**Loss of phosphodiesterase 10A expression is associated with progression and severity in Parkinson’s disease**

Niccolini F1,2, Foltynie T3, Reis Marques T4,Muhlert N5, Tzortzi AC6, Searle GE6,Natesan S4, Kapur S4, Rabiner EA6,7, Gunn RN2,6, Piccini P2, Politis M1,2

1 Neurodegeneration Imaging Group, Department of Basic and Clinical Neuroscience, Institute of Psychiatry, Psychology and Neuroscience, King’s College London, London, UK

2 Division of Brain Sciences, Department of Medicine, Imperial College London, London, UK

3 Sobell Department of Motor Neuroscience, UCL Institute of Neurology, London, UK

4 Department of Psychosis Studies, Institute of Psychiatry, Psychology and Neuroscience, King’s College London, London, UK

5 School of Psychology, Cardiff University, Cardiff, UK

6 Imanova Ltd., Centre for Imaging Sciences, Hammersmith Hospital, London, Imanova, London, UK

7 Department of Neuroimaging, Institute of Psychiatry, Psychology and Neuroscience, King’s College London, London, United Kingdom.

**Correspondance to:** Marios Politis, Neurodegeneration Imaging Group, Department of Clinical Neuroscience, Institute of Psychiatry, Psychology and Neuroscience (IoPPN), King’s College London, 16 De Crespigny Park, London SE5 8AF, UK.

E-mail: marios.politis@kcl.ac.uk

**Running title:** PDE-10A loss in Parkinson’s disease

**Abstract**

The mechanisms underlying neurodegeneration and loss of dopaminergic signalling in Parkinson’s disease are still only partially understood. Phosphodiesterase 10A (PDE-10A) is a basal ganglia expressed dual substrate enzyme, which regulates cAMP and cGMP signalling cascades, thus having a key role in the regulation of dopaminergic signalling in striatal pathways, and in promoting neuronal survival. This study aimed to assess *in vivo* the availability of PDE-10A in patients with Parkinson’s disease using PET molecular imaging with [11C]IMA107, a highly selective PDE-10A radioligand. We studied 24 patients with levodopa-treated, moderate to advanced Parkinson’s disease. Their PET imaging data were compared to those from a group of 12 healthy controls. Parametric images of [11C]IMA107 binding potential relative to non-displaceable binding (BPND) were generated from the dynamic [11C]IMA107 scans using the simplified reference tissue model with the cerebellum as the reference tissue. Corresponding region-of-interest analysis showed lower mean [11C]IMA107 BPND in the caudate (*P*<0.001), putamen (*P*<0.001) and globus pallidus (*P*=0.025) in Parkinson’s disease patients compared to healthy controls, that was confirmed with voxel-based analysis. Longer Parkinson’s duration correlated with lower [11C]IMA107 BPND in the caudate (*r*=−0.65; *P*=0.005), putamen (*r*=−0.51; *P*=0.025), and globus pallidus (*r*=−0.47; *P*=0.030). Higher Unified Parkinson's Disease Rating Scale part-III motor scores correlated with lower [11C]IMA107 BPND in the caudate (*r*=−0.54; *P*=0.011), putamen (*r*=−0.48; *P*=0.022), and globus pallidus (*r*=−0.70; *P*<0.001). Higher Unified Dyskinesia Rating Scale scores in those Parkinson’s disease with levodopa-induced dyskinesias (n=12), correlated with lower [11C]IMA107 BPND in the caudate (*r*=−0.73; *P*=0.031) and putamen (*r*=−0.74; *P*=0.031). Our findings demonstrate striatal and pallidal loss of PDE-10A expression, which is associated with Parkinson’s duration and severity of motor symptoms and complications. PDE-10A is an enzyme that could be targeted with novel pharmacotherapy, and this may help improve dopaminergic signalling and striatal output and, therefore, alleviate symptoms and complications of Parkinson’s disease.

**Keywords**

Parkinson’s disease; PDE-10A; PET; motor; LIDs.

**Abbreviations**

AIMS=Abnormal Involuntary Movement Scale; BPND=non-displaceable binding potential; LED=levodopa equivalent dose; LIDs=levodopa-induced dyskinesias; PDE-10A=phosphodiesterase 10A; PDQ-39=39-item Parkinson's disease Questionnaire; UDysRS=Unified Dyskinesia Rating Scale; UPDRS= Unified Parkinson’s Disease Rating Scale

# Introduction

Parkinson’s disease is a chronic and progressive neurodegenerative disorder characterised by the loss of dopaminergic neurotransmission in the denervated areas of the forebrain, above all in the striatum (Jellinger, 1991). The mechanisms underlying the progressive loss of dopamine neuron function and the development of motor symptoms and complications are still only partially understood. Dopamine replacement therapy such as with levodopa, remains the gold standard of therapy, several decades since the original work by Cotzias and colleagues (Cotzias *et al.*, 1967, 1969). Although levodopa has been a remarkable success in Parkinson’s therapeutics, has not delivered a complete solution for the patients. Following years of exposure, levodopa loses its efficacy and patients develop motor complications such as levodopa-induced dyskinesias (LIDs) (Lees *et al.*, 1977). There is an urgent need for complementary and alternative treatments and currently no treatment has been proven useful slowing down the progression of Parkinson’s disease or for managing LIDs effectively.

Phosphodiesterase 10A (PDE-10A) is a dual substrate enzyme located almost exclusively within the basal ganglia and mainly in the axons of the striatal medium spiny neurons, where it hydrolyses cAMP and cGMP (Fujishige *et al.,* 1999; Coskran *et al.,* 2006).In the striatal pathway,PDE-10A regulates cAMP/cGMP downstream signalling cascades (e.g. cAMP/PKA/DARPP-32) that control a diverse array of neural functions ranging from ion conductance to synaptic plasticity playing a key role in the regulation of dopaminergic signalling in the direct and indirect striatal pathways (Nishi *et al.*, 2008; Girault, 2012).

