1 **A Delphic Consensus assessment: Imaging and Biomarkers in Gastroenteropancreatic**

2 **Neuroendocrine Tumour Disease Management**

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ABSTRACT

The complexity of the clinical management of neuroendocrine neoplasia (NEN), is exacerbated by limitations in imaging modalities and a paucity of clinically useful biomarkers. Limitations in currently available imaging reflect difficulties in measuring an intrinsically indolent disease, resolution inadequacies, inter-/intra-facility device variability, and that RECIST (Response Evaluation Criteria in Solid Tumours) criteria are not optimal for NEN. Limitations of currently utilized biomarkers are that they are secretory biomarkers (chromogranin A, serotonin, neuron-specific enolase, pancreastatin), monoanalyte measurements, and lack sensitivity, specificity and predictive capacity. None meet NIH metrics for clinical usage. A multinational, multidisciplinary Delphi consensus meeting of NEN experts (*n*=33) assessed current imaging strategies as well as biomarkers in NEN management. Consensus (>75%) was achieved for 78% of 142 questions. The panel concluded that morphological imaging has diagnostic value. However, both imaging and current single-analyte biomarkers exhibit substantial limitations in measuring disease status and predicting therapeutic efficacy. RECIST remains sub-optimal as a metric. A critical unmet need is the development of a clinico-biological tool to provide enhanced information regarding precise disease status and treatment response. The group concluded that circulating mRNA was a more effective tool than current monoanalyte NEN biomarkers and clinical data were auspicious. It resolved that circulating multianalyte mRNA (NETest) had clinical utility in both diagnosis and monitoring disease status and therapeutic efficacy. Overall, it was concluded that a combination of tumour spatial and functional imaging with circulating transcripts (mRNA) would represent the future strategy for real-time monitoring of disease progress and therapeutic efficacy.

INTRODUCTION

The management of neuroendocrine neoplasms (NENs, also called "NETs") remains clinically challenging despite advances in classification systems [1], inauguration of novel therapies, innovations in imaging and the introduction of multidisciplinary management strategies [2]. In particular, the management of NEN reflects diverse approaches often based upon empiric pronouncements, local practical experience or the availability of certain therapies. Despite the 87 promulgation of effective and applicable guidelines (e.g., WHO/ENETs classification of 2010) [3, 4] and their regular reassessment, a critical limitation is the dearth of large, randomized prospective trials. The precise delineation of definable strategies is further constrained by the tumour heterogeneity (diverse cell types, disparate molecular regulatory mechanisms and ill-understood oncogenic drivers) [5, 6]. As a consequence, five-year survival rates diverge widely (15-95%), depending on the primary site, variable tumour biology, disease extent at diagnosis, available therapeutic options and designated centers of care [7-9]. Therapeutic options remain diverse and run the full gamut from mechanistic excision to pharmacological intervention and the infusion of radioactive somatostatin analogs [10]. Strategies include somatostatin receptor agonists, "targeted" agents (mTOR inhibitors, VEGF antagonists), immunotherapy (interferon), cytotoxic chemotherapy, peptide receptor radionuclide therapy (PRRT), external radiation, and interventional radiological or probe-directed ablation [11]. In those with "indolent tumour behavior", a watch-and-wait-strategy is considered appropriate in certain selected cases [12]. Apart from "early identified" (usually serendipitous) appendiceal, rectal or gastric NETs, cure is uncommon and overwhelmingly, the majority of treatment includes diverse combinations of strategies to delay local or metastatic disease progression [13]. Given their relatively slow growth, continual assessment by imaging, biomarker levels and overall survival represents the fundamental basis for all management strategies. The need to monitor tumour responsiveness, both in clinical trials and in routine practice, is mandatory given the range of expensive, empirical and often times toxic treatment choices utilized [14].

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For many non-neuroendocrine neoplasms, therapeutic responsiveness is assessed through imaging, but for NENs, this has well-described limitations [15-17]. Anatomic imaging using the *Response Evaluation Criteria in Solid Tumours* (RECIST) criteria exhibits well-documented limitations [18-20]. These include issues with lesion dimensionality and measurements thereof, effects of therapy on lesion appearance itself, difficulties with reproducibility and accurate delineation of metastatic disease, particularly extra-liver disease. The development of new lesions is probably the most powerful indicator of disease progression. 114 Functional imaging with somatostatin receptor-based strategies e.g., 68 Ga-SSA-PET/CT, has proved of considerable value [21], but limited spatial resolution (6-8 mms for PET-scanners) and partial volume effects, constrain the ability to delineate small lesions. As a consequence, timely, clinically reproducible assessments of progression remains unattainable [22, 23]. Changes in 118 the ⁶⁸ Ga-SSA tumour standardized uptake value (SUV) during treatment have not been a 119 reliable measure for therapy monitoring $[24, 25]$. ¹⁸ FDG-PET, though useful prognostically, is not established as an early harbinger of tumour progression [26]. Despite significant advances, current imaging strategies in NENs remain sub-optimal [27, 28] and exhibit significant limitations. In particular, the identification and delineation of residual (and occult) disease is difficult.

Credible general biomarkers with broad clinical utility for gastroenteropancreatic (GEP)- NENs remain unavailable although chromogranin A (CgA) and urinary 5 hydroxy-indoleacetic acid (5-HIAA; in serotonin-secreting tumours) have been used in this capacity [29]. Secretory (monoanalyte) biomarkers for specific tumour types (insulinoma: insulin, gastrinoma: gastrin, glucagonoma: glucagon, VIPoma: VIP), are effective serum indicators of tumour activity, but since this group of lesions represent a minority of NENs (<3-5%), their broad utility is limited. 130 CgA is a constitutive product of the neuroendocrine cell secretory granule and is measurable in serum or plasma. It has been variously reported to correlate with tumour biology and mass and prognosticate survival [30, 31]. Despite initial enthusiasm, the limitations of CgA have become

increasingly evident. There is considerable discrepancy as to whether alterations in CgA have clinical utility in the identification of progressive disease. Although there has been some improvement regarding comparable unit use, there is no reference CgA standard and wide variations exist in the assay measurements in different laboratories [30]. Furthermore, the sensitivity of CgA ranges from 60–90% with a specificity <50% (depending on the population studied) [32]. This reflects the CgA elevations associated with numerous non NEN-related conditions including renal failure, cardiac disease, other neoplasia as well as PPI administration [30].

The complexity and diversity of the biological behavior of a cancer or its response to therapy have been effectively addressed in scientific publications [33, 34]. The limitations of secretory products to define the permutations of oncogenic genomic regulators are apparent, and have led to the development of molecular technologies to better delineate cancer biology [35, 36]. This biological research has identified extensive interfacing mechanisms that delineate GEP-NEN neoplastic development [37]. A key unmet need is the identification of what constitutes the driver of neoplastic development (i.e., driver mutations) and whether this is clinically actionable i.e., targetable, and can be used as a predictive biomarker.

The majority of tumors (~95%) do not exhibit germline mutations [6, 38]. While genomic studies have revealed a number of sporadic genomic alterations, particularly in pancreatic NENs, the relationship between specific genes and tumour pathobiology remains unclear [5]. Unlike the majority of cancers, activating mutations are infrequent if not largely unknown in GEP-NEN [5] with most tumours exhibiting mutations (when identified) in tumour suppressor genes. While genomic studies seeking underlying driver mutations have proven disappointing [39, 40], transcriptome assessments have been useful in identifying and differentiating the different subtypes of NENs (based on origin e.g., pancreatic versus small intestinal, and aggressiveness e.g., non-progressive versus malignant/metastatic) [41, 42] and have demonstrable predictive utility at a tissue level [43]. More recently, blood-based assays (CTCs,

miRNA and circulating mRNA) have been developed. The most extensively investigated biomarker tool is blood-based multianalyte transcript analysis [44-54]. Blood gene expression of tumour biomarkers closely correlates with tumour tissue expression levels, and analysis of relevant clusters captures NEN biology facilitating accurate definition of clinical status [37]. The clinical application of such blood-based information to the management of NEN disease has therefore become a subject for investigation. Likewise, the concept of fusing such data with functional imaging to provide a synergistic monitoring platform is worthy of consideration, especially given the current limitations in accurate monitoring.

