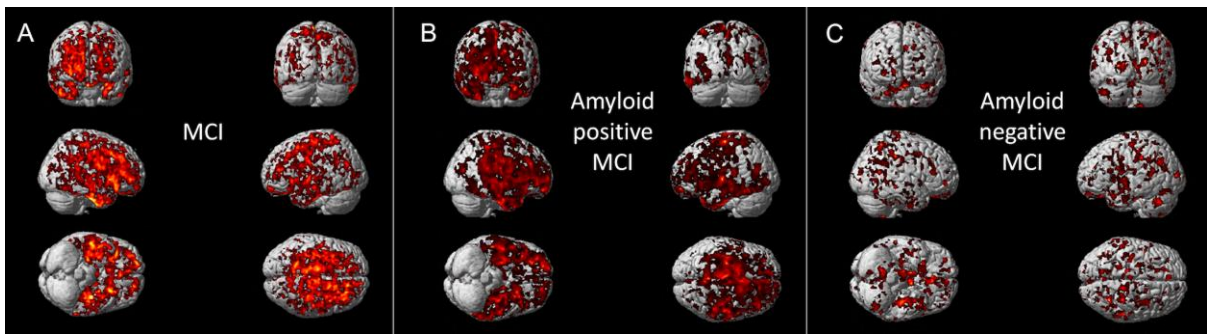


Figure 2