Preclinical studies in levodopa naïve animal models of Parkinson’s disease have shown that unilateral lesion of nigrostriatal dopaminergic projections leads to increased cAMP levels in the ipsilateral striatum compared to the contralateral non-lesioned side (Hossain and Weiner, 1993; Tenn and Niles, 1997; Giorgi *et al.,* 2008). In striatal neurons, cAMP synthesis is regulated by dopamine through the interaction with D1 and D2 receptors and its catabolism is mediated by phosphodiesterases such as PDE-10A (Rasmussen, 1970). In animal models of Parkinson’s disease, unilateral lesion of nigrostriatal projections not only induces an increase of cAMP but also down-regulates PDE-10A mRNA and protein levels in the striatum and globus pallidus compared to the contralateral non-lesioned side (Giorgi *et al.,* 2008, 2011). These data demonstrated that lesions of the midbrain dopaminergic neurons could regulate both PDE-10A expression and the rate of cAMP catabolism in the basal ganglia. The PDE-10A/cAMP interaction is essential for dopamine neurotransmission and could have a key role in the pathophysiology of Parkinson’s disease. Moreover, cyclic nucleotides levels were reduced at peak dyskinesias in rodent models of Parkinson’s disease and administration of PDE inhibitors before levodopa decreased the severity of dyskinesias (Giorgi *et al.*, 2008; Sancesario *et al.*, 2014).

Here, we investigated *in vivo* the expression of PDE-10A in Parkinson’s disease patients, using PET with [11C]IMA107, which is a specific and highly potent PDE-10A radioligand for human use (Plisson *et al.,* 2014). Our findings suggest loss of striatal and pallidal PDE-10A expression in patients with Parkinson’s disease, which is associated with progression of the disease and severity of motor signs and complications.

# Materials and methods

## Participants and clinical characteristics

Twenty-four patients with idiopathic Parkinson’s disease according to the UK Brain Bank criteria (12 of whom had a history of levodopa related motor complications, 12 with a stable levodopa motor response) were recruited from specialist Movement Disorders clinics at Imperial College Healthcare NHS Trust and National Hospital of Neurology & Neurosurgery, Queen Square, London (Table 1). Twelve healthy individuals (mean age ± SD: 51.1 ± 11.1) with no history of neurological or psychiatric disorders served as the control group. All participants screened successfully to undertake PET and MRI scanning under standard criteria (http://www.mrisafety.com; https://www.gov.uk/government/publications/arsac-notes-for-guidance), had no history of other neurological or psychiatric disorders, and were not under treatment with substances with known actions in PDEs (e.g. aminophylline, paraxanthine, theobromine, ibudilast and papaverine).

Parkinson’s disease patients were on levodopa treatment for at least six months at the time of study enrolment. Daily dopaminergic medication dose was calculated with a formula based on the theoretical equivalence to levodopa (Politis *et al.*, 2010; Supplementary Materials). Motor symptom severity was assessed with the Unified Parkinson’s Disease Rating Scale part-III (UPDRS-III) and staged with Hoehn & Yahr scale. For correlation with the PET data purposes with divided UPDRS-III tremor, rigidity, bradykinesia and axial subscores (Supplementary Materials). Presence/absence of motor complications was assessed with the UPDRS part-IV (UPDRS-IV). Severity of levodopa-induced dyskinesias (LIDs) was assessed with the Unified Dyskinesia Rating Scale (UDysRS). Motor assessments were performed OFF medication after overnight withdrawal of patient’s dopaminergic medications, and also following a challenge with levodopa 250/carbidopa 25, with an observational ON-medication period of 150 min. Quality of life was measured with the patient self-reported 39-item Parkinson's disease Questionnaire (PDQ-39).

The study was approved by the institutional review boards and the research ethics committee. Written informed consent was obtained from all study participants in accordance with the Declaration of Helsinki.

## Scanning procedures

PET and MR imaging was performed at Imanova Ltd, London, UK. All participants were scanned on Siemens Biograph Hi-Rez 6 PET-CT scanner (Erlangen, Germany), one hour following a standard levodopa 250/carbidopa 25 dose. A mean dose of 289 MBq [11C]IMA107 (SD: ± 39.2) [mean mass injected: 3.5 ug (SD: ± 1.8)] was administered intravenously as a slow bolus injection over 20s. All participants were scanned after withholding consumption of caffeinate beverages for 12 hours (Fredholm *et al.*, 1999).

Dynamic emission data were acquired continuously for 90 minutes following the injection of [11C]IMA107. The dynamic images were reconstructed into 26 frames (8 x 15 s, 3 x 60 s, 5 x 120 s, 5 x 300 s, and 5 x 600 s), using a filtered back projection algorithm (direct inversion Fourier transform) with a 128 matrix, zoom of 2.6 producing images with isotropic voxel size of 2 x 2 x 2 mm3, and smoothed with a transaxial Gaussian filter of 5 mm.

MRI scans were acquired with a 32-channel head coil on a Siemens Magnetom Verio (Erlangen, Germany), 3-Tesla MRI scanner and included a T1-weighted magnetization prepared rapid gradient echo sequence [MPRAGE; time repetition (TR) = 2300 ms, time echo (TE) = 2.98 ms, flip angle of 9°, time to inversion (TI) = 900 ms, matrix = 240 x 256]; fast grey matter T1 inversion recovery (FGATIR; TR = 3000 ms, TE = 2.96 ms, flip angle of 8°, TI = 409 ms, matrix = 240 x 256) (Sudhyadhom *et al.,* 2009) and fluid and white matter suppression (FLAWS; TR = 5000 ms, TE = 2.94 ms, flip angle of 5°, TI = 409/1100 ms, matrix = 240 x 256) (Tanner *et al.,* 2013) sequences for co-registration with the PET images and for improving delineation of subcortical brain regions. All sequences used a 1 mm3 voxel size, anteroposterior phase encoding direction, and a symmetric echo.