Although biomarkers have been used in conjunction with imaging as adjuncts to inform clinical decision making, "biochemical" responses using monoanalytes are often non-concordant with image-based assessments [10, 55]. The detailed analysis of other neoplastic diseases has led to the recognition that evaluation of monoanalyte secretory products (exocytotic or secreted proteins) alone fails to adequately describe the diversity of neoplastic pathobiology [56]. Thus, complex analytic strategies measuring diverse regulators of neoplastic cell biology interfaced with mathematical algorithms to facilitate interpretation have been developed for breast, lung and hematological malignancies [57-60]. A key unmet need therefore remains the development of a clinically applicable, multianalyte biomarker that captures NEN behavior and can be used to guide clinical management strategies. The use of such blood-based molecular information in combination with functional imaging would provide non-invasive real time multidimensional information in regard to tumour behavior.

Based upon the need for a better understanding of the relationship between imaging and therapeutic assessment in NEN disease and the emergence of molecular-based biomarkers that have utility in assessing disease status e.g., blood-based multianalyte transcript analysis NETest [37], a meeting of multidisciplinary experts in the field was convened in Casteldefells, Spain in March 2015. The goals of this forum were twofold. Firstly, to establish a consensus on the state of the art of imaging and biomarkers in NEN and secondly, to identify how these two

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information disciplines could be interfaced to provide added value in clinical decision-making and therapeutic response assessment. This meeting represents a follow-up of a previous, more biomarker focused Delphi consensus meeting that specifically examined the current status of circulating analytes in the management of GEP-NETs in respect of their individual metrics and clinical utility [61].

MATERIALS AND METHODS

Thirty-three multinational experts in the field of NEN disease diagnosis and management were identified including nuclear medicine physicians (*n*=12; A. Kjaer, E. Krenning, D. Kwekkeboom, L. Bodei, V. Ambrosini, R. Baum, J. Cwikla, G. Paganelli, S. Severi, H. Maecke, V. Prasad, I. Virgolini), radiologists (*n*=2: A. Sundin, K. Koopmans), endocrinologists (*n*=2; M. Pavel, A. Grossman), gastroenterologists (*n*=1, R. Jensen), oncologists (*n*=9, K. Oberg, M. Tesselaar, M. Kulke, N. Fazio, R. Salazar, J. Strosberg, A. Walenkamp, M. Cives, T. Meyer [*see* Authors contributions]), pathologists (*n*=1, A. Scarpa), basic scientists (*n*=3, M. Kidd, I. Drozdov, T. Korse) and surgeons (*n*=3: M. Falconi, A. Frilling, I. Modlin). The Delphi method [62] was utilized to achieve consensus on 142 questions, using a 75% agreement level as the basis for achieving consensus [61]. Questions were categorized into four major groups (Therapeutic Management, Imaging, Molecular Status of NETs, and Biomarkers). The first iteration of the statements to be discussed was developed by a core group (KO, EK, LB, IMM) and distributed to all participants eight weeks prior to the conference. This first round electronic assessment was undertaken to eliminate or redefine inconsistencies or ambiguous statements [61]. After integration of the primary assessment comments from all participants, this second list (revised) of statements/questions (yes or no responses) was electronically distributed one month ahead of the consensus meeting. All participants provided answers to this interrogatory. The collated results of the entire group responses were made available to all participants at the initiation of the meeting. The meeting format comprised two co-moderators for each discussion session.

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Any question with less than 75% prior agreement (either Consensus: Yes or Consensus: No) was then reviewed and discussed by the entire panel and re-voted on. Voting was anonymous (electronic touch pad) with re-wording of ambiguous, controversial or non-consensus statements as proposed by participants with the objective of attaining a 75% agreement threshold [61]. Up to five re-iterations of a proposal were undertaken before considering an issue resolved. Resolution was achieved in 78%. Not all questions (22%) resulted in a consensus.

RESULTS

A total of 142 questions and sub-questions were posed. First round electronic consensus was

achieved prior to the March 2015 meeting in 69 (48.5%). At the meeting, after

statement/question reformulation and repeat voting, final consensus was achieved on 111

(78%). The full lists of statements and voting results are documented in the *Appendix*. Three

participants (ID, HM, DK) were unable to attend the meeting and participate in the final round of

voting. The final consensus therefore includes input from these members at rounds 1 and 2 but not round 3.

A. Therapeutic Management

Consensus was achieved on 30 questions (47%) prior to the meeting. A further 16 (total of 72%) met consensus after discussion and re-voting. The panelists agreed that optimal management strategies required assessment of information based upon: histology, grade and stage, specific and non-specific symptoms, as well as knowledge regarding the patient's overall condition. However, they also decided that clinical knowledge alone was inadequate for predicting whether a NEN would be progressive or exhibit stable disease. Although a wait-and-see strategy was considered an acceptable management strategy, there was full concurrence that current diagnostic parameters were neither of adequate sensitivity nor specificity for defining progress.

Moreover, currently available Randomized Controlled Trial (RCT) data were considered insufficient to accurately delineate the optimal therapeutic sequence strategy in NEN disease. Overall, the group concluded that there was a paucity of rigorous data available to facilitate objective, clinical decision-making.

In respect of imaging, current standard diagnostic parameters are neither sensitive nor specific enough to define progress. Additional predictors of the individual course of disease are therefore required to identify individuals in whom early treatment may be of benefit. This would include additional imaging parameters. Limitations in the assessment of therapeutic responses with current imaging has a negative impact on patient management. Limitations in the discriminant index of both anatomic and functional imaging diminished the accuracy of assessment of therapeutic response. Somatostatin receptor (SSR) density was considered a relevant parameter but knowing the liver tumour load and pretreatment growth rate were considered important predictors of disease course. It was agreed that additional predictors of the individual course of a specific tumour are required to define those in whom early treatment may be of benefit. Biomarkers including but not limited to tissue gene signatures, circulating genetic information and mutational events were considered critical requirements for such a strategy.

The thresholds and cut-offs for defining histopathology, Ki67 were considered problematic for defining when chemotherapy should be considered. No consensus could be reached upon the precise applicable cut-off. Ki67 was not considered a relevant parameter for predicting SSA response. Surgery was considered the only curative treatment and a blood signature that could predict disease relapse following R0/R1 (primary or liver) resection was agreed upon as an important requirement. It was identified that selective internal radiation therapy (SIRT), radio frequency ablation (RFA) and trans-arterial (chemo-) embolization (TACE/TAE) were all effective in metastatic liver disease, though individual modalities differed in efficacy based upon patient selection and disease status [63]. Individual interventions were

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noted to have adverse events though lack of comparable data prevented rigorous comparison [63]. No consensus was reached regarding associations with adverse events. Regarding, somatostatin analogs (SSAs), use should not only be limited to midgut and pancreatic NENs with K-i67<10%, but no consensus could be reached as to whether SSAs were effective early in the disease course to prevent disease progression. Likewise, it was not accepted that there was evidence that above-label doses should be used in non-functioning progressive disease. There also was not sufficient data to support the use of SSAs as anti-proliferative agents in patients with significant metastatic burden e.g., >50% neuroendocrine tumour liver metastases (NELM) and/or extra-hepatic metastases. The panel was unsure whether Everolimus had a role in non-pancreatic NEN disease (it should be noted that this meeting occurred prior to the publication of the Radiant-4 study [64]). Controversy was also apparent regarding initial therapeutic use of chemotherapy. The group was of the opinion that PRRT might warrant consideration at an earlier time-point in the therapeutic strategy for management of NETs (it should be noted that this meeting occurred prior to the availability of the NETTER-1 study results [65]). It was, however, deemed appropriate to consider the use of PRRT before other targeted therapies. Overall, a substantial lack of consensus (~28%) was evident for GEP-NEN therapeutic management. This likely reflects the individualized, empiric-based approaches and the divergent views of European and US experts.