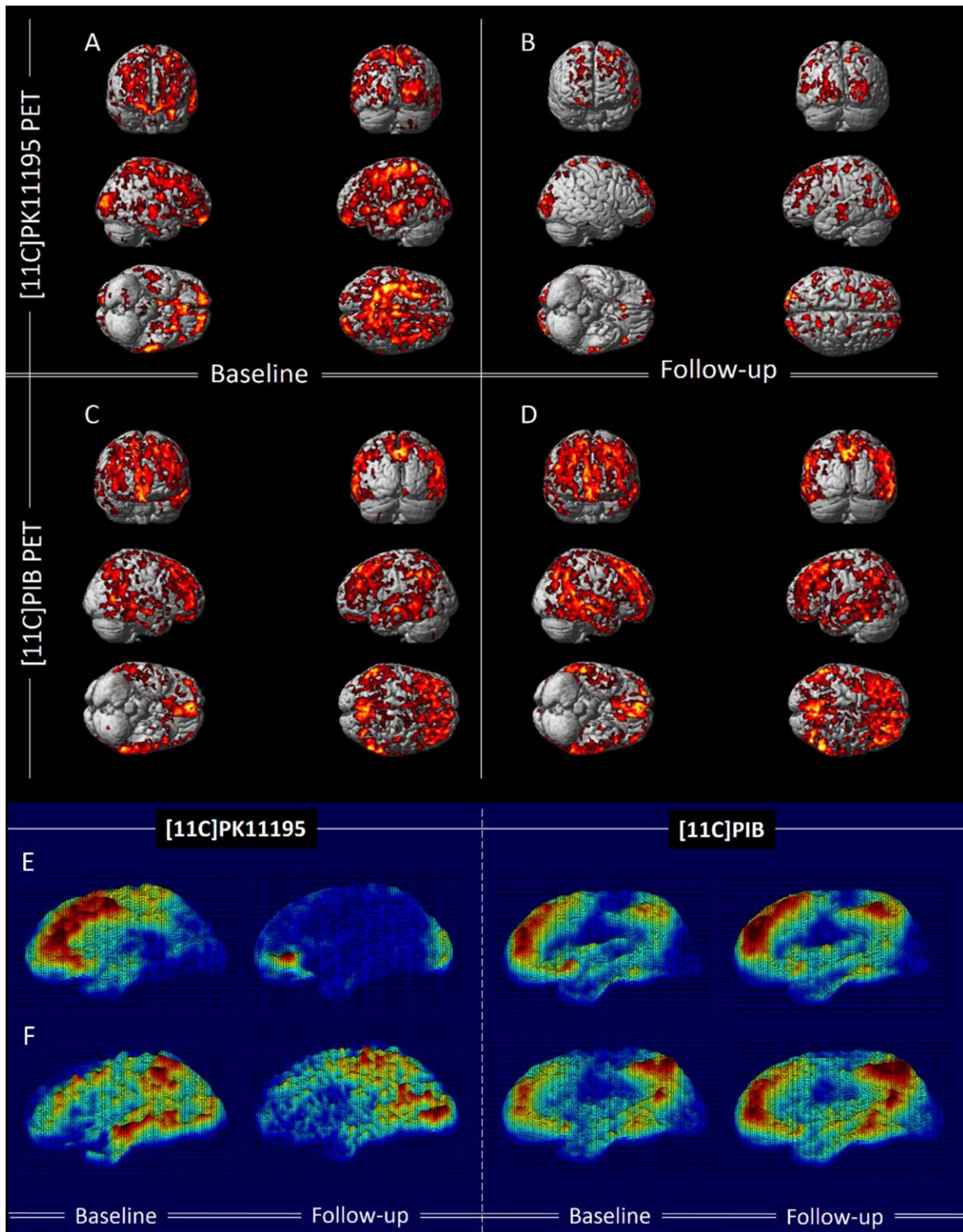
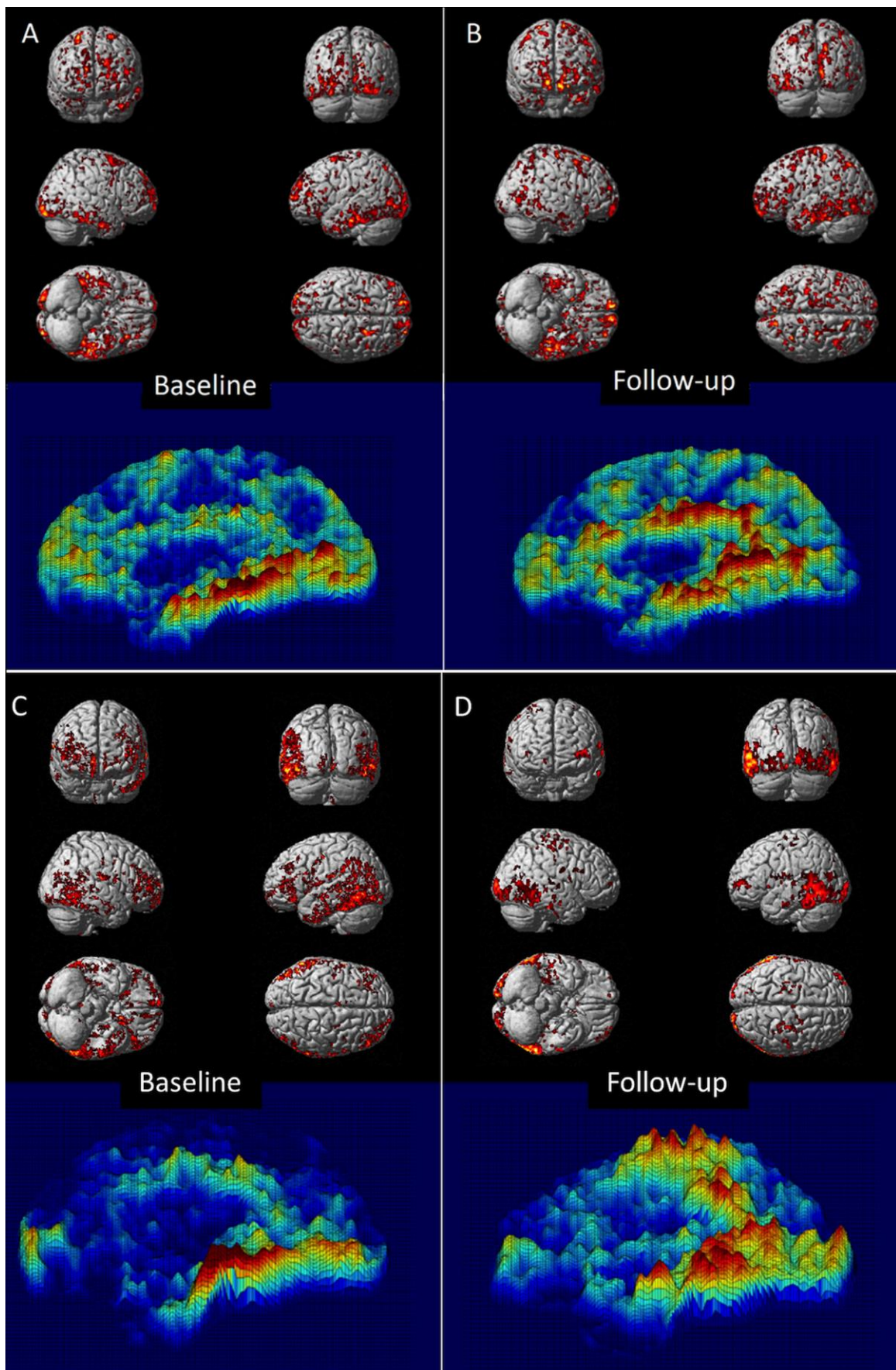