## Imaging data analysis

### *MRI-based volumetric analysis*

Since PDE-10A is an intracellular enzyme mainly expressed in the basal ganglia nuclei (Fujishige *et al.,* 1999; Coskran *et al.,* 2006), degeneration of these nuclei may affect the expression of the enzyme. Thus, we investigated volumetric changes in basal ganglia nuclei and thalamus in our cohort of Parkinson’s disease patients. We used the FreeSurfer’s image analysis suite (version 5.3.0 http://surfer.nmr.mgh.harvard.edu) to process individual MRI scans for deriving measures of cortical and subcortical volumes. The automated procedures for acquiring volumetric measures of brain structures have been previously described (Fischl *et al.,* 2002). This procedure automatically assigns a neuroanatomical label to each voxel in an MRI volume based on probabilistic information automatically estimated from a manually labelled training set. In brief, the segmentation is carried out as follows: first, an optimal linear transform is computed that maximizes the likelihood of the input image, given an atlas constructed from manually labelled images. Then, a nonlinear transform is initialized with the linear one, and the image is allowed to further deform to better match the atlas. Finally, a Bayesian segmentation procedure is carried out, and the maximum *a posteriori* estimate of the labelling is computed. The segmentation uses three pieces of information to disambiguate labels: (1) the prior probability of a given tissue class occurring at a specific atlas location, (2) the likelihood of the image intensity given that tissue class, and (3) the probability of the local spatial configuration of labels given the tissue class. This technique has previously been shown to be comparable in accuracy to manual labelling (Fischl *et al.,* 2002). Adjustments for intracranial volume were calculated for each region of interest (ROI) using validated methods within the FreeSurfer toolkit (Buckner *et al.,* 2004).

### *[11C]IMA107 PET data analysis*

#### Movement correction

Subjects were positioned supine with their transaxial planes parallel to the line intersecting the anterior-posterior commissure line. Head position was maintained with the help of individualized foam holders, monitored by video and was repositioned if movement was detected. Subjects were in a resting state with low light. Intrascan notes for participant’s movement were acquired during scanning.We applied correction for movement using a frame-by-frame realignment procedure as previously described (Montgomery *et al.,* 2006) with in-house software (c-wave) implemented in Matlab 8.2 (The MathWorks Inc.).Attenuated corrected images were realigned using a mutual information algorithm (Studholme *et al.,* 1997) excluding the first seven frames containing little information. Frame 17 was chosen as the reference frame because it offered good signal-to-noise ratio. Frames 8-26 of the original time series were then resliced and reassembled into a movement-corrected dynamic scan. Decay-corrected time–activity curves were derived and compared to those without movement correction. Amount and timing of any movement were assessed graphically and compared with intrascan notes.

#### Parametric images

First, integrated (ADD) images were created by summing the time series of [11C]IMA107 uptake scans collected 0–90 min after tracer administration. Then, parametric images of [11C]IMA107 non-displaceable binding potential (BPND) were generated using a basis function implementation of the simplified reference tissue model, with the cerebellum as the reference tissue for nonspecific binding using an in-house software (c-wave) implemented in Matlab 8.2 (Gunn et al., 1997). Previous PET studies have shown lower PDE-10A uptake in the cerebellum (Plisson *et al.,* 2011, 2014; Barret *et al.,* 2014) and a blocking study with selective PDE-10A inhibitors has shown no changes in cerebellar [11C]IMA107 binding (Imanova internal data), confirming the suitability of the cerebellum as a reference region for the determination of the regional estimation of BPND.

#### ROI-based analysis

To facilitate anatomical delineation of ROIs, PET images were anatomically co-registered and resliced to the corresponding volumetric FLAWS and FGATIR MR images and spatially normalized into the T1-weighted Montreal Neurologic Institute (MNI) 152 template using the Mutual Information Registration algorithm in Statistical Parametric Mapping version 8 (SPM8) software package implemented in Matlab 8.2. ROIs were delineated manually on the co-registered MRIs using ANALYZE version 11 (Mayo Foundation) medical imaging software package by the assessor who was blinded to groups allocation. We manually delineated basal ganglia structures due to the modest performance of automated parcellation techniques on structures such as the substantia nigra, which have poor contrast on structural T1-weighted MR images. To compensate for this, we acquired state-of-the-art FGATIR and FLAWS MRI sequences for each individual, which use T1-nulling to minimise white matter signal and, by improving contrast, increase the definition of basal ganglia structures. We have then used a reliable, robust and repeatable technique for manual delineation of basal ganglia structures (Tziortzi *et al.,* 2011). This technique has shown low BPND variability for both the intra- and inter-operator comparisons and good level of agreement with automatically derived ROIs (Tziortzi *et al.,* 2011). ROIs included caudate, putamen, ventral striatum, globus pallidus (external and internal segment), substantia nigra and motor thalamic nuclei. These brain regions express PDE-10A to a varying degree (Seeger *et al.,* 2003; Coskran *et al.,* 2006).

#### Voxel-based analysis

Spatial pre-processing and statistical analyses were performed using SPM8 implemented in Matlab 8.2. First, ADD images were co-registered to the corresponding T1-weighted MR images using a normalized mutual-information-based six parameter rigid registration. The resulting transformation matrix was then applied to the corresponding [11C]IMA107 BPND images. Then, T1-weighted MR images were spatially normalized to the T1 Montreal Neurologic Institute (MNI) template provided with SPM8 (Ashburner and Friston, 1999). The transformation parameters obtained were then applied to the co-registered BPND images. Parametric images were spatially smoothed using a 8mm full-width half-maximum Gaussian kernel. This spatial filter accommodates inter-individual anatomic variability and improves signal to noise for the statistical analysis. Voxel-wise statistics for between-group comparisons were computed using appropriately weighted contrasts to localize significant decreases in mean voxel [11C]IMA107 BPND values after applying the Basal Ganglia Human Area Template (Prodoehl *et al.,* 2008). The contrasts were used to derive Z-scores on a voxel basis using the general linear model (Friston *et al.,* 1995). The threshold for statistical significance was set to *P*<0.05 after family wise error (FWE) correction for multiple comparisons.