B. Imaging

Consensus was achieved in 72% of questions (**Figure 1**). There was agreement that CT or MRI 284 should be used in conjunction with functional imaging. 68 Ga-SSA-PET/CT was preferred to 111 In-285 pentetreotide scintigraphy for functional imaging. ⁶⁸Ga-SSA-PET/CT was considered the 286 preferred approach compared to ${}^{18}F$ -DOPA imaging for pancreatic and small intestinal NEN 287 diagnosis. 18 F-FDG-PET/CT was considered useful for differentiating high from low grade tumours which might have future implications for staging. The technique, however, has

prognostic implications although this requires validation in larger series. No consensus,

290 however, was reached regarding combining 18 F-FDG- and 68 Ga-SSA-PET/CT or the timing of

imaging for use of each of these modalities in a diagnostic setting.

Imaging was considered the best current modality for measuring treatment efficacy but no consensus was achieved regarding the optimal strategy, PET/CT or CT or MRI. It was agreed that RECIST criteria were not appropriate for defining therapeutic responses in NETs at least for biological therapy, and furthermore inclusion of morphologic parameters e.g., attenuation measurements, were not considered useful. No consensus was reached regarding whether "cold" analogs e.g., Sandostatin or Lanreotide (non-radioactive without bound isotopes), should be discontinued before somatostatin receptor imaging (SRI). Overall, the heterogeneity in SSR expression was considered a potential sensitivity limitation to this 300 approach since current ligands are SSR2/5 avid. Similarly, the SUV_{max} was also not considered an entirely reliable parameter for assessing patient management based on current ligand-302 receptor affinities [66]. Based upon currently available studies, different ⁶⁸ Ga-DOTA-SSA peptides (DOTA-TOC, DOTA-NOC and DOTA-TATE) were individually as effective in their diagnostic accuracy. All were considered to have clinical utility in determining clinical management.

Overall, imaging was considered more sensitive than existing biomarkers for detecting disease. The group concurred that more effective circulating biomarkers would be a useful adjunct for assessing treatment. It was agreed that current biomarkers such as CgA do not 309 correlate with imaging, particularly 68 Ga-DOTA-SSA and 18 F-FDG imaging. No consensus could be reached for the relationship between CT or MRI and CgA. Overall, the panel agreed that integration of a clinically relevant, biologically effective biomarker strategy into response criteria was required to improve NEN therapy monitoring.

C: Molecular Status of NETs

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Consensus was achieved in the majority of questions (95%). Metabolic pathways were agreed 316 to be poorly characterized. The $PI_3K/mTOR$ pathway was not considered to be the principal growth regulatory pathway in NENs. It is as yet unclear what constitutes the precise mechanistic basis of the critical growth regulatory pathways of neuroendocrine tumour cells. Despite the proposal of numerous putative targetable pathways, current agents are not generally accepted as being of robust clinical utility [67]. Alternative pathways remain to be defined. Mutations in the mTOR pathway were noted to occur in <15% of pancreatic NENs, and the objective response rate for Everolimus (mTOR pathway inhibitor) is ~10% with disease stabilization in ~75% [68]. The discrepancy between mutation rate and therapeutic efficacy is currently difficult to reconcile. Selective PI₃K inhibitors were considered useful for overcoming Everolimus resistance although the mechanisms of resistance remain to be defined. Mutations in the ATRX/DAXX pathways were not considered major indicators of clinical outcome and it was agreed they should not be routinely assessed in pancreatic NENs. In patients with multiple endocrine neoplasia type I (MEN1) syndrome (germline MEN-1 mutation), the type of menin mutation was not considered to be of prognostic significance. Alterations in methylation patterns were likewise not considered clinically useful, while O6-methylguanine DNA transferase deficiency was regarded as not significant in influencing the choice of therapy. Irrespective of the individual molecular abnormality described, cell line models were considered unreliable for identifying and confirming the utility of any targeted agent.

No consensus could be reached regarding the role of VEGF expression and tumour aggressiveness. It was agreed that immunohistochemistry for SSR was not needed to define a treatment strategy but immunohistochemistry (IHC) e.g., CDX2 and PAX6 was recommended when a primary site was unknown (CUP). Gene profiling, in this setting (CUP) was, however, not clinically recommended. Overall, it remained unclear how molecular alterations, particularly at a DNA level, could potentially improve clinical management strategies. It was concluded that molecular alterations as currently defined did not have a current role in NEN treatment, but the

panel did support continued investigation in these areas to further define the molecular basis of NEN disease.

D. Biomarkers

A consensus was reached in 89% of questions (**Figure 2**). It was agreed that despite the paucity of DNA-related clinically actionable biomarkers, genomics technology had significant potential for identifying novel tissue biomarkers. The conclusion, however, was that at present insufficient specific mutations and treatment-targetable mutations had been identified. As such, circulating DNA was therefore not considered a viable option for the development of a biomarker.

In general, circulating tumour cells (CTCs) were agreed not to be reliable, sensitive or specific for the detection (88% No) and diagnosis (92% No) of NENs. Furthermore, once tumours were diagnosed, CTCs were considered not to correlate with grade (77% No) or to have clinical utility as either a prognostic (85% No) or predictive biomarker (77% No). No consensus was achieved relating the utility of CTCs as an indicator of tumour burden. While miRNA was considered interesting and potentially useful as a circulating biomarker, the group agreed that current technology was not adequately robust to support clinical usage. Metabolomics was also considered of positive interest (83% Yes) as was the identification of novel blood GEP-NEN biomarkers. The consideration of metabolomic assessment in urine was not supported (83% No). Tumour transcriptomes and mRNA studies were agreed to be useful for identifying tissue biomarkers and more sensitive than standard biomarkers. Circulating mRNA assays were agreed to be worthy of further investigation given their potential clinical utility.

DISCUSSION

The Delphi method, originally developed by the RAND Corporation [62], has been used extensively to develop consensus in healthcare. We have previously assessed its utility in similar clinical decision-making settings [61, 69]. In this meeting, a substantial overall consensus (~80%) was achieved with 31 questions (~20%) ultimately unresolved (no consensus achieved). A consensus level of 75% was used as clear evidence of a majority opinion. Voting was anonymized (electronic) and followed by discussion when there was no consensus. The actual numbers of participants who completed all three rounds (*n*=30, 91% inclusion) is similar to other Delphi-based studies for NENs and met the acceptability criteria for validity [69, 70]. Therapeutic management and imaging achieved the lowest consensus (72%) compared to molecular biology and biomarkers (88-95%). This likely reflects two issues. Firstly, individual approaches to management (despite a focus on multidisciplinary methods) and secondly, 377 differential access to imaging $(^{68}Ga$ -DOTA-SSA PET/CT is currently not generally available in the US). There was a full consensus that surgery was potentially curative. Similarly, there was 379 broad consensus of the utility of 68 Ga-DOTA-SSA PET/CT both in establishing a diagnosis and having a role in staging, predicting response to PRRT and determining prognosis. There are a number of different national and societal neuroendocrine guidelines that variously evaluate the usage of biomarkers and imaging (North American – NANETs, National Comprehensive Cancer Network – NCCN, Canadian NETs and the European Neuroendocrine Tumor Society – ENETs, 384 [14, 71-75]. Each broadly supports the points defined in this Delphi Consensus but none specifically addresses the interface between imaging and biomarkers nor the best strategy to integrate anatomical and functional imaging with circulating molecular information. In particular, the current consensus meeting evaluated not only the utility of the different strategies (imaging and biomarkers) but how such modalities could be interfaced to provide a real-time assessment of the biological evolution of a neuroendocrine neoplasm. It was widely agreed that current approaches (RECIST) for assessing therapeutic responses were inadequate. In particular,

clinical knowledge was considered insufficient for early and accurate predictions of progressive or stable disease. Moreover, it was agreed that a clinically actionable, biologically-relevant biomarker should be included in treatment response assessments. This is consistent with the agreement reached in the previous Delphi consensus meeting (2014) that was designed to specifically address biomarker metrics and clinical utility [61].

Although biomarkers such as CgA are currently used in conjunction with imaging as adjuncts for clinical decision making (**Figure 3**), significant refinements are required [61]. In particular, implementations of more informative molecular tools such as multianalyte biomarkers are needed. Dynamic characterization of tumour behavior based upon blood-derived genomic information is likely to be of considerable clinical utility, especially if used as an adjunct to both spatial and functional imaging. This is underscored by the lack of utility and clinical effectiveness of solely secretory biomarkers. For example, CgA does not correlate with imaging, particularly 68 Ga-DOTA-SSA and ¹⁸ F-FDG imaging, while CgA biochemical "responses" to therapy are also typically non-concordant with imaging [61]. Indeed, a number of national and societal guidelines adjudge CgA to be "controversial" in clinical decision-making [14, 71].