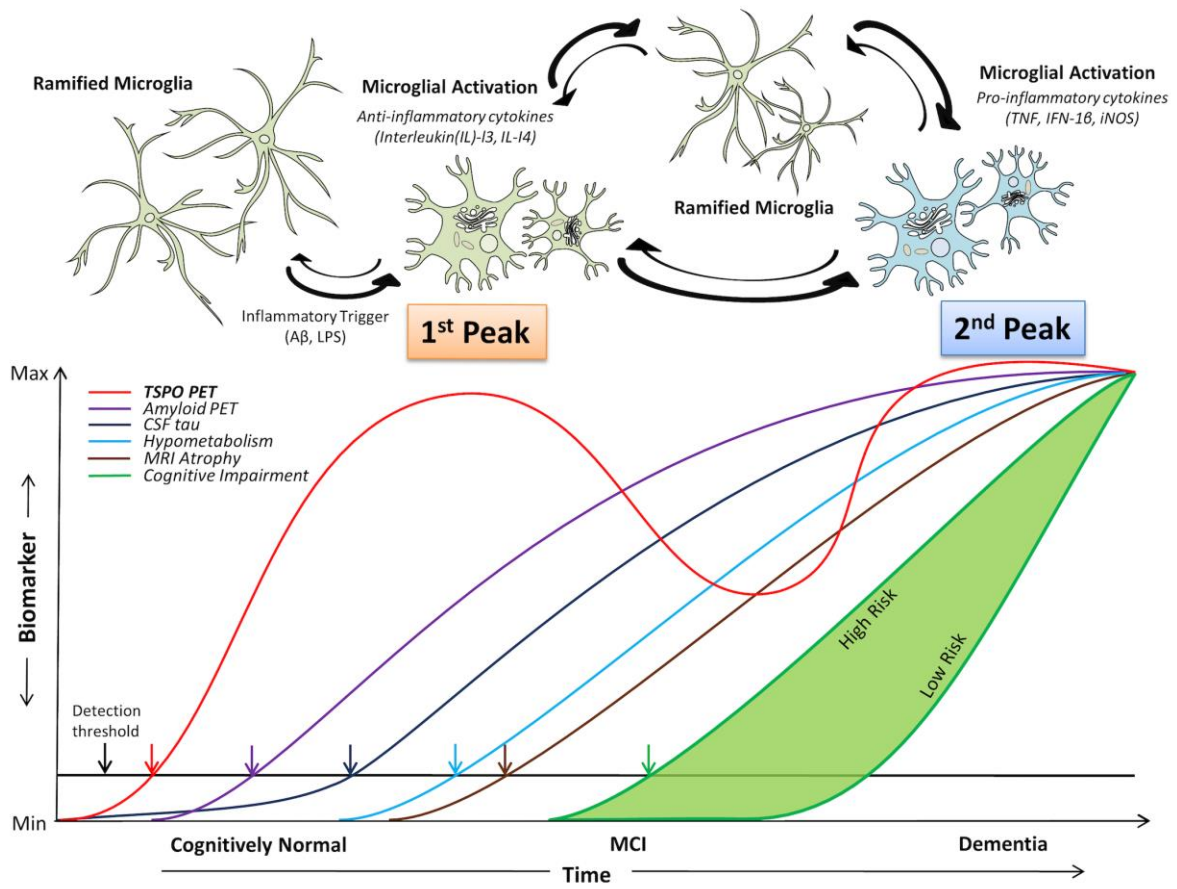