### Statistical analysis

Statistical analysis and graph illustration were performed with SPSS (version 20) and GraphPad Prism (version 6.0c) for MAC OS X, respectively. For all variables, variance homogeneity and Gaussianity were tested with Bartlett and Kolmogorov-Smirnov tests and we proceeded with parametric tests since our PET and clinical data were normally distributed. Multivariate analysis of variance (MANOVA) was used to assess the main effects of regional [11C]IMA107 BPND between Parkinson’s disease patients and healthy controls. If the overall multivariate test was significant, *P* values for each variable were calculated following Bonferroni’s multiple-comparisons test. We interrogated correlations between PET and clinical data using Pearson *r* and we applied the Benjamini-Hochberg correction in order to reduce false discovery rate (Benjamini and Hochberg, 1995). We set the false discovery rate cut-off at 0.05.Partial correlationanalysis was performed to control for the effect of disease duration on UDysRS score/[11C]IMA107 BPND correlation. All data are presented as mean±SD, and the level *α* was set for all comparisons at *P*<0.05, corrected. In addition, we performed SPM voxel-by-voxel comparison between controls and Parkinson’s disease patients. These comparisons were based on *a priori* hypothesis restricted to a volume of interest, which included the basal ganglia in both hemispheres (Prodoehl et al., 2008). This masking drastically reduces the number of voxel-by-voxel statistical comparisons, and a cluster-corrected threshold of *P*<0.05 used to test for statistically significant effects. Striatal [11C]IMA107 BPND results were corrected for multiple comparisons using Gaussian random field theory. Since no changes were found within the pallidum at this level, for completeness changes in pallidal PDE-10A expression were investigated using a more liberal threshold of *P*<0.05 (extent threshold of 10 voxels), to rule out the risk of a type II error (false negative).

# Results

## *Volumetric analysis*

Freesurfer analysis showed no volumetric differences in left and right basal ganglia and thalamic ROIs between the groups of Parkinson’s disease patients and healthy controls (Table S1).

## *ROI-based analysis*

We found significant differences in mean [11C]IMA107 BPND between Parkinson’s disease patients and healthy controls (*P*<0.001; Table 2; Fig 1; Fig S1). Parkinson’s disease patients had significantly lower mean [11C]IMA107 BPND in caudate (*P*<0.001; −28.4%), putamen (*P*<0.001; −25.5%;) and globus pallidus (*P*=0.025; −14.2%) compared to healthy controls. Within the globus pallidus, loss of [11C]IMA107 BPND was driven by the internal segment (*P*=0.018; −18.8%). There were no differences in [11C]IMA107 BPND between the Parkinson’s disease patients and healthy controls in ventral striatum (*P*>0.10; −1.6%), substantia nigra (*P*>0.10; −12.4%) and motor thalamic nuclei (*P*>0.10; −12.3%).

We assessed whether clinically asymmetric limb motor features correspond to [11C]IMA107 BPND decreases, and we found no significant differences in [11C]IMA107 BPND between the most and less affected sides (*P*=0.43; Table S2). Moreover, we found no effect of age in [11C]IMA107 BPND in all the regions examined in the group of healthy controls (*P*=0.32).

The 24 Parkinson’s disease patients were further divided into two subgroups, depending whether they were stable responders to levodopa (n=12) or had LIDs (n=12). We found significant differences in mean [11C]IMA107 BPND between Parkinson’s disease patients with stable levodopa response and those with LIDs (*P*=0.002; Table 3). Parkinson’s disease patients with LIDs had significantly lower mean [11C]IMA107 BPND in caudate (*P*=0.045; −18.60%), substantia nigra (*P*<0.001; −33.50%)and motor thalamic nuclei (*P*=0.009; −29.3%) compared to stable levodopa responders. Parkinson’s disease patients with LIDs had longer disease duration than Parkinson’s disease patients who were stable responders to levodopa (6.16 ±2.6 *vs* 13.17 ±4.3 years; *P*<0.001). After controlling for disease duration, we did not find significant differences in [11C]IMA107 BPND between Parkinson’s disease patients with stable levodopa response and those with LIDs (*P*>0.05).

## *Voxel-based analysis*

Voxel-by-voxel analysis of [11C]IMA107 parametric images between the group of Parkinson’s disease patients and healthy controls confirmed results obtained with ROI analysis. SPM analysis within the basal ganglia mask localized clusters of significant decreases in the right and left caudate (*P*<0.05 corrected), putamen (*P*<0.05 corrected) and globus pallidus (*P*<0.05 uncorrected) in the Parkinson’s disease patients when compared with the group of healthy controls (Table 4; Fig. 2). We repeated the SPM analysis using LED as a covariate and the results remained significant. We did not find any differences in [11C]IMA107 BPND at a voxel-level between the Parkinson’s disease patients with stable levodopa response and those with LIDs.

## Correlations

## *Disease severity*

Longer Parkinson’s disease duration correlated with lower [11C]IMA107 BPND in caudate (*r*=−0.65; *P*=0.005), putamen (*r*=−0.51; *P*=0.025) and globus pallidus (*r*=−0.47; *P*=0.03; Fig. 3A). Higher UPDRS-III motor scores (corrected for disease duration) were also correlated with lower [11C]IMA107 BPND in the caudate (*r*=−0.54; *P*=0.011), putamen (*r*=−0.48; *P*=0.022) and globus pallidus (*r*=−0.70; *P*<0.001; Fig 3B).

With regards to UPDRS-III subscores (corrected for disease duration), higher bradykinesia subscores correlated with lower [11C]IMA107 BPND in the caudate (*r*=−0.58; *P*=0.007), putamen (*r*=−0.53; *P*=0.010) and globus pallidus (*r*=−0.69; *P*<0.001; Fig 3C). Higher rigidity correlated with lower [11C]IMA107 BPND in the caudate (*r*=−0.51; *P*=0.022), putamen (*r*=−0.48; *P*=0.022) and globus pallidus (*r*=−0.67; *P*<0.001; Fig 3D). Higher axial subscores correlated only with lower [11C]IMA107 BPND in the globus pallidus (*r*=−0.57; *P*=0.009; Fig S2).We did not find any correlations between tremor subscores, Hoehn & Yahr staging scores, PDQ-39 ratings and PDE-10A data.