Imaging alone, however, also has its limitations. The panel agreed that current strategies, although useful in diagnosis, were unlikely to be improved in NENs in the near future. For example, measurements of changes in Hounsfield Units, proposed in the Choi criteria for measuring GIST treatment responses [15], may not be useful in GEP-NENs. Although suitable 410 for a rough estimate, SUV_{max} determined by 68 Ga-SSA-PET/CT, was also not considered to be ideal, since SSR heterogeneity in individual tumours is a problematic factor for sensitive 412 assessment of treatment response. Moreover, the differences in intrinsic variabilities in SUV $_{\text{max}}$ in separate PET/CT scanners at different institutions was a limitation for image-based 414 assessment and patient follow-up [54]. Changes in tumour SUV_{max} during PRRT also do not 415 always correlate to the outcome [25, 76] and in tumours with SUV_{max} > 20-25, SUV does not linearly correlate with SSR expression [77]. Other imaging biomarkers, such as activated

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samples and may be altered by somatostatin analogs) [82]. Similarly, metabolomics investigations were considered of interest since functional and non-functional tumors are readily 445 separated (R^2 =0.98) [83]. Further clinical data was necessary to further assess clinical utility. In respect of CTCs, the consensus was that this parameter remained problematic at the present time. While there is some literature to support CTCs [84, 85], all represent a single center study and hence enthusiasm was diminished. Concerns were also raised in regard to technological aspects of the measurement. Analysis of results demonstrate the clinical sensitivity (number of patients with detectable CTCs) is low, 33% in the first study and 49% in the second. Such low numbers may reflect variable EpCAM expression used for tumor cell capture. Irrespective of technical issues, it remains difficult to reconcile the utility of a test that is based on the absence or presence of 1 circulating tumor cell. This opinion directly recapitulated that expressed at the biomarker-focused Delphic consensus meeting (2014) where a separate group of international experts expressed a similar lack of enthusiasm for the clinical utility of circulating tumor cell technology [61]. None of these parameters (CTC, miRNA, metabolomics) are currently clinically recommended in guidelines. Overall, blood-based multianalyte transcript analysis [44, 45], with a clinical sensitivity >95%, was considered by the group to be more sensitive than standard biomarkers and of potential clinical utility. This is concordant with the consensus from the previous Delphi panel (2014) which evaluated the efficacy, metrics and clinical utility of current NET biomarkers [61]. Its precise application to guiding therapy was considered to require further evaluation. Current preliminary data [6, 46] were, however, noted to have specifically addressed clinical utility in sporadic, well-differentiated GEP-NETs. A role in familial NETs (including germline MEN-1 and VHL mutations) is currently under evaluation. The efficacy of a molecular tool capable of detecting germline disease evolution over time is of particular clinical relevance given the low accuracy of current biomarkers and the limitations of imagery (sensitivity and radiation exposure) as a life-long monitoring tool [86]. The areas of efficacy were identified as assessment of the effectiveness of curative surgery, assessment of the efficacy of SSA therapy,

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prediction of disease stability/progression and identification of response to PRRT. The signature was decreased by surgery and values corresponded to the completeness of tumour removal [49]. In addition, elevated levels following R0 resection predicted subsequent disease recurrence. In a different study, elevated transcript levels were prognostic of SSA failure/disease progression [51]. Of note was the observation that alterations in transcript levels occurred significantly earlier than RECIST- or SRI-based measures of disease progression [51]. Finally, levels were prognostic for PRRT efficacy and could be used to evaluate therapy, correlating with image-based assessments [53]. The observation that NEN gene blood levels 477 correlated with 68 Ga-DOTA-SSA PET/CT imaging and could define disease status was considered worthy of further clinical study [52]. In the latter study, a quotient including specific 479 genes as well as the SUV $_{\text{max}}$ accurately predicted clinical status. Thus, stable disease could be differentiated from progression using a time point amalgam of a single image/blood sample. The group considered that the combination of imaging and circulating blood biomarker offered a potential for fusing these two functional modalities of treatment assessment into a clinical index of disease status. This novel consideration had not been previously evaluated at the initial Delphi analysis (2014) which developed a biomarker-centric analysis of disease management. The larger and more diverse international cohort of experts that comprised the current Delphi group was designed to assess the effectiveness and facility of the integration of validated imaging strategies as a combinatorial clinical assessment tool with biomarkers.

In conclusion, there was consensus among a large (*n*=33) group of NEN disease experts from diverse medical and scientific disciplines and countries that current imaging and circulating biomarkers for NEN disease have substantial limitations for predicting disease activity and for measuring therapeutic efficacy. In addition, RECIST remains sub-optimal as a metric of disease status and better tools for assessment as well as improved techniques for imaging require development. These views broadly recapitulate published guidelines for GEP-NETs [14, 71-75] while providing a more in depth and detailed evaluation of the strengths and weaknesses of the

different strategies and how best they might be integrated to provide synergistic information of clinical utility. It was concluded that a critical requirement was the development of a multianalyte molecular tool that can better identify disease status and define treatment response. In this respect, the use of circulating RNA as a biomarker was confirmed to supersede the effectiveness of standard monoanalyte biomarkers and have potential clinical applicability. This assessment corroborated the outcome of the previous biomarker-centric Delphi consensus meeting [61]. Current data suggests added value for the transcript analysis in the monitoring of diverse therapeutic modalities, particularly in conjunction with other parameters to monitor disease progression (**Figure 4**). The NEN experts concluded that combinations of imaging and blood-based molecular information provided by transcriptome analysis could offer the most promising future strategy for refining and improving the evaluation of therapy.

DECLARATION OF INTEREST

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518 **AUTHOR CONTRIBUTIONS**

519 All authors were involved in the development of the manuscript and the recommendations. All

520 authors contributed equally. T. Meyer accepted financial and travel support, voted in all the

- 521 Delphi consensus iterations but ultimately declined to participate in the manuscript.
- 522

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527 **REFERENCES**

528 1 Kidd M, Modlin I, Oberg K: Towards a new classification of gastroenteropancreatic

- 529 neuroendocrine neoplasms. Nat Rev Clin Oncol 2016;7:85.
- 530 2 Oberg K: Neuroendocrine tumors of the digestive tract: Impact of new classifications and new 531 agents on therapeutic approaches. Curr Opin Oncol 2012;24:433-440. doi:
- 532 410.1097/CCO.1090b1013e328353d328357ba.
- 533 3 Bosman FT: Who classification of tumours of the digestive system. Lyon, World Health 534 Organization.; International Agency for Research on Cancer., 2010.
- 535 4 Salazar R, Wiedenmann B, Rindi G, Ruszniewski P: Enets 2011 consensus guidelines for the 536 management of patients with digestive neuroendocrine tumors: An update. Neuroendocrinology 537 2012;95:71-73. doi: 10.1159/000335600. Epub 000332012 Feb 000335615.
- 538 5 Kidd M, Modlin I, Bodei L, Drozdov I: Decoding the molecular and mutational ambiguities of 539 gastroenteropancreatic neuroendocrine neoplasm pathobiology. Cellular and Molecular 540 Gastroenterology and Hepatology 2015;1:131-153.
- 541 6 Lewis MA, Yao JC: Molecular pathology and genetics of gastrointestinal neuroendocrine 542 tumours. Curr Opin Endocrinol Diabetes Obes 2013;4:4.
- 543 7 Modlin IM, Oberg K, Chung DC, Jensen RT, de Herder WW, Thakker RV, Caplin M, Delle Fave G, 544 Kaltsas GA, Krenning EP, Moss SF, Nilsson O, Rindi G, Salazar R, Ruszniewski P, Sundin A:
- 545 Gastroenteropancreatic neuroendocrine tumours. Lancet Oncol 2008;9:61-72.
- 546 8 Yao JC, Hassan M, Phan A, Dagohoy C, Leary C, Mares JE, Abdalla EK, Fleming JB, Vauthey JN, 547 Rashid A, Evans DB: One hundred years after "carcinoid": Epidemiology of and prognostic factors for 548 neuroendocrine tumors in 35,825 cases in the united states. J Clin Oncol 2008;26:3063-3072.
- 549 9 Garcia-Carbonero R, Capdevila J, Crespo-Herrero G, Diaz-Perez JA, Martinez Del Prado MP, 550 Alonso Orduna V, Sevilla-Garcia I, Villabona-Artero C, Beguiristain-Gomez A, Llanos-Munoz M, Marazuela 551 M, Alvarez-Escola C, Castellano D, Vilar E, Jimenez-Fonseca P, Teule A, Sastre-Valera J, Benavent-552 Vinuelas M, Monleon A, Salazar R: Incidence, patterns of care and prognostic factors for outcome of 553 gastroenteropancreatic neuroendocrine tumors (gep-nets): Results from the national cancer registry of 554 spain (rgetne). Ann Oncol 2010;21:1794-1803. doi: 1710.1093/annonc/mdq1022. Epub 2010 Feb 1795. 555 10 Kulke MH, Siu LL, Tepper JE, Fisher G, Jaffe D, Haller DG, Ellis LM, Benedetti JK, Bergsland EK,
- 556 Hobday TJ, Van Cutsem E, Pingpank J, Oberg K, Cohen SJ, Posner MC, Yao JC: Future directions in the