Figure 3



**Figure 4**



**Table 1: Demographic details of subjects with mild cognitive impairment and control subjects**

	MCI	HC [11C]PIB	HC [11C](R)PK11195
N	8	14	8
Mean age (years $\pm$ sd)	67.7 $\pm$ 6.6	64.4 $\pm$ 5.9	65.5 $\pm$ 5.5
Age range	55-77	54-75	58-71
Gender (female/total)	4/8	4/14	4/8
MMSE (mean $\pm$ sd)	27.6 $\pm$ 1.2	30	30
Amyloid load (A $\beta$ +/total)	4/8	/	/

SD = standard deviation; MMSE = mini-mental state exam; MCI = subjects with mild cognitive impairment; HC = healthy control.

**Table 2A: Regional [11C](R)PK11195 BP and [11C]PIB RATIO in subjects with mild cognitive impairment and healthy control (HC) subjects at baseline and follow-up.**

<i>MCI subjects</i>		<i>Frontal</i>		<i>Temporal</i>		<i>Parietal</i>		<i>Occipital</i>	
		<b>PK11195</b>	<b>PIB</b>	<b>PK11195</b>	<b>PIB</b>	<b>PK11195</b>	<b>PIB</b>	<b>PK11195</b>	<b>PIB</b>
<b><i>Baseline</i></b>	Mean	0.42	1.53	0.47	1.54	0.43	1.56	0.48	1.47
	SD	0.1	0.58	0.09	0.54	0.08	0.59	0.09	0.49
	Increase (B vs. HC) <sup>1</sup>	42%	32%	44%	34%	47%	34%	35%	21%
	P value	0.0079*	0.0135*	0.0021*	0.0060*	0.0080*	0.0102*	0.0071*	0.0325*
<b><i>Follow-up</i></b>	Mean	0.36	1.65	0.39	1.62	0.37	1.64	0.42	1.54
	SD	0.08	0.64	0.03	0.6	0.04	0.67	0.03	0.52
	Increase (F vs. HC) <sup>1</sup>	20%	43%	18%	41%	26%	41%	18%	27%
	P value	0.114	0.0054*	0.0430*	0.0037*	0.0500*	0.0073*	0.0347*	0.0144*
<b><i>HC</i></b>	Mean	0.3	1.16	0.33	1.15	0.29	1.16	0.36	1.21
	SD	0.1	0.07	0.08	0.05	0.12	0.06	0.09	0.06
<b><i>B vs. F</i></b>	Longitudinal change <sup>2</sup>	<b>-14%</b>	<b>8%</b>	<b>-16%</b>	<b>5%</b>	<b>-13%</b>	<b>5%</b>	<b>-11%</b>	<b>5%</b>
	Paired t test	0.0265	0.0157*	0.0088*	0.0101*	0.0149*	0.0267	0.0204*	0.0032*