## *Motor complications*

Higher UDysRS scores correlated with lower [11C]IMA107 BPND in the caudate (*r*=−0.73, *P*=0.031) and putamen (*r*=−0.74, *P*=0.031; Fig 4). After controlling for disease duration, the correlation remained significant between higher UDysRS scores and lower [11C]IMA107 BPND in the caudate (*r*=−0.66, *P*=0.027) and putamen (*r*=−0.68, *P*=0.022).

***Dopaminergic medication***

We found a significant interaction between higher daily LED and longer disease duration (*r*=0.52; *P*=0.014), higher UPDRS-III scores (*r*=0.055; *P*=0.012), and lower [11C]IMA107 BPND in the ROIs examined (*r*=-0.44; *P*=0.036). Subsequently, we repeated the PET-clinical correlation analysis adding LED as a covariate and found no changes in the level of significance of the previously reported correlations between lower [11C]IMA107 BPND and (a) longer disease duration (*r*=−0.55; *P*=0.020), and (b) higher UPDRS-III motor scores (*r*=−0.64; *P*=0.008).

# Discussion

Our findings indicate loss of PDE-10A signalling in the striatum and globus pallidus of patients with Parkinson’s disease, which is associated with the duration of the disease and the severity of motor signs and complications such as LIDs. Our findings are consistent with preclinical data showing that lesion of dopaminergic neurons induces significant decreases in striatal and pallidal PDE-10A mRNA and protein levels (Giorgi *et al.,* 2011; Sancesario *et al.,* 2014), and suggest that nigrostriatal degeneration affects the expression of PDE-10A.

Our ROI analysis demonstrated 14-28% significant loss of striatal and pallidal PDE-10A expression in patients with moderate to advanced Parkinson’s disease who had mean disease duration of 9.7 years, were on dopamine-replacement therapy, and had no significant brain atrophy in the ROIs examined. We confirmed these results at a voxel level. Although Parkinson’s disease patients were older than the group of healthy controls, we did not find any effect of age in PDE-10A expression in the ROIs examined. This observation is in line with recent experimental work, which indicates no significant effect of age in striatal PDE-10A mRNA and protein levels in mice (Kelly *et al.,* 2014).

There were strong associations between loss of striatal and pallidal PDE-10A expression and Parkinson’s duration and severity of motor symptoms. We found correlations between loss of striatal and pallidal PDE-10A expression, longer disease duration and worse motor symptoms as assessed by UPDRS-III. The motor symptoms that were driving this correlation were bradykinesia and rigidity, whereas worse axial symptoms correlated with decreased PDE-10A expression only in globus pallidus. Loss of PDE-10A expression was not associated with tremor signs.

PET and SPECT studies with dopaminergic markers have shown correlations between dopaminergic decline, Parkinson’s progression and worse clinical severity (Otsuka *et al.*, 1996; Booij *et al.*, 1997; Broussolle *et al.*, 1999; Marek *et al.*, 2001; Hsiao *et al*., 2014). A previous study has reported correlations between striatal decreases in [18F]dopa uptake and longer disease duration and worse UPDRS-III motor scores at a 0.01 and 0.05 *α* level, respectively (Broussolle *et al.*, 1999). [18F]dopa and [18F]DTBZ PET studies have shown correlations between dopaminergic decline, worse bradykinesia and rigidity, but not tremor scores in groups of 17-27 Parkinson’s disease patients (Otsuka *et al.*, 1996; Broussolle *et al.*, 1999). In our study, decreased PDE-10A expression correlated with longer Parkinson’s disease duration and worse severity of bradykinesia and rigidity, at a similar level of statistical significance to that previously seen with dopaminergic imaging markers.

Although previous PET and SPECT studies have predominantly reported correlations between decreases in putaminal dopaminergic markers and Parkinson’s motor symptoms (Otsuka *et al.*, 1996; Broussolle *et al.*, 1999), here we found that also caudate PDE-10A decreases were associated with worse clinical scores. Our findings are consistent with a minority of previous dopaminergic PET and SPECT studies, which have shown correlations between lower caudate VMAT-2 and DAT levels and worse clinical scores (Marek *et al.,* 2001; Hsiao *et al.,* 2014), and therefore provide support for the role of caudate in the development of motor symptoms in Parkinson’s disease.

A striking feature of nigrostriatal dopaminergic degeneration in idiopathic Parkinson’s disease is the clinically asymmetric presentation. We assessed whether clinically asymmetric limb motor features correspond to PDE-10Adecreases, and we found no significant differences in PDE-10A expression between most and less affected sides in our cohort of Parkinson’s disease patients. It is possible that loss of PDE-10A expression does not follow the pattern of dopaminergic degeneration, however we speculate that this result could be due to the fact that the vast majority of the Parkinson’s patients had bilateral disease, all Parkinson’s patients were on levodopa therapy and had their PET scans while ON levodopa medication. A recent experimental study has demonstrated that levodopa treatment reduced PDE-10A levels in the unlesioned striatum of 6-OHDA lesioned animals (Sancesario *et al.*, 2014). In humans, would be very interesting to investigate the relationship between clinically asymmetric motor features and PDE-10A expression in early *de novo* Parkinson’s patients in a future study.

Our results also suggest that exogenous dopaminergic medication could have a degree of influence in striatal PDE-10A expression in patients with Parkinson’s disease. However, covariate analysis indicated that this influence was weak and it did not alter the results of our study. It is important to note that the vast majority of our cohort of Parkinson’s patients had established disease and significant motor symptoms with bilateral presentation. Experimental studies have shown that levodopa treatment balances PDE-10A levels by reducing PDE-10A expression in the unlesioned striatum, without significantly affecting the PDE-10A expression in the lesioned striatum of 6-OHDA animals (Sancesario *et al.*, 2014). We believe that chronic effect of levodopa could be investigated by studying in parallel a cohort of unilaterally affected *de novo* Parkinson’s patients, whereas the acute levodopa effect could be better understood if OFF and ON levodopa PDE-10A PET scans are performed and compared in the same Parkinson’s patients.