557 treatment of neuroendocrine tumors: Consensus report of the national cancer institute neuroendocrine 558 tumor clinical trials planning meeting. J Clin Oncol 2011;29:934-943. 559 11 Frilling A, Modlin I, Kidd M, Russell C, Breitenstein S, Salem R, Kwekkeboom D, Lau W-Y, Klersy C, 560 Vilgrain V, Davidson B, Siegler M, Caplin M, Solcia E, Schilsky RL, Metastases ftWGNL: Recommendations 561 for management of patients with neuroendocrine liver metastases. Lancet Oncol 2014;15:e8-21. 562 12 Alexandraki KI, Kaltsas GA, Grozinsky-Glasberg S, Chatzellis E, Grossman AB: Appendiceal 563 neuroendocrine neoplasms: Diagnosis and management. Endocr Relat Cancer 2016;23:R27-41. doi: 564 10.1530/ERC-1515-0310. Epub 2015 Oct 1519. 565 13 Pavel M: Translation of molecular pathways into clinical trials of neuroendocrine tumors. 566 Neuroendocrinology 2013;97:99-112. doi: 110.1159/000336089. Epub 000332012 Apr 000336011. 567 14 Kunz PL, Reidy-Lagunes D, Anthony LB, Bertino EM, Brendtro K, Chan JA, Chen H, Jensen RT, Kim 568 MK, Klimstra DS, Kulke MH, Liu EH, Metz DC, Phan AT, Sippel RS, Strosberg JR, Yao JC: Consensus 569 guidelines for the management and treatment of neuroendocrine tumors. Pancreas 2013;42:557-577. 570 doi: 510.1097/MPA.1090b1013e31828e31834a31824. 571 15 Choi H, Charnsangavej C, Faria SC, Macapinlac HA, Burgess MA, Patel SR, Chen LL, Podoloff DA, 572 Benjamin RS: Correlation of computed tomography and positron emission tomography in patients with 573 metastatic gastrointestinal stromal tumor treated at a single institution with imatinib mesylate: Proposal 574 of new computed tomography response criteria. J Clin Oncol 2007;25:1753-1759. 575 16 Sundin A, Rockall A: Therapeutic monitoring of gastroenteropancreatic neuroendocrine tumors: 576 The challenges ahead. Neuroendocrinology 2012;96:261-271. doi: 210.1159/000342270. Epub 577 000342012 Oct 000342212. 578 17 Bodei L, Sundin A, Kidd M, Prasad V, Modlin I: The status of neuroendocrine tumor imaging: 579 From darkness to light? Neuroendocrinology 2015;101:1-17. 580 18 Eisenhauer EA, Therasse P, Bogaerts J, Schwartz LH, Sargent D, Ford R, Dancey J, Arbuck S, 581 Gwyther S, Mooney M, Rubinstein L, Shankar L, Dodd L, Kaplan R, Lacombe D, Verweij J: New response 582 evaluation criteria in solid tumours: Revised recist guideline (version 1.1). Eur J Cancer 2009;45:228-247. 583 doi: 210.1016/j.ejca.2008.1010.1026. 584 19 Neperud J, Mahvash A, Garg N, Murthy R, Szklaruk J: Can imaging patterns of neuroendocrine 585 hepatic metastases predict response yttruim-90 radioembolotherapy? World J Radiol 2013;5:241-247. 586 doi: 210.4329/wjr.v4325.i4326.4241. 587 20 Denecke T, Baur AD, Ihm C, Steffen IG, Tischer E, Arsenic R, Pascher A, Wiedenmann B, Pavel M: 588 Evaluation of radiological prognostic factors of hepatic metastases in patients with non-functional 589 pancreatic neuroendocrine tumors. Eur J Radiol 2013;82:e550-555. doi: 590 510.1016/j.ejrad.2013.1006.1017. Epub 2013 Jul 1024. 591 21 Toumpanakis C, Kim MK, Rinke A, Bergestuen DS, Thirlwell C, Khan MS, Salazar R, Oberg K: 592 Combination of cross-sectional and molecular imaging studies in the localization of 593 gastroenteropancreatic neuroendocrine tumors. Neuroendocrinology 2014;21:21. 594 22 Ruf J, Schiefer J, Kropf S, Furth C, Ulrich G, Kosiek O, Denecke T, Pavel M, Pascher A, 595 Wiedenmann B, Amthauer H: Quantification in ga-dota(0)-phe(1)-tyr(3)-octreotide positron emission 596 tomography/computed tomography: Can we be impartial about partial volume effects? 597 Neuroendocrinology 2013;97:369-374. 598 23 Virgolini I, Ambrosini V, Bomanji JB, Baum RP, Fanti S, Gabriel M, Papathanasiou ND, Pepe G, 599 Oyen W, De Cristoforo C, Chiti A: Procedure guidelines for pet/ct tumour imaging with 68ga-dota-600 conjugated peptides: 68ga-dota-toc, 68ga-dota-noc, 68ga-dota-tate. European journal of nuclear 601 medicine and molecular imaging 2010;37:2004-2010. 602 24 Gabriel M, Oberauer A, Dobrozemsky G, Decristoforo C, Putzer D, Kendler D, Uprimny C, Kovacs 603 P, Bale R, Virgolini IJ: 68ga-dota-tyr3-octreotide pet for assessing response to somatostatin-receptor604 mediated radionuclide therapy. J Nucl Med 2009;50:1427-1434. doi: 1410.2967/jnumed.1108.053421. 605 Epub 052009 Aug 053418. 606 25 Haug AR, Auernhammer CJ, Wangler B, Schmidt GP, Uebleis C, Goke B, Cumming P, Bartenstein 607 P, Tiling R, Hacker M: 68ga-dotatate pet/ct for the early prediction of response to somatostatin 608 receptor-mediated radionuclide therapy in patients with well-differentiated neuroendocrine tumors. J 609 Nucl Med 2010;51:1349-1356. Epub 2010 Aug 1318. 610 26 Binderup T, Knigge U, Loft A, Federspiel B, Kjaer A: 18f-fluorodeoxyglucose positron emission 611 tomography predicts survival of patients with neuroendocrine tumors. Clin Cancer Res 2010;16:978-985. 612 Epub 2010 Jan 2026. 613 27 Castano JP, Sundin A, Maecke HR, Villabona C, Vazquez-Albertino R, Navarro E, Oberg K: 614 Gastrointestinal neuroendocrine tumors (nets): New diagnostic and therapeutic challenges. Cancer 615 Metastasis Rev 2014;5:5. 616 28 Faivre S, Ronot M, Dreyer C, Serrate C, Hentic O, Bouattour M, Bruno O, Couvelard A, Vilgrain V, 617 Raymond E: Imaging response in neuroendocrine tumors treated with targeted therapies: The 618 experience of sunitinib. Target Oncol 2012;7:127-133. doi: 110.1007/s11523-11012-10216-y. Epub 619 12012 May 11515. 620 29 Modlin I, Kidd M, Taylor A, Drozdov I, Bodei L: Neuroendocrine tumor biomarkers: Current 621 status and perspectives. Neuroendocrinology 2014;100:265-277. 622 30 Modlin IM, Gustafsson BI, Moss SF, Pavel M, Tsolakis AV, Kidd M: Chromogranin a--biological 623 function and clinical utility in neuro endocrine tumor disease. Ann Surg Oncol 2010;17:2427-2443. 624 31 Yao JC, Pavel M, Phan AT, Kulke MH, Hoosen S, St Peter J, Cherfi A, Oberg KE: Chromogranin a 625 and neuron-specific enolase as prognostic markers in patients with advanced pnet treated with 626 everolimus. J Clin Endocrinol Metab 2011;96:3741-3749. Epub 2011 Oct 3712. 627 32 Lawrence B, Gustafsson BI, Kidd M, Pavel M, Svejda B, Modlin IM: The clinical relevance of 628 chromogranin a as a biomarker for gastroenteropancreatic neuroendocrine tumors. Endocrinol Metab 629 Clin North Am 2011;40:111-134, viii. doi: 110.1016/j.ecl.2010.1012.1001. 630 33 Hanahan D, Weinberg RA: The hallmarks of cancer. Cell 2000;100:57-70. 631 34 Hanahan D, Weinberg RA: Hallmarks of cancer: The next generation. Cell 2011;144:646-674. doi: 632 610.1016/j.cell.2011.1002.1013. 633 35 Walenkamp A, Crespo G, Fierro Maya F, Fossmark R, Igaz P, Rinke A, Tamagno G, Vitale G, Oberg 634 K, Meyer T: Hallmarks of gastrointestinal neuroendocrine tumours: Implications for treatment. Endocr 635 Relat Cancer 2014;21:R445-460. doi: 410.1530/ERC-1514-0106. 636 36 Wang E, Zaman N, McGee S, Milanese JS, Masoudi-Nejad A, O'Connor-McCourt M: Predictive 637 genomics: A cancer hallmark network framework for predicting tumor clinical phenotypes using genome 638 sequencing data. Semin Cancer Biol 2014;18:00050-00059. 639 37 Kidd M, Drozdov I, Modlin I: Blood and tissue neuroendocrine tumor gene cluster analysis 640 correlate, define hallmarks and predict disease status. Endocr Relat Cancer 2015;22:561-575. doi: 641 510.1530/ERC-1515-0092. Epub 2015 Jun 1532. 642 38 Dreijerink KM, Derks JL, Cataldo I, Scarpa A, Valk GD, Speel EJ: Genetics and epigenetics of 643 pancreatic neuroendocrine tumors and pulmonary carcinoids. Front Horm Res 2015;44:115- 644 38.:10.1159/000382138. Epub 000382015 Aug 000382114. 645 39 Banck MS, Kanwar R, Kulkarni AA, Boora GK, Metge F, Kipp BR, Zhang L, Thorland EC, Minn KT, 646 Tentu R, Eckloff BW, Wieben ED, Wu Y, Cunningham JM, Nagorney DM, Gilbert JA, Ames MM, Beutler 647 AS: The genomic landscape of small intestine neuroendocrine tumors. J Clin Invest 2013;15 648 40 Jiao Y, Shi C, Edil BH, de Wilde RF, Klimstra DS, Maitra A, Schulick RD, Tang LH, Wolfgang CL, 649 Choti MA, Velculescu VE, Diaz LA, Jr., Vogelstein B, Kinzler KW, Hruban RH, Papadopoulos N: Daxx/atrx, 650 men1, and mtor pathway genes are frequently altered in pancreatic neuroendocrine tumors. Science 651 2011;331:1199-1203.