<i>MCI subjects</i>		<i>A-cing</i>		<i>P-cing</i>		<i>Thalamus</i>		<i>Striatum</i>		<i>Hippo</i>	
		<b>PK11195</b>	<b>PIB</b>	<b>PK11195</b>	<b>PIB</b>	<b>PK11195</b>	<b>PIB</b>	<b>PK11195</b>	<b>PIB</b>	<b>PK11195</b>	<b>PIB</b>
<b><i>Baseline</i></b>	Mean	0.53	1.69	0.57	1.73	0.43	1.26	0.35	1.59	0.48	1.35
	SD	0.14	0.62	0.11	0.69	0.07	0.33	0.07	0.47	0.11	0.13
	Increase (B vs. HC) <sup>1</sup>	59%	36%	46%	41%	12%	24%	36%	32%	41%	11%
	P value	0.0024*	0.0069*	0.0021*	0.0056*	0.2013	0.0086*	0.0066*	0.0030*	0.0087*	0.0031*
<b><i>Follow-up</i></b>	Mean	0.36	1.77	0.43	1.85	0.41	1.28	0.33	1.66	0.38	1.38
	SD	0.09	0.71	0.1	0.77	0.09	0.33	0.06	0.54	0.08	0.18
	Increase (F vs. HC) <sup>1</sup>	7%	43%	10%	51%	7%	26%	26%	38%	11%	17%
	P value	0.3113	0.0056*	0.2202	0.0030*	0.3323	0.005*	0.0221*	0.0024*	0.2047	0.0111*
<b><i>HC</i></b>	Mean	0.33	1.24	0.39	1.23	0.38	1.02	0.26	1.2	0.34	1.18
	SD	0.09	0.11	0.1	0.05	0.14	0.09	0.06	0.07	0.09	0.11
<b><i>B vs. F</i></b>	Longitudinal change <sup>2</sup>	<b>-29%</b>	<b>5%</b>	<b>-23%</b>	<b>7%</b>	<b>-2%</b>	<b>1%</b>	<b>-6%</b>	<b>4%</b>	<b>-16%</b>	<b>3%</b>
	Paired t test	0.0079*	0.0823	0.0062*	0.0116*	0.3243	0.3336	0.1445	0.0363	0.0248*	0.0692

\*Significant p <0.05 with correction for multiple comparisons. Increase (B vs. HC)<sup>1</sup> indicates the group-wise ROI comparison between baseline MCI patients and healthy controls, while Increase (F vs. HC)<sup>1</sup> indicates the group-wise ROI comparison between follow-up MCI patients and



healthy controls; Longitudinal change<sup>2</sup> indicates the group-wise ROI comparison between baseline and follow-up. HC = Healthy controls; B vs. F = Baseline vs. Follow-up; A-Cing = Anterior Cingulate; P-Cing = Posterior Cingulate; Hippo = Hippocampus; Frontal = Frontal Lobe; Temporal = Temporal Lobe; Parietal = Parietal Lobe; Occipital = Occipital Lobe; PK11195 = [11C](R)PK11195; PIB = [11C]PIB.

**Table 2B: Regional [11C](R)PK11195 BP in amyloid positive MCI subjects (A $\beta$ + MCI) and amyloid negative MCI subjects (A $\beta$ - MCI) at baseline and during follow-up.**

<i>MCI subjects (A<math>\beta</math>+ &amp; A<math>\beta</math>-)</i>	<i>Frontal</i>		<i>Temporal</i>		<i>Parietal</i>		<i>Occipital</i>		
	<b>A<math>\beta</math>+ MCI</b>	<b>A<math>\beta</math>- MCI</b>	<b>A<math>\beta</math>+ MCI</b>	<b>A<math>\beta</math>- MCI</b>	<b>A<math>\beta</math>+ MCI</b>	<b>A<math>\beta</math>- MCI</b>	<b>A<math>\beta</math>+ MCI</b>	<b>A<math>\beta</math>- MCI</b>	
<b><i>Baseline</i></b>	Mean	0.44	0.41	0.52	0.42	0.43	0.44	0.52	0.45
	SD	0.06	0.07	0.07	0.07	0.08	0.07	0.08	0.08
	Increase (B vs. HC) <sup>1</sup>	47%	36%	60%	29%	46%	49%	44%	25%
	P value	0.0110*	0.0601	0.0025*	0.0021*	0.0333*	0.0396*	0.0161*	0.0029*
<b><i>Follow-up</i></b>	Mean	0.36	0.36	0.39	0.38	0.37	0.36	0.43	0.42
	SD	0.09	0.06	0.03	0.05	0.05	0.03	0.04	0.03

	Increase (F vs. HC) <sup>1</sup>	19%	21%	19%	17%	27%	24%	18%	17%
	P value	0.2019	0.1306	0.0383*	0.1078	0.0757	0.0725	0.0527	0.0504
<b><i>B vs. F</i></b>	Longitudinal change <sup>2</sup>	<b>-19%</b>	<b>-11%</b>	<b>-25%</b>	<b>-9%</b>	<b>-13%</b>	<b>-16%</b>	<b>-18%</b>	<b>-6%</b>
	Paired t test	0.0542	0.1900	0.0062*	0.1969	0.0389	0.1043	0.0274	0.2291