Our findings raise the possibility that, as with dopaminergic acting drugs such as levodopa and dopamine agonists, PDE-10A modulating drugs may have a therapeutic role in Parkinson’s disease. Similarly to previous studies with dopaminergic markers, PDE-10A loss did not correlate with tremor, which may be explicable by tremor pathophysiology being more closely related to serotonergic deficits (Loane *et al.*, 2013).

Axial signs such as imbalance, falls and freezing of gait are not associated with the degree of nigrostriatal dopaminergic denervation and they are often refractory to treatment with dopaminergic medication (Bohnen *et al.*, 2009). In our study, we found that worse axial signs were associated with loss of PDE-10A in globus pallidus. Deep brain stimulation of the globus pallidus has been shown to have variable effects on gait and balance problems in Parkinson’s disease (Krack *et al.*, 1998; Pötter-Nerger and Volkmann, 2013), and can even provoke gait freezing in patients with cervical dystonia (Berman *et al.*, 2009). Moreover, recent functional MRI studies have demonstrated that freezing of gait is associated with decreased BOLD signal in the globus pallidus during a virtual reality gait task (Shine *et al.*, 2013; Peterson *et al.*, 2014). We speculate that loss of PDE-10A in the globus pallidus may contribute to the development of axial signs in Parkinson’s disease.

Dopaminergic signalling is regulated by PDE-10A through the cAMP/PKA/DARPP-32 signalling cascade (Greengard *et al.*, 1999; Nishi *et al.*, 2008; Girault, 2012). PDE-10A has different roles in modulating cAMP signalling in the striato-nigral and striato-pallidial postsynaptic pathways. In striato-nigral neurons, dopamine loss leads to a reduced synthesis of D1 receptor-stimulated cAMP (Herve *et al.*, 2001); decreased PDE-10A expression will therefore increase cAMP levels and could in part compensate for the reduced cAMP signalling. In striato-pallidal neurons, dopamine loss decreases the inhibitory effect of D2 receptor on cAMP synthesis (Stoof and Kebabian, 1981).Decreased PDE-10A levels may further increase cAMP levels by enhancing the negative consequences of dopamine loss on D2 receptor signalling and potentiate adenosine A2A receptor signalling. Hence, loss of PDE-10A may result in functional imbalance between the striato-nigral and striato-pallidal dopaminergic pathways depressing motor activity and contributing to the development of Parkinson’s disease symptoms.

Half of our Parkinson’s disease patients had more advanced disease and were exhibiting LIDs. In Parkinson’s disease patients with LIDs, ROI-based analysis of PET signalling showed 19-34% reductions in PDE-10A expression in the caudate, substantia nigra and motor thalamic nuclei, compared to Parkinson’s disease patients with stable response to levodopa. These significant changes in the Parkinson’s disease LIDs subgroup were not confirmed at a voxel level and did not survive after using disease duration as a covariate in the analysis. These observations raise the possibility that further PDE-10A decreases in Parkinson’s patients with LIDs are the result of advanced disease rather directly related to the development of LIDs. However, when we assessed dyskinesia severity, rather than simply dyskinesia presence/absence, worse LIDs scores independently correlated with loss of striatal PDE-10A expression. It is unclear at this stage whether PDE-10A has a direct relevance to the development of LIDs in Parkinson’s disease, or whether this is simply an epiphenomenon. The prevalence of LIDs increases and PDE-10 expression decreases with advancing Parkinson’s disease, but a study comparing PDE-10A expression between Parkinson’s disease patients, matched for disease duration, with and without LIDs would be able to answer this question.

Preclinical studies have suggested that PDE-10A could be involved in the development of Parkinson’s disease LIDs. In dyskinetic Parkinson’s disease rats, striatal second messenger signalling was altered at the peak of dyskinesias due to the lost ability of striatal medium spiny neurons to induce both long-term potentiation and long-term depression (Picconi *et al.*, 2003; 2008; 2011). This means that stimulation of postsynaptic striatal neurons from levodopa-derived dopamine would fail and dysregulation of PDE-10A could be a pathogenic mechanism underlying the dysfunction of second messenger signalling. In dyskinetic rodent models of Parkinson’s disease, cAMP levels in the cortico-striatal-pallidal pathway were significantly reduced at peak of dyskinesias compared to non-dyskinetic animals, and administration of the non-selective PDE inhibitor, zaprinast, before levodopa effectively reduced the severity of dyskinesias and prevented the decreases in cyclic nucleotides levels (Giorgi *et al.*, 2008). A recent study has shown lower striatal cAMP and cGMP levels in dyskinetic animals at 60 minutes after levodopa administration (Sancesario *et al.*, 2014). During the phase of decreasing and extinction of dyskinesias (from 90 to 150 minutes after levodopa), cAMP/cGMP levels were up-regulated in dyskinetic animals. Thus, changes in cyclic nucleotide levels are linked to the appearance and regression of LIDs. Cyclic nucleotides levels are regulated by both dopamine-induced synthesis and PDE-related catabolism. It has been previously demonstrated that LIDs are associated with short-term sharp rises in synaptic dopamine levels following levodopa administration (de la Fuente-Fernandez *et al.*, 2001; Politis *et al.*, 2014). Hence, altered cyclic nucleotides levels due to abnormal dopamine-induced synthesis and PDE-10A-related catabolism might contribute to the development of LIDs.

Recent PET studies have demonstrated the importance of PDE-10A in other movement disorders such as Huntington’s disease (Ahmad *et al.*, 2014; Russell *et al.*, 2014). Striatal PDE-10A levels were decreased by 48-70% in manifest Huntington’s disease gene carriers (Ahmad *et al.*, 2014; Russell *et al.*, 2014). Similarly to what observed in our cohort of Parkinson’s disease patients, loss of striatal PDE-10A expression correlated with worse Unified Huntington’s Disease Rating Scale–Motor scores in manifest Huntington’s disease gene carriers (Russell *et al.*, 2014). Collectively, these observations suggest an extended role for the importance of PDE-10A expression in the clinical presentation of movement disorders.