Page 24 of 34

652 41 Kidd M, Modlin IM, Drozdov I: Gene network-based analysis identifies two potential subtypes of 653 small intestinal neuroendocrine tumors. BMC Genomics 2014;15:595.:10.1186/1471-2164-1115-1595. 654 42 Duerr EM, Mizukami Y, Ng A, Xavier RJ, Kikuchi H, Deshpande V, Warshaw AL, Glickman J, Kulke 655 MH, Chung DC: Defining molecular classifications and targets in gastroenteropancreatic neuroendocrine 656 tumors through DNA microarray analysis. Endocr Relat Cancer 2008;15:243-256. 657 43 Drozdov I, Kidd M, Nadler B, Camp RL, Mane SM, Hauso O, Gustafsson BI, Modlin IM: Predicting 658 neuroendocrine tumor (carcinoid) neoplasia using gene expression profiling and supervised machine 659 learning. Cancer 2009;115:1638-1650. 660 44 Modlin I, Drozdov I, Kidd M: The identification of gut neuroendocrine tumor disease by multiple 661 synchronous transcript analysis in blood. Plos One 2013;e63364 662 45 Modlin I, Drozdov I, Alaimo D, Callahan S, Teixeira N, Bodei L, Kidd M: A multianalyte pcr blood 663 test outperforms single analyte elisas for neuroendocrine tumor detection Endocr Relat Cancer 664 2014;21:615-628. 665 46 Halperin DM, Kulke MH, Yao JC: A tale of two tumors: Treating pancreatic and extrapancreatic 666 neuroendocrine tumors. Annu Rev Med 2014;17:17. 667 47 Modlin I, Drozdov I, Kidd M: Gut neuroendocrine tumor blood qpcr fingerprint assay: 668 Characteristics and reproducibility. Clinical Chemistry 2014;52:419-429. 669 48 Modlin IM, Aslanian H, Bodei L, Drozdov I, Kidd M: A pcr blood test outperforms chromogranin a 670 in carcinoid detection and is unaffected by ppis. Endocr Connect 2014;14:14-0100. 671 49 Modlin IM, Frilling A, Salem RR, Alaimo D, Drymousis P, Wasan HS, Callahan S, Faiz O, Weng L, 672 Teixeira N, Bodei L, Drozdov I, Kidd M: Blood measurement of neuroendocrine gene transcripts defines 673 the effectiveness of operative resection and ablation strategies. Surgery 2016;159:336-347. doi: 674 310.1016/j.surg.2015.1006.1056. Epub 2015 Oct 1019. 675 50 Modlin IM, Kidd M, Bodei L, Drozdov I, Aslanian H: The clinical utility of a novel blood-based 676 multi-transcriptome assay for the diagnosis of neuroendocrine tumors of the gastrointestinal tract. Am J 677 Gastroenterol 2015;110:1223-1232. doi: 1210.1038/ajg.2015.1160. Epub 2015 Jun 1222. 678 51 Cwikla JB, Bodei L, Kolasinska-Cwikla A, Sankowski A, Modlin IM, Kidd M: Circulating transcript 679 analysis (netest) in gep-nets treated with somatostatin analogs defines therapy. J Clin Endocrinol Metab 680 2015;8 681 52 Bodei L, Kidd M, Modlin IM, Prasad V, Severi S, Ambrosini V, Kwekkeboom DJ, Krenning EP, 682 Baum RP, Paganelli G, Drozdov I: Gene transcript analysis blood values correlate with (68)ga-dota-683 somatostatin analog (ssa) pet/ct imaging in neuroendocrine tumors and can define disease status. Eur J 684 Nucl Med Mol Imaging 2015;42:1341-1352. doi: 1310.1007/s00259-00015-03075-00259. Epub 02015 685 May 00257. 686 53 Bodei L, Kidd M, Modlin IM, Severi S, Drozdov I, Nicolini S, Kwekkeboom DJ, Krenning EP, Baum 687 RP, Paganelli G: Measurement of circulating transcripts and gene cluster analysis predicts and defines 688 therapeutic efficacy of peptide receptor radionuclide therapy (prrt) in neuroendocrine tumors. Eur J 689 Nucl Med Mol Imaging 2015;23:23. 690 54 Modlin IM, Drozdov I, Bodei L, Kidd M: Blood transcript analysis and metastatic recurrent small 691 bowel carcinoid management. BMC Cancer 2014;14:564.:10.1186/1471-2407-1114-1564. 692 55 Kidd M, Bodei L, Modlin IM: Chromogranin a: Any relevance in neuroendocrine tumors? Curr 693 Opin Endocrinol Diabetes Obes 2015;30:30. 694 56 Engels CC, Ruberta F, de Kruijf EM, van Pelt GW, Smit VT, Liefers GJ, Matsushima T, Shibayama 695 M, Ishihara H, van de Velde CJ, Kuppen PJ: The prognostic value of apoptotic and proliferative markers in 696 breast cancer. Breast Cancer Res Treat 2013;142:323-339. doi: 310.1007/s10549-10013-12748-y. Epub 697 12013 Nov 10546.