<b><i>MCI subjects (Aβ+ &amp; Aβ-)</i></b>		<b><i>A-cing</i></b>		<b><i>P-cing</i></b>		<b><i>Thalamus</i></b>		<b><i>Striatum</i></b>		<b><i>Hippo</i></b>	
		<b>Aβ+</b>	<b>Aβ-</b>	<b>Aβ+</b>	<b>Aβ-</b>	<b>Aβ+ MCI</b>	<b>Aβ-</b>	<b>Aβ+</b>	<b>Aβ-</b>	<b>Aβ+ MCI</b>	<b>Aβ-</b>
		<b>MCI</b>	<b>MCI</b>	<b>MCI</b>	<b>MCI</b>		<b>MCI</b>	<b>MCI</b>	<b>MCI</b>		<b>MCI</b>
<b><i>Baseline</i></b>	Mean	0.55	0.50	0.61	0.53	0.43	0.43	0.35	0.36	0.53	0.43
	SD	0.16	0.07	0.10	0.09	0.07	0.06	0.08	0.04	0.11	0.08

<i>Follow-up</i>	Increase (B vs. HC) <sup>1</sup>	67%	51%	57%	35%	13%	12%	35%	37%	55%	27%
	P value	0.0474*	0.0364*	0.0117*	0.0684	0.2275	0.0105*	0.0693	0.0313*	0.0270*	0.0515
	Mean	0.32	0.39	0.42	0.44	0.43	0.39	0.30	0.35	0.35	0.43
	SD	0.08	0.07	0.08	0.10	0.08	0.09	0.04	0.05	0.06	0.05
	Increase (F vs. HC) <sup>1</sup>	-3%	17%	6%	13%	11%	3%	16%	37%	4%	25%
	P value	0.4218	0.1609	0.3408	0.2313	0.2775	0.4435	0.1350	0.0180*	0.3958	0.0472*
<i>B vs. F</i>	Longitudinal change <sup>2</sup>	<b>-42%</b>	<b>-22%</b>	<b>-32%</b>	<b>-16%</b>	<b>-2%</b>	<b>-8%</b>	<b>-14%</b>	<b>-1%</b>	<b>-33%</b>	<b>-2%</b>
	Paired t test	0.0418	0.0681	0.0165*	0.1227	0.4521	0.3406	0.0508	0.4823	0.0117*	0.3852

\*Significant  $p < 0.05$  with correction for multiple comparisons. Increase (B vs. HC)<sup>1</sup> indicates the group-wise ROI comparison between baseline MCI patients and healthy controls, while Increase (F vs. HC)<sup>1</sup> indicates the group-wise ROI comparison between follow-up MCI patients and healthy controls; Longitudinal change<sup>2</sup> indicates the group-wise ROI comparison between baseline and follow-up. B vs. F = Baseline vs. Follow-up; A-Cing = Anterior Cingulate; P-Cing = Posterior Cingulate; Hippo = Hippocampus; Frontal = Frontal Lobe; Temporal = Temporal Lobe; Parietal = Parietal Lobe; Occipital = Occipital Lobe; A $\beta$ + MCI = Amyloid positive MCI; A $\beta$ - MCI = Amyloid negative MCI.

**Table 3: Partial volume corrected (PVC) regional [11C](R)PK11195 in subjects with mild cognitive impairment at baseline and follow-up**

PVC [11C](R)PK11195 BP <sup>1</sup>		<i>Frontal</i>	<i>Temporal</i>	<i>Parietal</i>	<i>Occipital</i>	<i>A-cing</i>	<i>P-cing</i>	<i>Thalamus</i>	<i>Striatum</i>	<i>Hippo</i>
<i>Baseline</i>	Mean	0.48	0.44	0.47	0.48	0.54	0.56	0.48	0.38	0.44
	SD	0.10	0.09	0.06	0.09	0.14	0.12	0.06	0.06	0.15
<i>Follow-up</i>	Mean	0.41	0.38	0.39	0.45	0.32	0.39	0.43	0.35	0.35
	SD	0.08	0.03	0.05	0.06	0.10	0.11	0.10	0.04	0.05
<i>B vs. F</i>	Longitudinal change <sup>2</sup>	<b>-15%</b>	<b>-15%</b>	<b>-17%</b>	<b>-7%</b>	<b>-41%</b>	<b>-31%</b>	<b>-9%</b>	<b>-8%</b>	<b>-21%</b>
	Paired t-test	0.0389*	0.0306*	0.0082*	0.1926	0.0085*	0.0150*	0.1480	0.0819	0.0475*