In conclusion, our findings provide evidence for a novel neurochemical change in Parkinson’s disease, which is linked with the progression and severity of motor symptoms. [11C]IMA107 PET may be a valuable tool to understand the pathophysiology of Parkinson’s disease. PDE-10A could serve as a novel therapeutic target for manipulation with pharmacotherapy, and PDE-10A modulating drugs by acting independently or synergistically with levodopa could potentially have a therapeutic role in the alleviation of Parkinson’s disease symptoms and complications.

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**FIGURE LEGENDS**

**Fig. 1** AlteredPDE-10A expression in manually defined brain regions of Parkinson’s disease patients. **(A)** Mean [11C]IMA107 BPND parametric images derived from 12 healthy controls and 24 Parkinson’s disease patients in stereotaxic space overlaid onto the T1 weighted MNI template showing significant loss of striatal and pallidal PDE-10A signal in the Parkinson’s disease patients. **(B)** Column bar graph showing mean [11C]IMA107 BPND in anatomically defined brain regions between Parkinson’s disease patients and healthy controls. Colour bar reflects range of [11C]IMA107 BPND intensity. Error bars represent mean ± SD. \**P*<0.05; \*\**P*<0.01; \*\*\**P*<0.001.

**Fig. 2** Statistical parametric maps of [11C]IMA107 BPND (MNI co-ordinates x=17.99, y=3.59, z=7.42).Yellow-red areas represent voxel clusters with significant decreases in [11C]IMA107 BPND within the basal ganglia mask in Parkinson’s disease patients. The colour stripe indicates z-values.

**Fig. 3** Correlations between loss of PDE-10A and progression and severity of Parkinson’s disease. **(A)** Longer disease duration correlated with lower [11C]IMA107 BPND in caudate (*r*=−0.65; *P*=0.005), putamen (*r*=−0.51; *P*=0.025) and globus pallidus (r=−0.47; *P*=0.03) **(B)** Higher UPDRS-III scores correlated with lower [11C]IMA107 BPND ratio in caudate (*r*=−0.54; *P*=0.011), putamen (*r*=−0.48; *P*=0.022) and globus pallidus (*r*=−0.70; *P*<0.001) **(C)** Higher bradykinesia subscore was correlated with lower [11C]IMA107 BPND in caudate (*r*=−0.58; *P*=0.007), putamen (*r*=−0.53; *P*=0.010) and globus pallidus (*r*=−0.69; *P*<0.001) **(D)** Higher rigidity subscore was correlated with lower [11C]IMA107 BPND in caudate (*r*=−0.51; *P*=0.022), putamen (*r*=−0.48; *P*=0.022) and globus pallidus (*r*=−0.67; *P*<0.001).

**Fig. 4** Correlations between loss of PDE-10A and severity of levodopa-induced dyskinesias in 12 Parkinson’s disease patients with motor complications. Higher UDysRS scores correlated with lower [11C]IMA107 BPND in **(A)** caudate (*r*=−0.73, *P*=0.031) and **(B)** putamen (*r*=−0.74, *P*=0.031).

**TABLES**

**Table 1** Clinical characteristics of Parkinson’s disease patients.

| **Subject No** | **Sex** | **Age** | **Disease duration1** | **PD medication duration (yrs)2** | **Daily LED (mg)3** | **H&Y OFF4** | **H&Y ON** | **UPDRS-III OFF5** | **UPDRS-III ON** | **UPDRS-IV** | **UDysRS5** | **PDQ-396** |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| 1 | M | 69.6 | 12 | 11 | 1377 | 4 | 4 | 68 | 38 | 11 | 81 | 49 |
| 2 | F | 78.5 | 24 | 20 | 1329 | 4 | 4 | 71 | 54 | 17 | 109 | 30 |
| 3 | M | 66.1 | 14 | 13 | 2662.5 | 4 | 2 | 54 | 19 | 17 | 94 | 49 |
| 4 | F | 56.2 | 12 | 11 | 1715 | 2 | 2 | 46 | 16 | 1 | 98 | 33 |
| 5 | F | 59.0 | 13 | 12 | 973 | 4 | 3 | 61 | 11 | 11 | 55 | 49 |
| 6 | M | 59.9 | 9 | 8 | 520 | 2.5 | 2 | 29 | 9 | 5 | 86 | 16 |
| 7 | M | 56.8 | 12 | 10 | 2937 | 3 | 2 | 33 | 10 | 12 | 102 | 56 |
| 8 | F | 70.1 | 12 | 10 | 946 | 3 | 1.5 | 20 | 11 | 13 | 80 | 45 |
| 9 | M | 72.5 | 16 | 12 | 2873.75 | 4 | 3 | 64 | 13 | 17 | 114 | 35 |
| 10 | M | 69.4 | 9 | 8 | 1874.5 | 2 | 1 | 35 | 8 | 9 | 99 | 44 |
| 11 | F | 53.1 | 17 | 14 | 697 | 2 | 2 | 53 | 4 | 10 | 126 | 43 |
| 12 | M | 72.4 | 8 | 6 | 452 | 3 | 3 | 32 | 15 | 12 | 82 | 56 |
| 13 | M | 72.0 | 8 | 8 | 640 | 2 | 2 | 34 | 14 | 0 | 0 | 41 |
| 14 | M | 71.8 | 8 | 7 | 1515 | 2 | 2 | 48 | 36 | 0 | 0 | 36 |
| 15 | F | 67.4 | 5 | 0.9 | 400 | 1 | 1 | 10 | 2 | 0 | 0 | 22 |
| 16 | F | 71.3 | 5 | 3 | 850 | 2 | 1 | 35 | 6 | 0 | 0 | 45 |
| 17 | M | 75.5 | 4 | 3 | 300 | 2 | 1 | 21 | 5 | 0 | 0 | 7 |
| 18 | F | 57.2 | 1 | 10 | 150 | 1 | 0 | 6 | 0 | 0 | 0 | 7 |
| 19 | M | 72.9 | 11 | 9 | 960 | 2 | 1 | 39 | 15 | 0 | 0 | 7 |
| 20 | F | 71.3 | 5 | 5 | 510 | 1.5 | 1.5 | 11 | 6 | 0 | 0 | 62 |
| 21 | F | 59.8 | 7 | 6 | 480 | 1.5 | 1 | 27 | 6 | 0 | 0 | 10 |
| 22 | F | 62.2 | 4 | 3 | 920 | 2 | 1 | 30 | 11 | 0 | 0 | 72 |
| 23 | M | 70.5 | 8 | 5 | 490 | 2 | 1 | 19 | 8 | 0 | 0 | 6 |
| 24 | M | 75.5 | 8 | 6 | 1336.25 | 2 | 1 | 23 | 13 | 0 | 0 | 15 |
| **AVG****(± SD)** | **13M/****11F** | **67.13** (±**7.2**) | **9.67 (±5.0)** | **8.37 (±4.3)** | **1121.17 (±799.0)** | **2.44 (±1.0)** | **1.79 (±1.0)** | **36.21 (±18.5)** | **13.75 (±12.4)** | **5.63 (±6.6)\*** | **46.92 (±49.6)\*** | **34.79 (±19.5)** |