699 Zhang L, Metspalu A: Metagenes associated with survival in non-small cell lung cancer. Cancer Inform 700 2011;10:175-183. Epub 2011 Jun 2012. 701 58 Miller WR, Larionov A, Renshaw L, Anderson TJ, Walker JR, Krause A, Sing T, Evans DB, Dixon JM: 702 Gene expression profiles differentiating between breast cancers clinically responsive or resistant to 703 letrozole. J Clin Oncol 2009;27:1382-1387. doi: 1310.1200/JCO.2008.1316.8849. Epub 2009 Feb 1317. 704 59 Jaeger U, Kainz B: Monitoring minimal residual disease in aml: The right time for real time. Ann 705 Hematol 2003;82:139-147. Epub 2003 Feb 2011. 706 60 Lopez-Knowles E, Wilkerson PM, Ribas R, Anderson H, Mackay A, Ghazoui Z, Rani A, Osin P, 707 Nerurkar A, Renshaw L, Larionov A, Miller WR, Dixon JM, Reis-Filho JS, Dunbier AK, Martin LA, Dowsett 708 M: Integrative analyses identify modulators of response to neoadjuvant aromatase inhibitors in patients 709 with early breast cancer. Breast Cancer Res 2015;17:35.:10.1186/s13058-13015-10532-13050. 710 61 Oberg K, Modlin I, DeHerder W, Pavel M, Klimstra D, Frilling A, Metz D, Heaney A, Kwekkeboom 711 D, Strosberg J, Meyer T, Moss S, Washington M, Wolin E, Liu E, Goldenring J: Biomarkers for 712 neuroendocrine tumor disease: A delphic consensus assessment of multianalytes, genomics, circulating 713 cells and monoanalytes. Lancet Oncol 2015;16:e435046. 714 62 Linstone H, Turoff M: The delphi method: Techniques and applications Newark, NJ, New Jersey 715 Institute of Technology, 2002. 716 63 Frilling A, Clift AK: Therapeutic strategies for neuroendocrine liver metastases. Cancer 717 2014;1:28760. 718 64 Yao JC, Fazio N, Singh S, Buzzoni R, Carnaghi C, Wolin E, Tomasek J, Raderer M, Lahner H, Voi M, 719 Pacaud LB, Rouyrre N, Sachs C, Valle JW, Delle Fave G, Van Cutsem E, Tesselaar M, Shimada Y, Oh DY, 720 Strosberg J, Kulke MH, Pavel ME: Everolimus for the treatment of advanced, non-functional 721 neuroendocrine tumours of the lung or gastrointestinal tract (radiant-4): A randomised, placebo-722 controlled, phase 3 study. Lancet 2016;387:968-977. doi: 910.1016/S0140-6736(1015)00817-X. Epub 723 02015 Dec 00817. 724 65 Strosberg J, Wolin E, Chasen B, Kulke MH, Bushnell DL, Caplin M, Baum RP, Mittra E, Hobday T, 725 Hendifar A, Oberg K, Lopera Sierra M, Ruszniewski P, Kwekkeboom D: 177-lu-dotatate significantly 726 improves progression-free survival in patients with midgut neuroendocrine tumours: Results of the 727 phase iii netter-1 trial. Eur J Cancer 2015;51 (Suppl 3):6LBA (S710). 728 66 Kwekkeboom DJ, Kam BL, van Essen M, Teunissen JJ, van Eijck CH, Valkema R, de Jong M, de 729 Herder WW, Krenning EP: Somatostatin-receptor-based imaging and therapy of gastroenteropancreatic 730 neuroendocrine tumors. Endocr Relat Cancer 2010;17:R53-73. Print 2010 Mar. 731 67 Yao JC, Lagunes DR, Kulke MH: Targeted therapies in neuroendocrine tumors (net): Clinical trial 732 challenges and lessons learned. Oncologist 2013;18:525-532. doi: 510.1634/theoncologist.2012-0434. 733 Epub 2013 Apr 1624. 734 68 Yao JC, Shah MH, Ito T, Bohas CL, Wolin EM, Van Cutsem E, Hobday TJ, Okusaka T, Capdevila J, 735 de Vries EG, Tomassetti P, Pavel ME, Hoosen S, Haas T, Lincy J, Lebwohl D, Oberg K: Everolimus for 736 advanced pancreatic neuroendocrine tumors. N Engl J Med 2011;364:514-523. 737 69 Klimstra DS, Modlin IR, Adsay NV, Chetty R, Deshpande V, Gonen M, Jensen RT, Kidd M, Kulke 738 MH, Lloyd RV, Moran C, Moss SF, Oberg K, O'Toole D, Rindi G, Robert ME, Suster S, Tang LH, Tzen CY, 739 Washington MK, Wiedenmann B, Yao J: Pathology reporting of neuroendocrine tumors: Application of 740 the delphic consensus process to the development of a minimum pathology data set. Am J Surg Pathol 741 2010;34:300-313. doi: 310.1097/PAS.1090b1013e3181ce1447. 742 70 Strosberg JR, Fisher GA, Benson AB, Malin JL, Cherepanov D, Broder MS, Anthony LB, Arslan B, 743 Gibbs JF, Greeno E, Iyer RV, Kim MK, Maples W, Philip PA, Strosberg J, Wolin EM: Systemic treatment in 744 unresectable metastatic well-differentiated carcinoid tumors: Consensus results from a modified delphi 745 process. Pancreas 2013;42:397-404. doi: 310.1097/MPA.1090b1013e31826d31823a31817.

698 57 Urgard E, Vooder T, Vosa U, Valk K, Liu M, Luo C, Hoti F, Roosipuu R, Annilo T, Laine J, Frenz CM,