\*p<0.05; PVC [11C](R)PK11195 PVC BP<sup>1</sup> = [11C](R)PK11195 partial volume corrected binding potential; B vs. F = Baseline vs. Follow-up; Longitudinal change<sup>2</sup> indicates the group-wise ROI comparison between baseline and follow-up PVC [11C](R)PK11195 BP; A-Cing = Anterior Cingulate; P-Cing = Posterior Cingulate; Hippo = Hippocampus; Frontal = Frontal Lobe; Temporal = Temporal Lobe; Parietal = Parietal Lobe; Occipital = Occipital Lobe.

**Table 4. Changes in extent of area of microglial activation and the extent of area of amyloid deposition in subjects with Alzheimer’s disease and subjects with mild cognitive impairment between baseline and follow-up.**

		<i>MCI1</i> <i>(Aβ+)</i>	<i>MCI2</i> <i>(Aβ+)</i>	<i>MCI3</i> <i>(Aβ-)</i>	<i>MCI4</i> <i>(Aβ-)</i>	<i>MCI5</i> <i>(Aβ+)</i>	<i>MCI6</i> <i>(Aβ-)</i>	<i>MCI7</i> <i>(Aβ+)</i>	<i>MCI8</i> <i>(Aβ-)</i>
<i>[11C](R)PK11195</i> <i>volume</i>	<b>Baseline</b>	307584	68527	8894	136917	94941	230360	450468	61452
	<b>Follow-up</b>	173297	55552	33710	56623	74585	30253	124229	138047
	<b>Changes</b>	<i>-134287</i>	<i>-12975</i>	<i>24816</i>	<i>-80294</i>	<i>-20356</i>	<i>-200107</i>	<i>-326239</i>	<i>76595</i>
<i>[11C]PIB volume</i>	<b>Baseline</b>	368415	622213	12176	5294	366916	20617	599768	24371
	<b>Follow-up</b>	484441	633263	21246	15943	432670	13281	383028	18303
	<b>Changes</b>	<i>116026</i>	<i>11050</i>	<i>9070</i>	<i>10649</i>	<i>65754</i>	<i>-7336</i>	<i>55534</i>	<i>-6068</i>
<i>[11C](R)PK11195</i> <i>volume</i>		<i>AD1</i> <i>(Aβ+)</i>	<i>AD2</i> <i>(Aβ+)</i>	<i>AD3</i> <i>(Aβ+)</i>	<i>AD4</i> <i>(Aβ-)</i>	<i>AD5</i> <i>(Aβ+)</i>	<i>AD6</i> <i>(Aβ+)</i>	<i>AD7</i> <i>(Aβ+)</i>	<i>AD8</i> <i>(Aβ+)</i>
	<b>Baseline</b>	204649	16943	99337	162536	341955	41803	312547	36998
	<b>Follow-up</b>	26742	53223	373971	311870	83399	46645	829597	110810
<i>[11C]PIB volume</i>	<b>Changes</b>	<i>-177907</i>	<i>36280</i>	<i>274634</i>	<i>149334</i>	<i>-258556</i>	<i>4842</i>	<i>517050</i>	<i>73812</i>
	<b>Baseline</b>	338747	376803	217421	27454	36697	273064	347069	560597
	<b>Follow-up</b>	157210	374892	225236	92410	15648	341343	366402	1094708
	<b>Changes</b>	<i>-181537</i>	<i>-1911</i>	<i>7815</i>	<i>64956</i>	<i>-21049</i>	<i>68279</i>	<i>19333</i>	<i>534111</i>

Values in italics represent the changes in number of significant voxels between baseline and follow-up; *Aβ+* = Amyloid positive MCI; *Aβ-* = Amyloid negative MCI.