1From time of first appearance of PD motor symptoms; 2LED: Levodopa Equivalent Dose; 3H&Y: Hoehn and Yahr; 4UPDRS: Unified PD Rating Scale; 5UDysRS: Unified Dyskinesia Rating Scale; 6PDQ-39: 39-item Parkinson's Disease Questionnaire. \*AVG (±SD) of 12 PD patients with LIDs.

Table 2 [11C]IMA107 BPND in the groups of Parkinson’s disease patients and healthy controls.

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **ROIs1** | **Healthy Controls** | **PD patients** | ***P* value\*** | **% change in PD** |
| Striatum(mean±SD) | 1.81 (±0.28) | 1.40 (±0.23) | **< 0.001** | **−22.8%** |
| Caudate (mean±SD) | 1.37 (±0.25) | 0.98 (±0.18) | **< 0.001** | **−28.4%** |
| Putamen (mean±SD) | 2.21 (±0.31) | 1.64 (±0.29) | **< 0.001** | **−25.5%** |
| Ventral Striatum (mean±SD) | 1.16 (±0.24) | 1.14 (±0.22) | > 0.10 | −1.6% |
| Globus Pallidus (mean±SD) | 1.75 (±0.33) | 1.50 (±0.24) | **0.025** | **−14.3%** |
| Globus Pallidus external (mean±SD) | 1.91 (±0.39) | 1.72 (±0.25) | > 0.10 | −9.6% |
| Globus Pallidus internal (mean±SD) | 1.54 (±0.27) | 1.25 (±0.23) | **0.018** | **−18.8%** |
| Substantia Nigra (mean±SD) | 0.49 (±0.12) | 0.55 (±0.17) | > 0.10 | +12.4% |
| Motor thalamic nuclei (mean±SD) | 0.45 (±0.07) | 0.39 (±0.11) | > 0.10 | −12.3% |

1A-ROIs: anatomical regions of interest. \**P* values are Bonferroni corrected.

## Table 3 [11C]IMA107 BPND in the groups of Parkinson’s disease patients with levodopa-induced dyskinesias and stable response to levodopa.

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **ROIs1** | **PD stable (n=12)** | **PD LIDs (n=12)** | ***P* value\*** | **% change****in PD LIDs** |
| Striatum (mean±SD) | 1.48 (±0.24) | 1.31 (±0.18) | >0.10 | −11.7% |
| Caudate (mean±SD) | 1.08 (±0.19) | 0.88 (±0.10) | **0.045** | −**18.6%** |
| Putamen (mean±SD) | 1.74 (±0.31) | 1.55 (±0.23) | >0.10 | −10.9% |
| Ventral Striatum (mean±SD) | 1.23 (±0.21) | 1.04 (±0.19) | >0.10 | −15.4% |
| Globus Pallidus (mean±SD) | 1.60 (±0.24) | 1.40 (±0.21) | >0.10 | −12.4% |
| Globus Pallidus external (mean±SD) | 1.82 (±0.24) | 1.63 (±0.23) | >0.10 | −10.8% |
| Globus Pallidus internal (mean±SD) | 1.35 (±0.24) | 1.15 (±0.17) | >0.10 | −15.0% |
| Substantia Nigra (mean±SD) | 0.67 (±0.15) | 0.44 (±0.11) | **<0.001** | −**33.5%** |
| Motor thalamic nuclei (mean±SD) | 0.46 (±0.11) | 0.32 (±0.07) | **0.009** | −**29.3%** |

1A-ROIs: anatomical regions of interest. \**P* values are Bonferroni corrected but are not corrected for disease duration. Statistical significance was lost after using disease duration as a covariate in the analysis.

**Table 4** Voxel-based analysis: bilateral striatal significant decreases in [11C]IMA107 BPND in Parkinson’s patients compared with the group of healthy controls.

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **MNI coordinates** | **Area** | **Cluster sizes**  | **Z score** | ***P* value\*** |
| ***x*** | ***y*** | ***z*** |  |  |  |  |
| -14 | -4 | 22 | L Caudate | 139 | 4.53 | 0.002 |
| 14 | -4 | 22 | R Caudate | 130 | 4.01 | 0.004 |
| -28 | -8 | 12 | L Putamen | 519 | 4.28 | 0.006 |
| 28 | -8 | 12 | R Putamen | 420 | 3.83 | 0.028 |
| 22 | -6 | 2 | L Globus Pallidus | 74 | 2.90 | 0.031 |
| -22 | -6 | 2 | R Globus Pallidus | 46 | 2.71 | 0.038 |

\**P* values are FWE corrected except for globus pallidus where *P* values are uncorrected.