746 71 Kulke MH, Shah MH, Benson AB, 3rd, Bergsland E, Berlin JD, Blaszkowsky LS, Emerson L, 747 Engstrom PF, Fanta P, Giordano T, Goldner WS, Halfdanarson TR, Heslin MJ, Kandeel F, Kunz PL, 748 Kuvshinoff BW, 2nd, Lieu C, Moley JF, Munene G, Pillarisetty VG, Saltz L, Sosa JA, Strosberg JR, Vauthey 749 JN, Wolfgang C, Yao JC, Burns J, Freedman-Cass D: Neuroendocrine tumors, version 1.2015. J Natl Compr 750 Canc Netw 2015;13:78-108. 751 72 Singh S, Asa SL, Dey C, Kennecke H, Laidley D, Law C, Asmis T, Chan D, Ezzat S, Goodwin R, Mete 752 O, Pasieka J, Rivera J, Wong R, Segelov E, Rayson D: Diagnosis and management of gastrointestinal 753 neuroendocrine tumors: An evidence-based canadian consensus. Cancer Treat Rev 2016;47:32- 754 45.:10.1016/j.ctrv.2016.1005.1003. Epub 2016 May 1017. 755 73 Singh S, Dey C, Kennecke H, Kocha W, Maroun J, Metrakos P, Mukhtar T, Pasieka J, Rayson D, 756 Rowsell C, Sideris L, Wong R, Law C: Consensus recommendations for the diagnosis and management of 757 pancreatic neuroendocrine tumors: Guidelines from a canadian national expert group. Ann Surg Oncol 758 2015;22:2685-2699. doi: 2610.1245/s10434-10014-14145-10430. Epub 12014 Nov 10434. 759 74 Falconi M, Eriksson B, Kaltsas G, Bartsch DK, Capdevila J, Caplin M, Kos-Kudla B, Kwekkeboom D, 760 Rindi G, Kloppel G, Reed N, Kianmanesh R, Jensen RT: Enets consensus guidelines update for the 761 management of patients with functional pancreatic neuroendocrine tumors and non-functional 762 pancreatic neuroendocrine tumors. Neuroendocrinology 2016;103:153-171. doi: 110.1159/000443171. 763 Epub 000442016 Jan 000443175. 764 75 Niederle B, Pape UF, Costa F, Gross D, Kelestimur F, Knigge U, Oberg K, Pavel M, Perren A, 765 Toumpanakis C, O'Connor J, O'Toole D, Krenning E, Reed N, Kianmanesh R: Enets consensus guidelines 766 update for neuroendocrine neoplasms of the jejunum and ileum. Neuroendocrinology 2016;103:125- 767 138. doi: 110.1159/000443170. Epub 000442016 Jan 000443112. 768 76 Gabriel M, Decristoforo C, Kendler D, Dobrozemsky G, Heute D, Uprimny C, Kovacs P, Von 769 Guggenberg E, Bale R, Virgolini IJ: 68ga-dota-tyr3-octreotide pet in neuroendocrine tumors: Comparison 770 with somatostatin receptor scintigraphy and ct. J Nucl Med 2007;48:508-518. 771 77 Velikyan I, Sundin A, Sorensen J, Lubberink M, Sandstrom M, Garske-Roman U, Lundqvist H, 772 Granberg D, Eriksson B: Quantitative and qualitative intrapatient comparison of 68ga-dotatoc and 68ga-773 dotatate: Net uptake rate for accurate quantification. J Nucl Med 2014;55:204-210. doi: 774 210.2967/jnumed.2113.126177. Epub 122013 Dec 126130. 775 78 Langer P, Kann PH, Fendrich V, Richter G, Diehl S, Rothmund M, Bartsch DK: Prospective 776 evaluation of imaging procedures for the detection of pancreaticoduodenal endocrine tumors in 777 patients with multiple endocrine neoplasia type 1. World J Surg 2004;28:1317-1322. Epub 2004 Nov 778 1311. 779 79 van Asselt SJ, Brouwers AH, van Dullemen HM, van der Jagt EJ, Bongaerts AH, Kema IP, 780 Koopmans KP, Valk GD, Timmers HJ, de Herder WW, Feelders RA, Fockens P, Sluiter WJ, de Vries EG, 781 Links TP: Eus is superior for detection of pancreatic lesions compared with standard imaging in patients 782 with multiple endocrine neoplasia type 1. Gastrointest Endosc 2015;81:159-167.e152. doi: 783 110.1016/j.gie.2014.1009.1037. 784 80 Shay JW, Reddel RR, Wright WE: Cancer. Cancer and telomeres--an alternative to telomerase. 785 Science 2012;336:1388-1390. 786 81 Sei Y, Zhao X, Forbes J, Szymczak S, Li Q, Trivedi A, Voellinger M, Joy G, Feng J, Whatley M, Jones 787 MS, Harper UL, Marx SJ, Venkatesan AM, Chandrasekharappa SC, Raffeld M, Quezado MM, Louie A, 788 Chen CC, Lim RM, Agarwala R, Schaffer AA, Hughes MS, Bailey-Wilson JE, Wank SA: A hereditary form of 789 small intestinal carcinoid associated with a germline mutation in inositol polyphosphate multikinase. 790 Gastroenterology 2015;149:67-78. doi: 10.1053/j.gastro.2015.1004.1008. Epub 2015 Apr 1059. 791 82 Li SC, Khan M, Caplin M, Meyer T, Oberg K, Giandomenico V: Somatostatin analogs treated small 792 intestinal neuroendocrine tumor patients circulating micrornas. PLoS One 2015;10:e0125553. doi: 793 0125510.0121371/journal.pone.0125553. eCollection 0122015.

794 83 Kinross JM, Drymousis P, Jimenez B, Frilling A: Metabonomic profiling: A novel approach in 795 neuroendocrine neoplasias. Surgery 2013;154:1185-1192; discussion 1192-1183.

796 84 Khan MS, Kirkwood A, Tsigani T, Garcia-Hernandez J, Hartley JA, Caplin ME, Meyer T: Circulating 797 tumor cells as prognostic markers in neuroendocrine tumors. J Clin Oncol 2013;31:365-372. doi: 798 310.1200/JCO.2012.1244.2905. Epub 2012 Dec 1217.

799 85 Khan MS, Tsigani T, Rashid M, Rabouhans JS, Yu D, Luong TV, Caplin M, Meyer T: Circulating

800 tumor cells and epcam expression in neuroendocrine tumors. Clin Cancer Res 2011;17:337-345. Epub 801 2011 Jan 2011.

802 86 de Laat JM, Pieterman CR, Weijmans M, Hermus AR, Dekkers OM, de Herder WW, van der

803 Horst-Schrivers AN, Drent ML, Bisschop PH, Havekes B, Vriens MR, Valk GD: Low accuracy of tumor

804 markers for diagnosing pancreatic neuroendocrine tumors in multiple endocrine neoplasia type 1

- 805 patients. J Clin Endocrinol Metab 2013;98:4143-4151. doi: 4110.1210/jc.2013-1800. Epub 2013 Aug 806 4116.
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FIGURE LEGENDS

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- **Figure 1**. Clinical utility of imaging overview (Section B).
- 811 Imaging for diagnosis (*left*) was considered effective (71% positive); ⁶⁸Ga-DOTA-SSA PET/CT
- 812 was considered more useful than either In-pentetreotide scintigraphy (100%) or 18 F-DOPA-
- 813 PET/CT (89%) for diagnosis of well-differentiated NENs. 18 F-DOPA-PET/CT was agreed to
- accurately differentiate (88%) low from high grade tumours. Imaging in therapeutic assessment
- (*right*) was overall considered suboptimal (36%). No consensus (grey) could be reached
- 816 regarding the utility of either CT/MRI (40%) or PET-CT (46%) in the assessment of therapy. A
- combination of CT/MRI and functional imaging were considered useful (84%) There was a
- negative assessment of current methodologies including RECIST criteria (82%) and Hounsfield
- Units (Choi criteria) (76%).
- 820 68 Ga = 68 Ga-DOTA-SSA PET/CT; 111 In = 111 In-pentetreotide scintigraphy; 18 F = 18 F-DOPA-

PET/CT; HU = Hounsfield Units

Figure 2. Biomarker assessment. (Section D).

Current monoanalyte blood biomarkers including CgA, serotonin, and pancreastatin were

overall considered inadequate (80%). The utility for individual strategies was assessed as

- negative for CTC's (70%) and positive, in ascending order, for miRNA (67%), metabolomics
- (75%) and circulating mRNA (80%).
-
- **Figure 3**. Proposed Strategy for Assessing Therapeutic Efficacy.
- An integration of functional imaging and biomarker measurement including circulating tumour
- mRNA will provide combinatorial information on a real time basis of disease status. The
- combination of individual imaging strategies will quantify tumour location/extent and in addition
- 833 delineate somatostatin receptor expression (SRI typically 68 Ga-DOTA-SSA PET/CT) and

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834 tumour metabolism (¹⁸F-FDG-PET/CT). Circulating mRNA will measure tumour biological 835 activity and identify treatment response.

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837 **Figure 4**. *Conceptual proposal for the evaluation of therapeutic efficacy.* This provides an 838 integration of functional imaging and tumour molecular biology utilizing circulating multianalyte 839 assays with algorithm analyses (MAAA)s, mRNA or miRNA. Disease progress can be 840 delineated using a combination of functional imaging modalities quantifying somatostatin 841 receptor expression (SSR) by 68 Ga-DOTA-SSA PET/CT and tumour metabolism using either 842 ¹⁸ F-DOPA PET/CT (in well-differentiated tumours) or 18F-FDG (mainly in undifferentiated forms 843 or to assess tumour aggressiveness). The MAAA e.g., circulating mRNA, provides an accurate 844 reflection of tumour activity. Overall, the combination of functional imaging $(^{68}Ga\text{-SSA}$ and $^{18}F\text{-}$ 845 FDG-PET/CT) and circulating mRNA could, in the future, help to delineate treatment efficacy. 846

Figure 1

Figure 2

PROPOSED STRATEGY FOR ASSESSING THERAPEUTIC EFFICACY

Figure 3

Figure 4